
Figures and figure supplements

The activity-dependent histone variant H2BE modulates the life span of olfactory neurons

Stephen W Santoro, Catherine Dulac

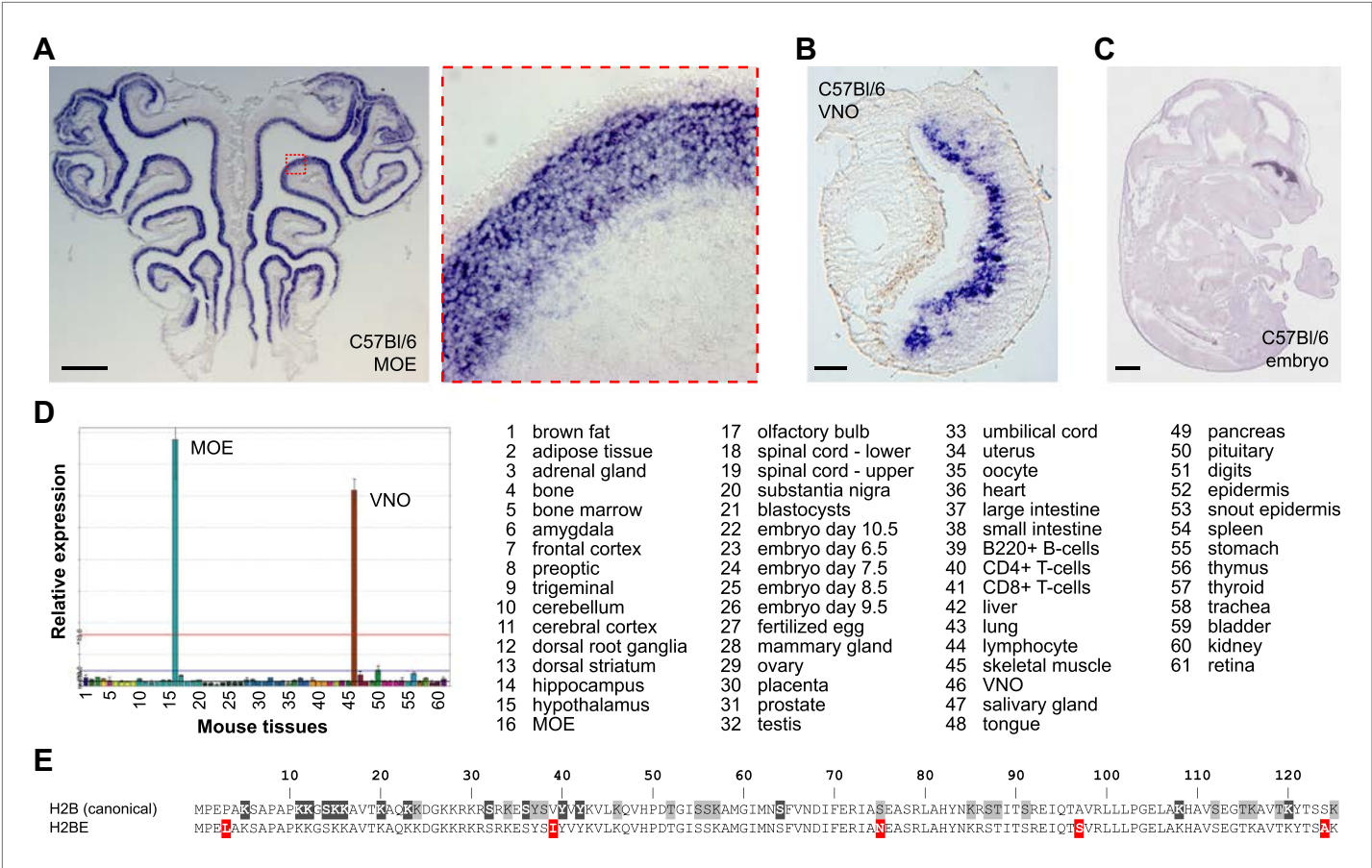


Figure 1. Mouse H2BE is detected exclusively in chemosensory neurons. **(A)** Analysis of *H2be* mRNA in the MOE of an 8-week old mouse, showing expression limited to sensory neurons. Boxed region is magnified (right). **(B)** Analysis of *H2be* mRNA in the VNO of an 8-week old mouse, showing expression limited to sensory neurons, especially in the apical zone. **(C)** Analysis of *H2be* mRNA in a sagittal section of an E14.5 mouse embryo (image from genepaint.org; ID: ES2590). **(D)** Profile of *H2be* mRNA levels in 61 mouse tissues (listed, right), showing exclusive expression in MOE and VNO. Data from GNF, now maintained by BioGPS (<http://biogps.org/>). **(E)** Alignment of H2BE and canonical H2B sequences. H2BE variant positions are highlighted in red; H2B PTM sites supported by >5 or ≤5 reports are highlighted in dark and light gray, respectively (<http://www.phosphosite.org>). Scale bars for (A), 500 μm; (B), 100 μm; (C), 1000 μm.

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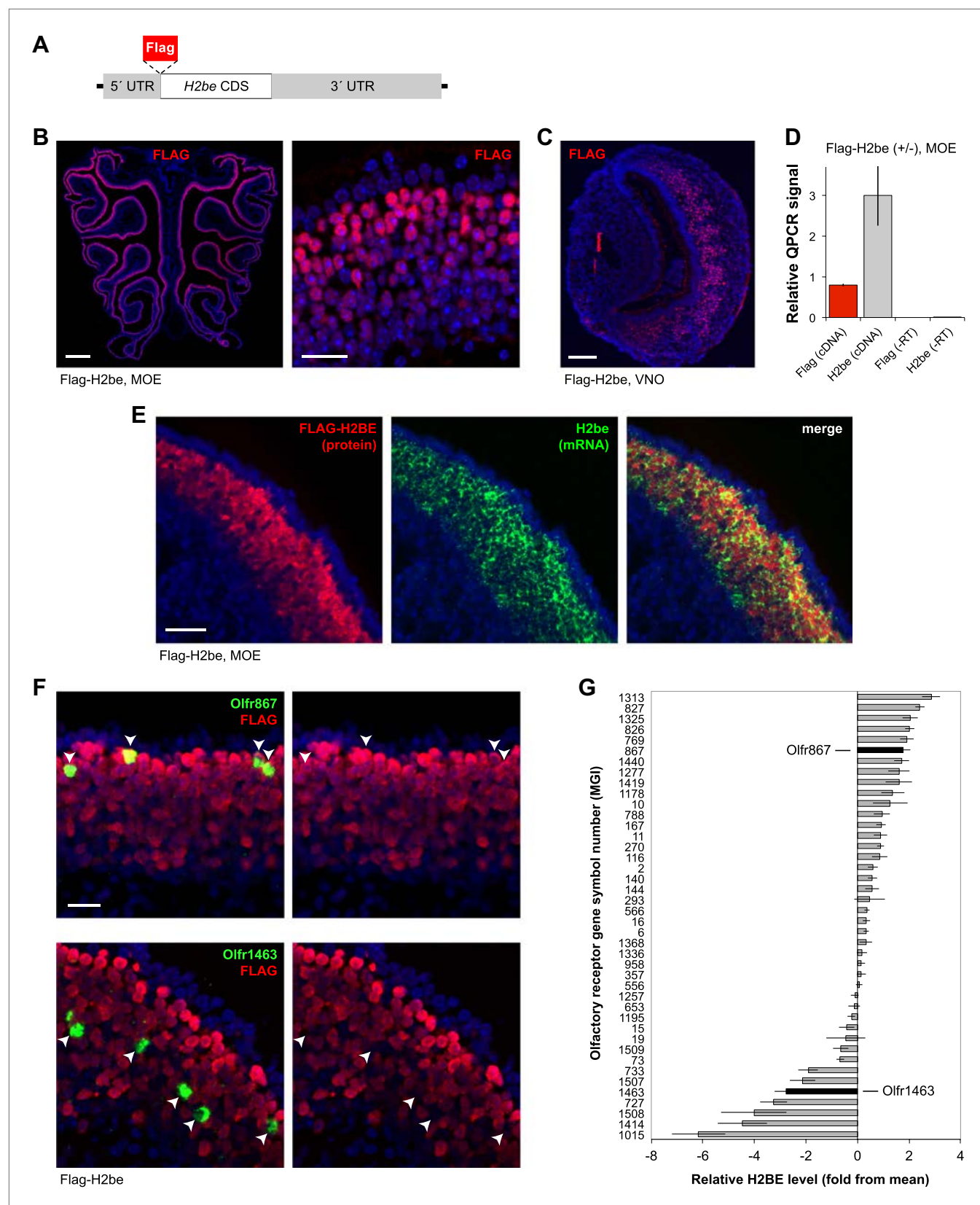


Figure 2. Generation of an *H2be:Flag-H2be* transgenic mouse (referred to as Flag-H2be) reveals that H2BE levels are stereotyped according to OR identity. (A) Flag-H2be transgenic construct, generated through modification of a BAC containing the *H2be* genomic region by insertion of a

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FLAG-encoding sequence immediately upstream of the *H2be* CDS. **(B,C)** Representative images of FLAG-H2BE in MOE (B) and VNO (C) from 10-week old Flag-H2be transgenic mice. **(D)** Quantitative PCR (qPCR) analysis of Flag-H2be transgene mRNA levels in whole MOE tissue from three-week old Flag-H2be^(+/-) transgenic mice. Signals were normalized to a value of three, corresponding to a primer pair recognizing all three *H2be* mRNAs (two endogenous and one transgenic). A primer pair specific for the Flag-tagged transgenic allele produces a normalized signal of approximately 1, indicating similar per-allele expression levels for the transgenic and endogenous *H2be* alleles. Negative control samples (-RT) were prepared by omitting reverse transcriptase during cDNA synthesis. **(E)** Colocalization analysis of FLAG-H2BE protein and *H2be* mRNA in the MOE of a 3-week old Flag-H2be^(+/-) transgenic mouse. Observation of occasional basally-located neurons that are *H2be*-mRNA-positive and FLAG-H2BE-negative is likely due to an expected lag in protein production and accumulation following *H2be* transcription onset during neuronal development. **(F)** Colocalization analysis of *Olf867* or *Olf1463* (arrowheads) and FLAG-H2BE, showing representative ORs associated with high or low levels of H2BE, respectively. Mouse age: 10 weeks. **(G)** Quantification of average H2BE levels in neurons expressing specific ORs (n = from 4 to 36 neurons per OR examined; mean, 18). Gene symbols are from the Mouse Genome Informatics database (MGI; <http://www.informatics.jax.org/>). Scale bars for (B, left), 500 μ m; (B, right) and (F), 20 μ m; (C) and (E), 100 μ m.

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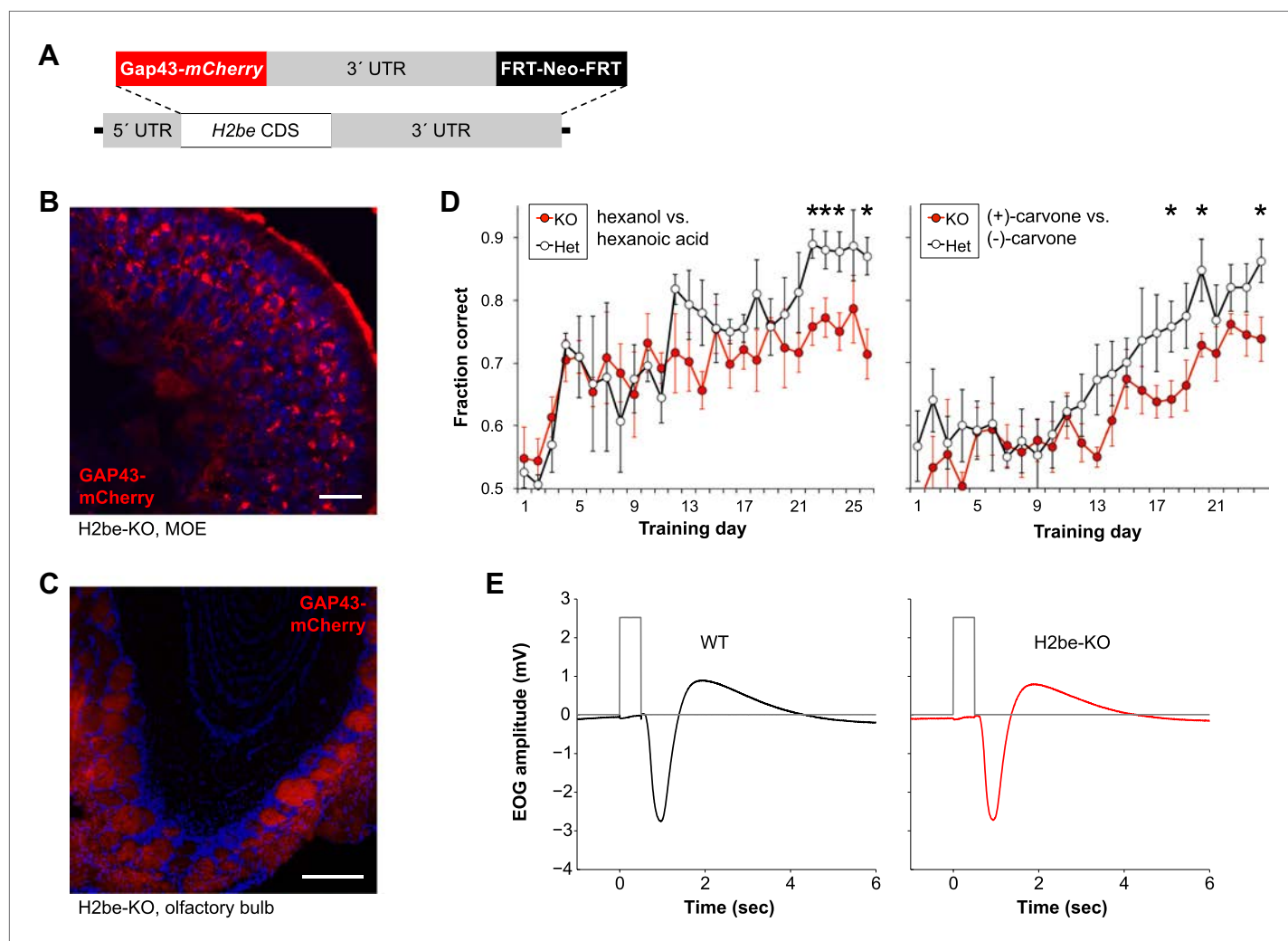


Figure 3. Generation of an H2be-KO/GAP43-mCherry-KI mouse line (referred to as H2be-KO) reveals that loss of *H2be* causes defects in olfaction. **(A)** H2be-KO allele, constructed through replacement of the endogenous *H2be* CDS with a membrane-targeted mCherry-encoding sequence (*Gap43-mCherry*). **(B,C)** Intrinsic GAP43-mCherry fluorescence in the MOE (B) and OB (C) of H2be-KO mice, showing GAP43-mCherry localization to the cell membranes and processes of olfactory neurons. Mouse ages: (B), 6 months; (C), 2 months. Scale bar for (B), 20 μ m; (C), 200 μ m. **(D)** Performance of approximately 3-month old water-restricted H2be-KO and control littermates in discriminating between hexanol/hexanoic acid (left) or (+)/(-)-carvone (right) odor pairs to obtain water ($n = 5$ per genotype). * $p < 0.05$. **(E)** Effects of *H2be* loss-of-function on odor-evoked electrical responses in the MOE. Electro-olfactogram traces (black and red) represent average responses to a 0.5-s stream of air from the head space of a 1% solution of isoamyl-acetate in mineral oil. Gray traces show timing of switching between the delivery of clean, de-odorized air (low), and odor-containing air (high). Results shown are representative of multiple trials, odorants, and concentrations; experimental procedures were adapted from those described previously (Waggener and Coppola, 2007).

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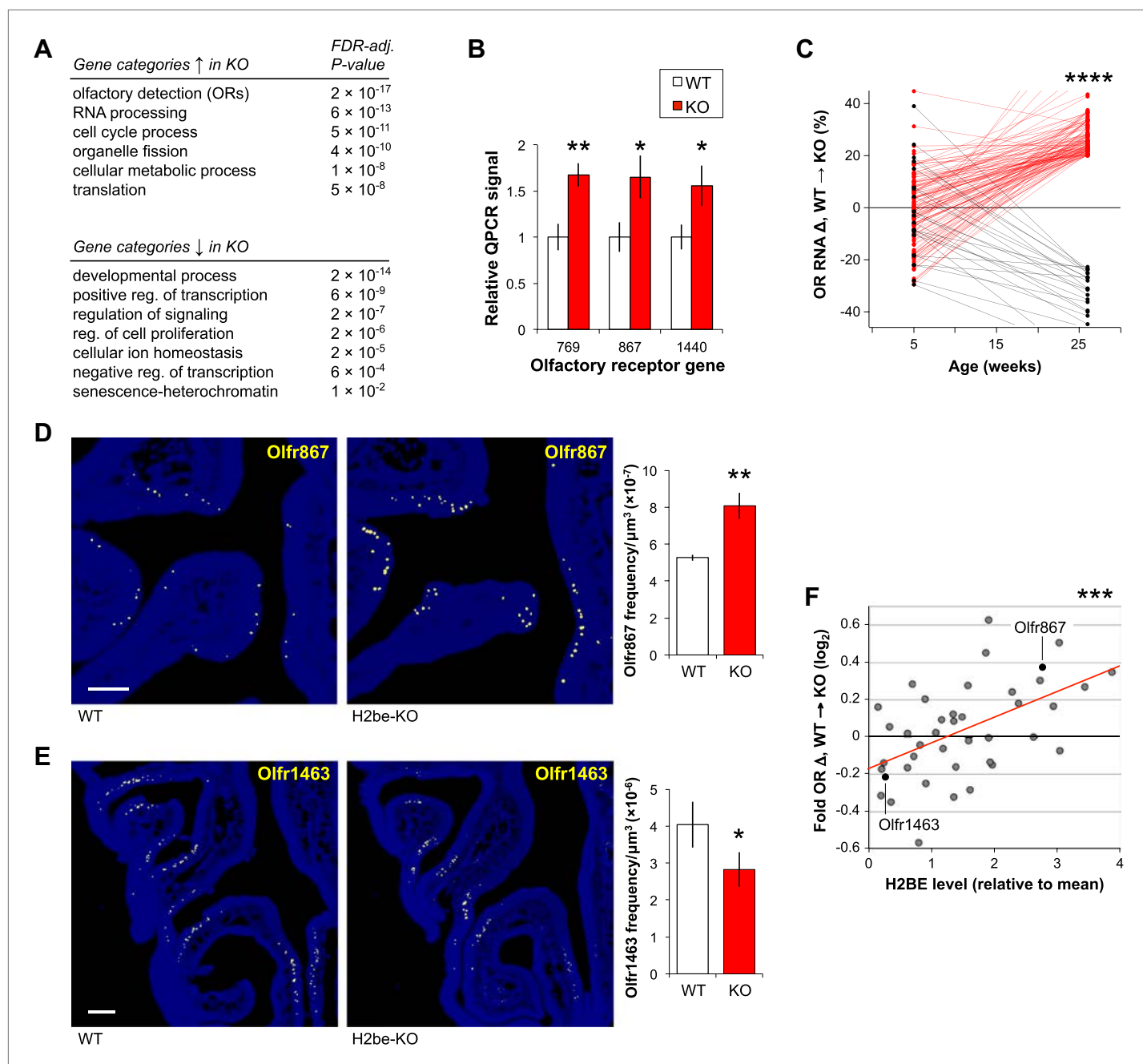


Figure 4. Loss of *H2be* causes defects in gene expression and OR expression frequencies. **(A)** Enriched gene ontology categories among genes up- (top) or down-regulated (bottom) in 6-month old *H2be*-KO vs WT MOEs, based on microarray analyses of whole MOE tissue ($n = 6$ samples/genotype, 2 animals/sample). **(B)** Multiplex qPCR analysis of OR mRNA levels in MOE tissue of six-month old *H2be*-KO and WT mice. Signals represent normalized ratios of specific OR mRNAs to *Cnga2*, which is unaltered in *H2be*-KO mice and used as an internal control ($n = 6$). **(C)** Expression differences in *H2be*-KO and WT MOEs (based on microarray analysis), plotted as a function of age. For simplicity, only ORs with differences >20% at age 6 months are shown, with up- and down-regulated ORs shown in red and black, respectively; statistical analysis included all ORs interrogated. **(D,E)** Representative images (left) and quantification (right) of *Olfr867* (D) and *Olfr1463* (E) expression frequencies in 6-month old *H2be*-KO and WT littermates ($n = 3$ mice, 10 sections per mouse). Scale bars, 200 μm. **(F)** Relationship between OR gene expression defects in 6-month old male *H2be*-KO mice (based on microarray analysis) and stereotypical *H2BE* levels as measured in male Flag-*H2be* mice. Red line, best fit. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 10^{-30}$.

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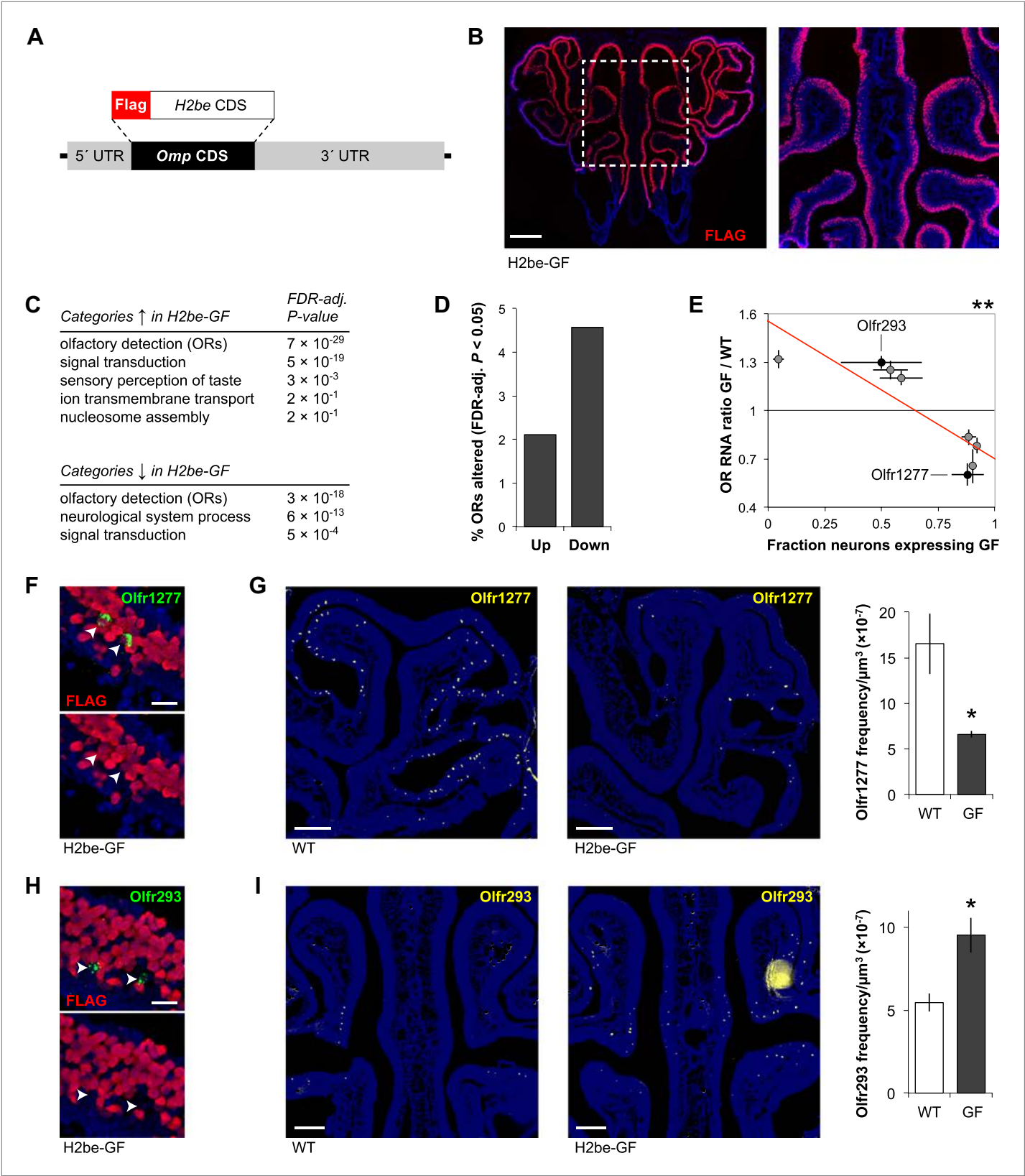


Figure 5. Generation of an *Omp:Flag-H2be* transgenic mouse (referred to as H2be-GF) reveals that ectopic overexpression of *H2be* in olfactory neurons alters gene expression and OR expression frequencies. (A) H2be-GF transgenic construct, generated through replacement of the *Omp* CDS with a

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FLAG-H2BE-encoding sequence in a vector containing the *Omp* genomic region. **(B)** Analysis of FLAG-H2BE in the MOE of a 5-week old H2be-GF mouse, showing high transgene expression in all mature neurons, except for a band near zone 2 (**Sullivan et al., 1996**). Boxed region is magnified (right). **(C)** Gene ontology categories enriched among genes up- (top) or down-regulated (bottom) in 5-week old H2be-GF vs WT MOEs, based on microarray analyses of whole MOE tissue ($n = 4$ samples, 3 animals per sample) and WT ($n = 6$ samples, 2 animals per sample) mice. **(D)** Percentage of OR genes with significantly differential expression (FDR-adjusted $p < 0.05$) in 5-week old H2be-GF and WT mice. **(E)** Relationship between OR gene expression defects in H2be-GF mice and transgene co-expression penetrance. Red line, best fit. **(F,H)** Colocalization of *Olf1277* or *Olf293* (arrowheads) and FLAG-H2BE in 5-week old H2be-GF mice, showing representative ORs associated with high- or low transgene penetrance, respectively. **(G,I)** Representative images (left) and quantification (right) of *Olf1277* (G) and *Olf293* (I) expression frequencies in 5-week old H2be-GF and WT littermates ($n = 3$ mice; 10 sections per mouse). * $p < 0.05$, ** $p < 0.01$. Scale bars for (B), 500 μm ; (F) and (H), 20 μm ; (G) and (I), 200 μm .

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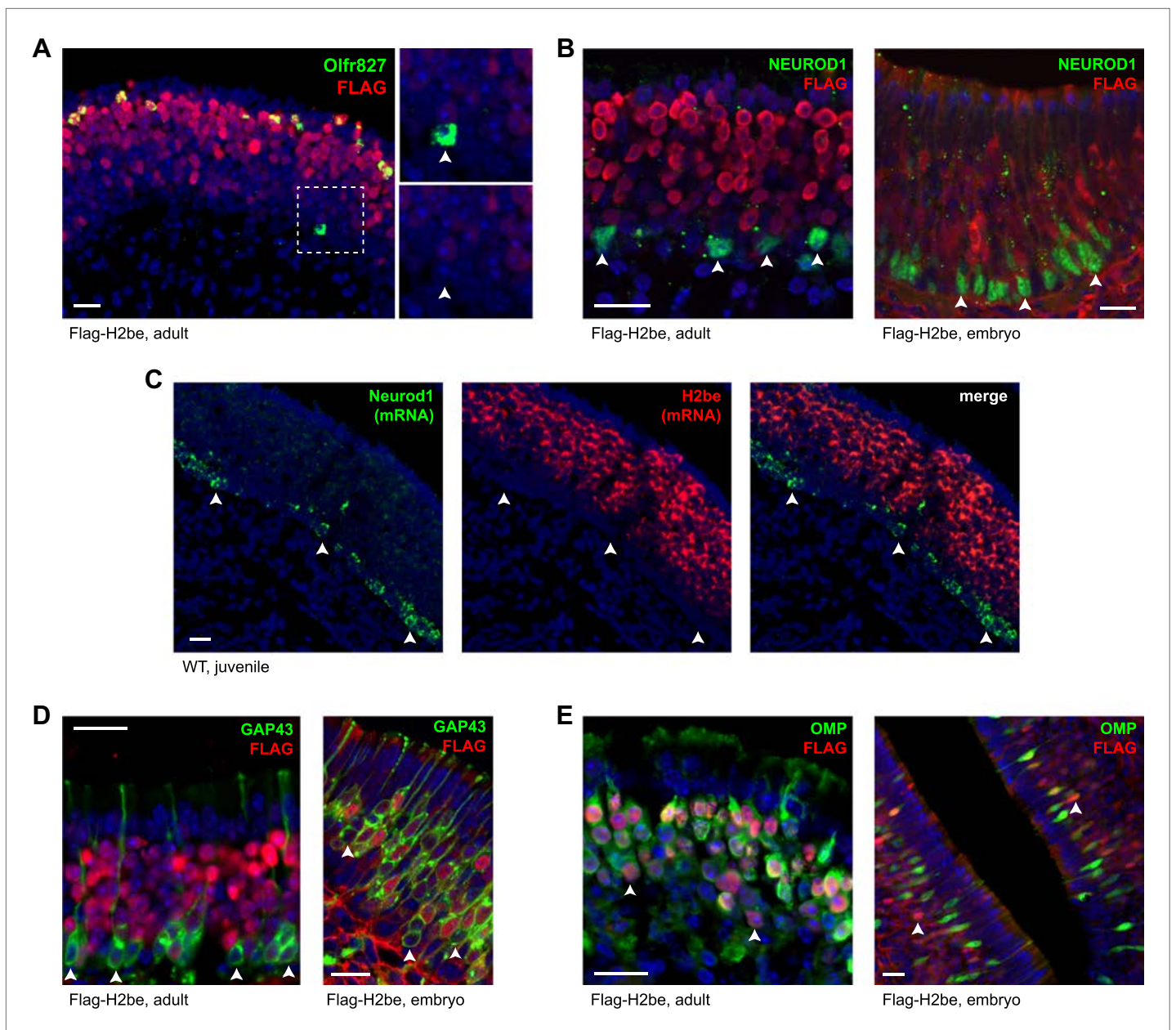


Figure 6. The expression onset of *H2be* follows OR choice during neuronal development. **(A)** H2BE is undetectable in an *Olfr827*⁺ neuron that is newly-differentiated (note its basal position in the epithelium; boxed and magnified, right), but expressed at uniformly high levels in mature *Olfr827*⁺ neurons. **(B,D,E)** MOE expression patterns in adult (left) or embryo (right) of *H2be* relative to *Neurod1*, *Gap43*, and *Omp*, which have expression onsets prior to, concurrently with, and following OR choice, respectively. All NEUROD1⁺ cells (B) and a fraction of basal GAP43⁺ (newly-differentiated) neurons (C) are H2BE⁻ (arrowheads), whereas a fraction of H2BE⁺ neurons are OMP⁻ (E; arrowheads). **(C)** Colocalization of endogenous *H2be* and *Neurod1* mRNAs in the MOE of a 3-week old WT mouse. Mouse ages: (A), 3 months; (B, left), 4 months; (C), 3 weeks; (D, left) and (E, left), 10 months. Scale bars for (A) to (E), 20 μ m.

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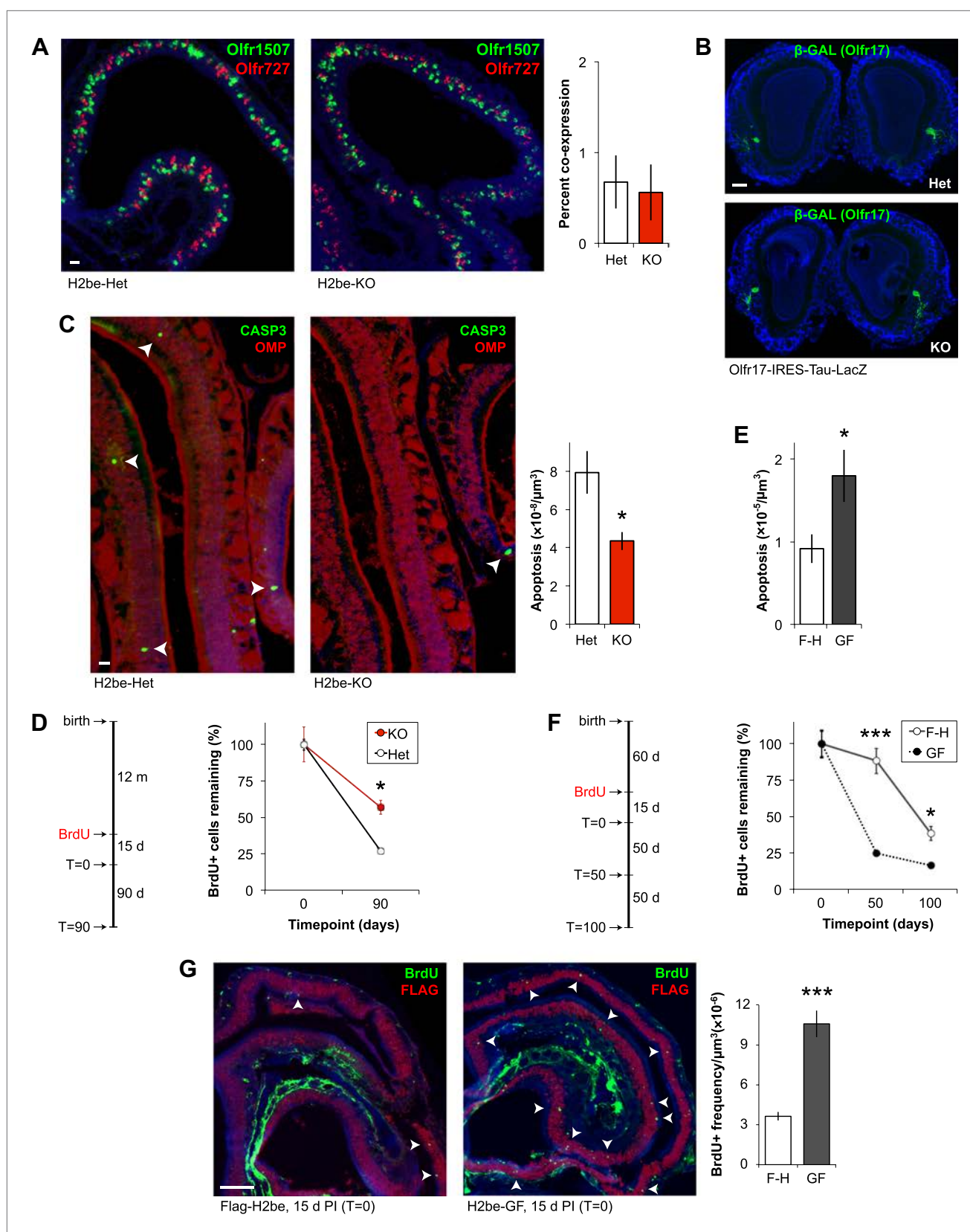


Figure 7. H2BE affects olfactory neuronal longevity, not OR choice. (A) Representative images (left) and quantification (right) of *Olfr1507* and *Olfr727* co-expression frequencies in H2be-KO and control littermates ($n = 10$ sections). (B) Labeled axons of *Olfr17*-expressing neurons form indistinguishable Figure 7. Continued on next page

Figure 7. Continued

glomeruli in the OBs of H2be-KO and control littermates. **(C,E)** Representative images (C, left; arrowheads) and quantification (C, right; E) of apoptosis in mature neurons of H2be-KO (C) or H2be-GF (E) mice compared to controls (C: $n = 3$ mice, 12 sections per mouse; E: $n = 10$ sections). **(D,F)** Schematic of experimental analysis timeline (left) and quantification (right) of relative BrdU⁺ neuron frequencies in H2be-KO (D) and H2be-GF (F) mice compared to controls, respectively (D: $n = 3$ mice per timepoint, 12 sections per mouse; F: $n = 10$ sections per timepoint). F-H: Flag-H2be. **(G)** Representative images (left) and quantification (right) of neurogenesis in H2be-GF (GF) mice and Flag-H2be (F-H) controls by analysis of frequencies of BrdU⁺ neurons (arrowheads) 15 days post-injection of BrdU ($T = 0$ timepoint; $n = 10$ sections). * $p < 0.05$, *** $p < 0.001$. Mouse ages: (A) and (E), 4 months; (B) and (G), 2 months; (C), 15 months. Scale bars for (A) and (C), 20 μm ; (B), 200 μm ; (G), 100 μm .

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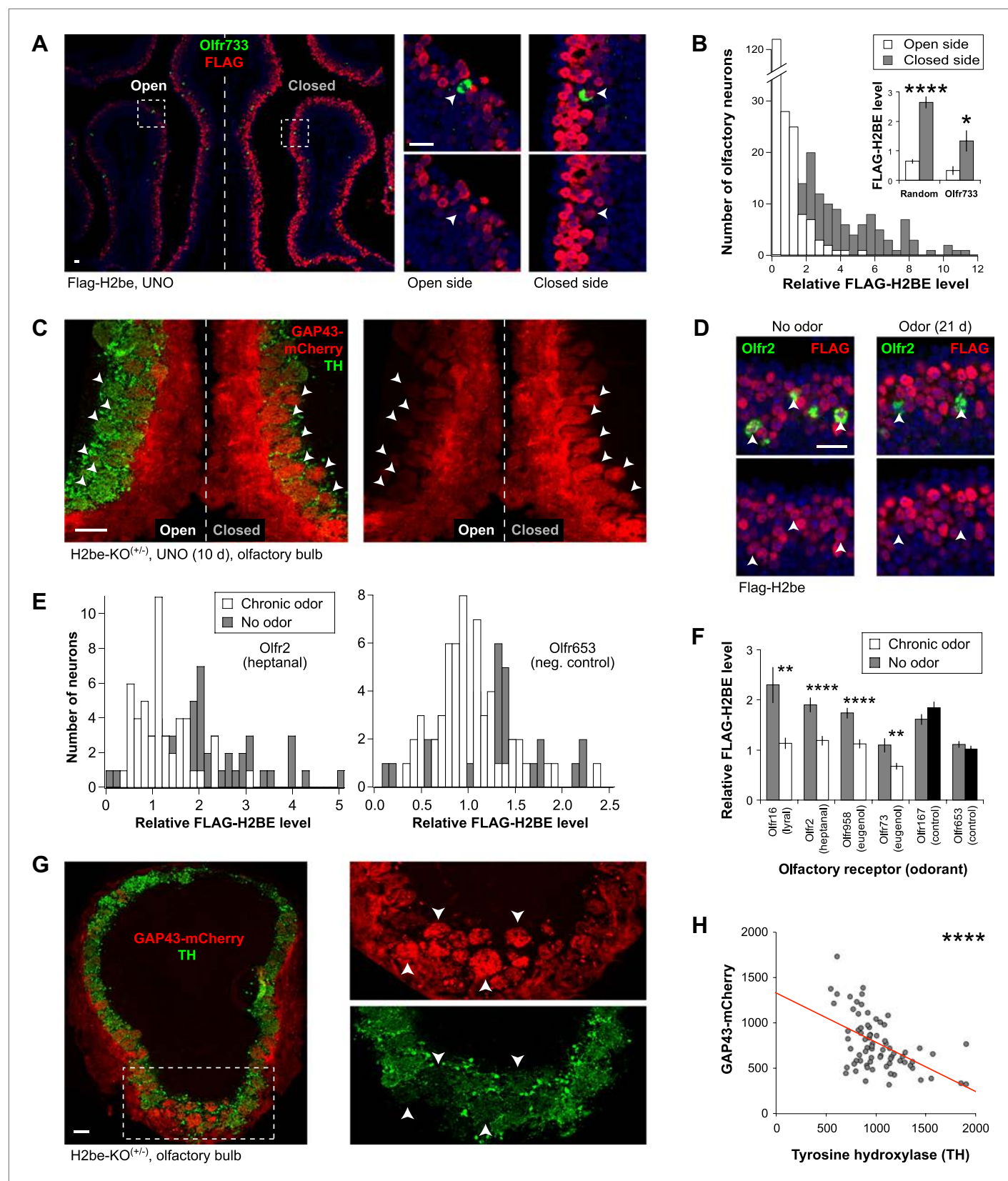


Figure 8. H2be is regulated by activity. (A,B) Effects of unilateral naris occlusion (UNO) on H2BE level in the MOE. (A) Representative images of FLAG-H2BE and *Olf733* colocalization (boxed regions magnified, right; *Olf733*⁺ neurons, arrowheads). (B) Distributions (main) and averages (inset) of Figure 8. Continued on next page

Figure 8. Continued

H2BE level within nuclei of randomly-sampled (main; inset, left; $n = 200$) or *Olfr733*⁺ (inset, right; $n = 10$) neurons on the two sides of the MOE. (C) Effects of UNO on intrinsic GAP43-mCherry fluorescence in glomeruli (arrowheads) within the OB of an H2be-KO heterozygous mouse. Reduced tyrosine hydroxylase (TH; a marker of olfactory activity) staining on the closed side indicates completeness of naris closure. (D–F) Effects of odor exposure on H2BE level in olfactory neurons. Representative images (D) and quantification (E, left) of FLAG-H2BE in *Olfr2*⁺ neurons (D, arrowheads) exposed to odors or mineral oil (no odor). (E, right) Quantification of FLAG-H2BE in *Olfr653*⁺ neurons (chosen randomly as a negative control) exposed to odors or mineral oil (no odor). (F) Average H2BE level within neurons expressing odor-stimulated or control ORs ($n = 20$ –60). (G,H) Representative image (G; boxed region magnified, right) and quantification (H) of the relationship between GAP43-mCherry and tyrosine hydroxylase intensities within glomeruli of an H2be-KO heterozygous mouse. Red line, best fit. * $p < 0.05$; ** $p < 0.01$; **** $p < 0.0001$. Mouse ages: (A) and (C), 4 weeks; (D), 8 weeks; (G), 10 weeks. Scale bars for (A) and (D), 20 μm ; (C) and (G), 100 μm .

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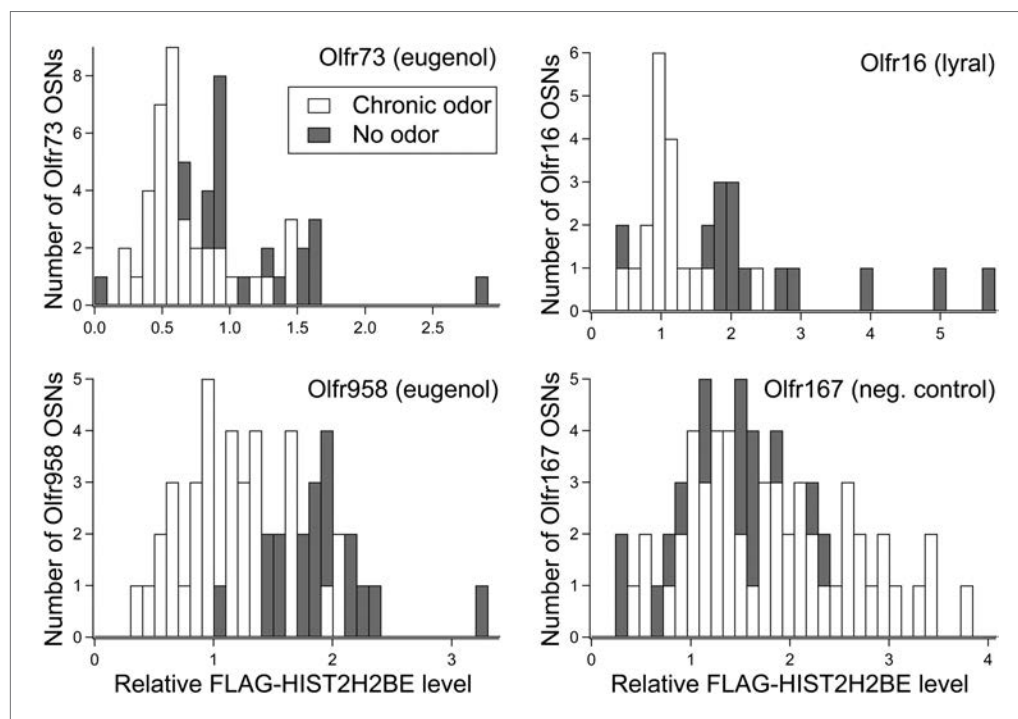


Figure 8—figure supplement 1. Distribution of relative H2BE levels within neurons expressing *Olfr73*, *Olfr958*, *Olfr16*, and *Olfr167* for odor-exposed and control littermates. Stimulating odors are indicated, except for *Olfr167*, which serves as a negative control.

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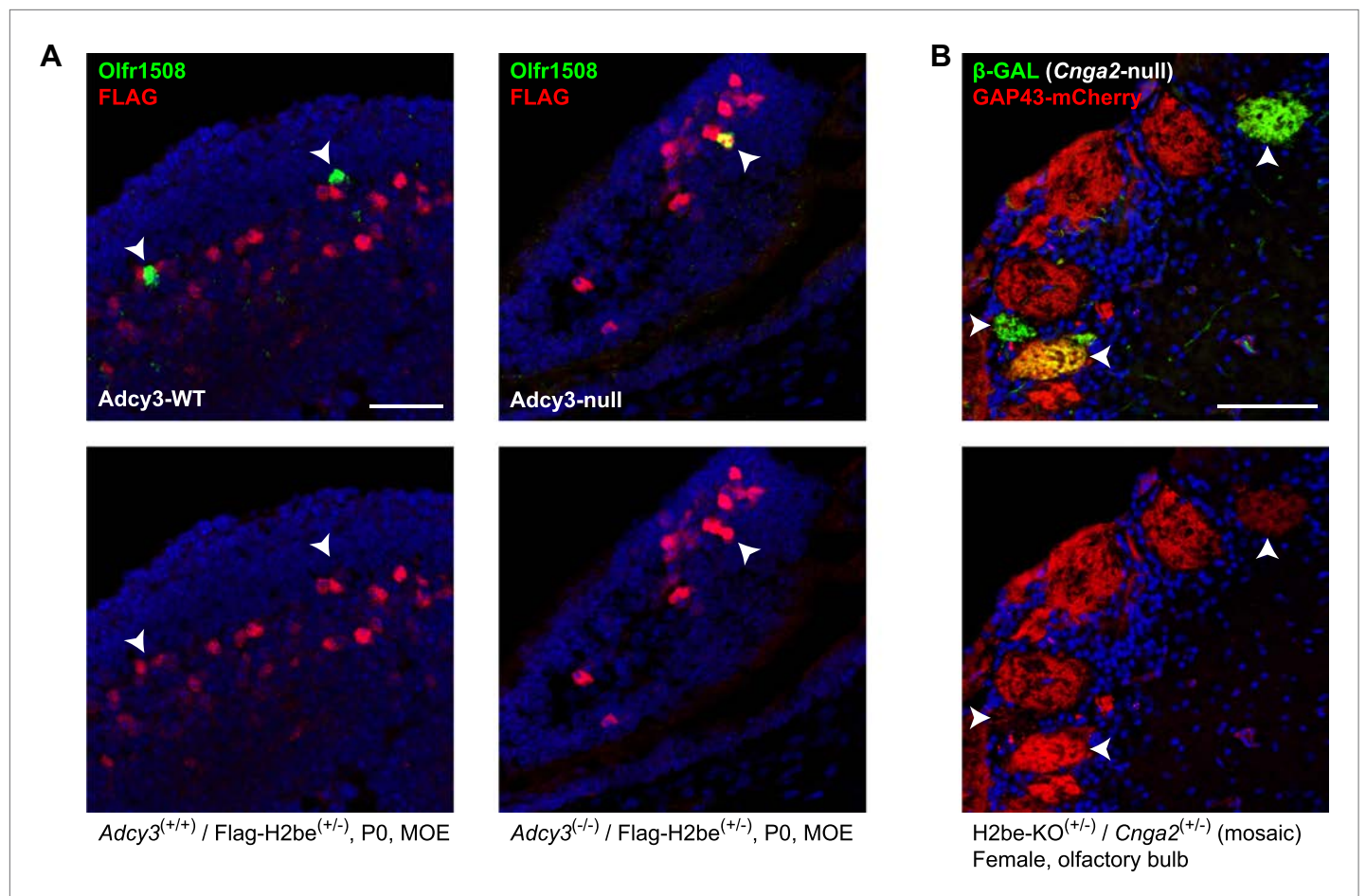


Figure 9. H2BE levels are cAMP-, but not Ca^{2+} -dependent. **(A)** Representative example of the effects of *Adcy3* loss-of-function on H2BE levels in newborn (P0) mice. In *Adcy3*^(+/+) mice, *Olfr1508*⁺ neurons contain extremely low H2BE levels (left, arrowheads), while in *Adcy3*^(-/-) mice, *Olfr1508*⁺ neurons frequently contain extremely high levels (right, arrowhead), indicating that cAMP participates in the negative regulation of H2BE. Note: age P0 was chosen due to the low postnatal survival rate of *Adcy3*^(-/-) mice. **(B)** Effects of *Cnga2* loss-of-function on *H2be* expression. Mice in which *Cnga2* (an X-chromosomal gene necessary for odor-evoked Ca^{2+} signaling) was replaced with the *Tau-LacZ* gene (Zhao and Reed, 2001) were crossed with *H2be*-KO mice to generate three-week old *Cnga2*-KO-*Tau-LacZ*^(+/+)/*H2be*-KO-*Gap43-mCherry*^(+/+) compound heterozygous females. In these mice, one half of new olfactory neurons express TAU-LACZ instead of *CNGA2* and project to glomeruli distinct from neurons expressing *Cnga2* (Zheng et al., 2000). Analysis of β -GAL⁺ (TAU-LACZ) and GAP43-mCherry intensities within glomeruli revealed that *Cnga2*⁻ neurons (β -GAL⁺; arrowheads) do not have higher levels of GAP43-mCherry than *Cnga2*⁺ neurons (β -GAL⁻), indicating that *H2be* expression is not negatively regulated by Ca^{2+} signaling. Scale bars for (A), 40 μm ; (B), 100 μm .

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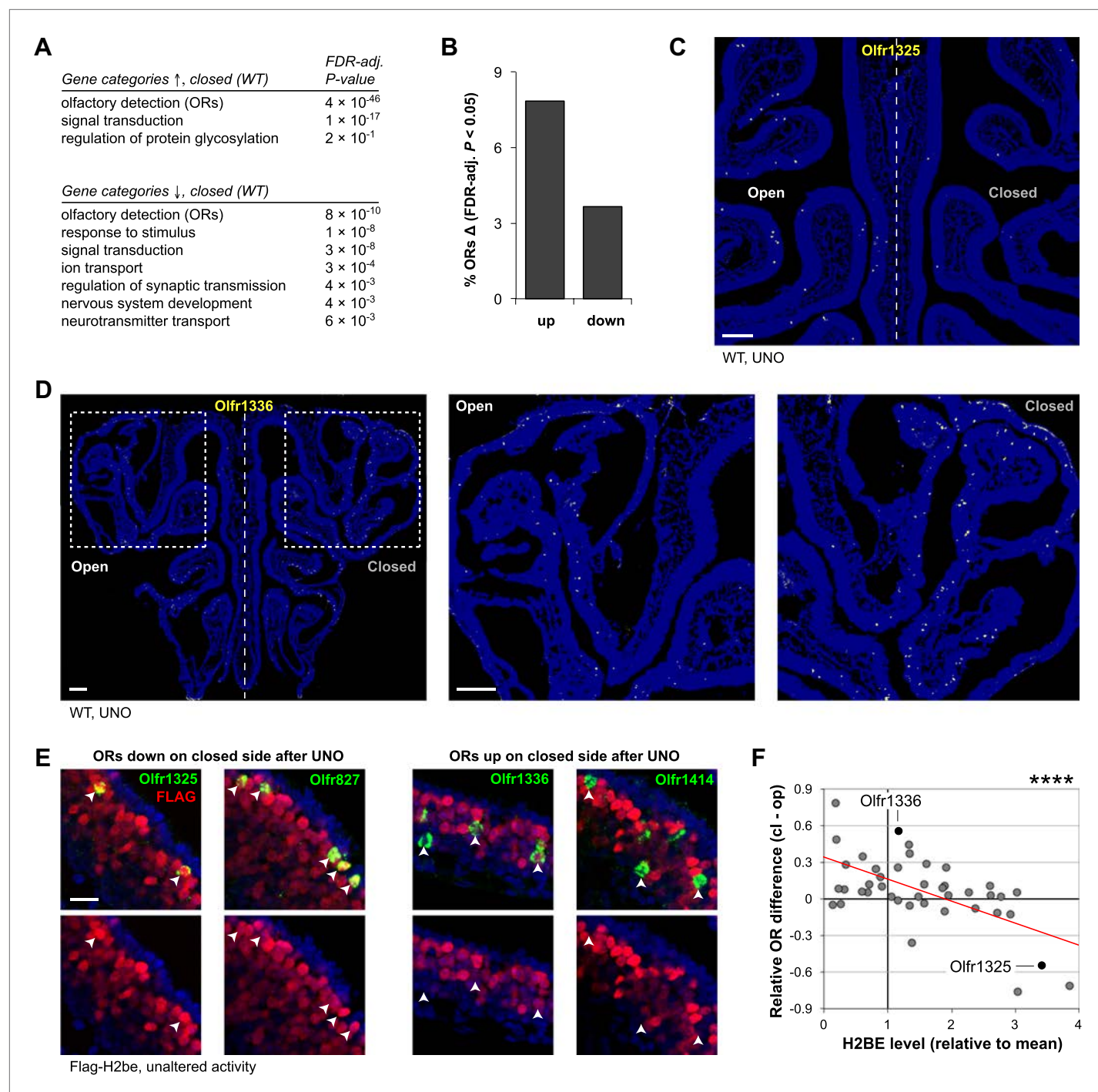


Figure 10. Unilateral naris occlusion (UNO) alters gene expression and OR expression frequencies. **(A)** Gene ontology (biological process) terms enriched at the top of a gene list ranked descendingly according to differential expression on the two MOE halves from 5-week old WT mice subjected to UNO (21 days), based on microarray analysis ($n = 3$ samples per MOE side, four animals per sample). **(B)** Percentage of OR genes with significantly differential (FDR-adjusted $p < 0.05$) expression on the two sides of the MOE of WT mice after UNO (21 days; values from microarray data). **(C,D)** Representative images of *Olfr1325* (C) and *Olfr1336* (D) expression in the MOE after UNO. **(E)** Representative images of normal FLAG-H2BE levels in neurons associated with ORs that are down- (left) or up-regulated (right) in frequency after olfactory deprivation. **(F)** Relationship between UNO-mediated OR gene expression differences on the two sides of the MOE (values from microarray data for WT mice subjected to UNO for 21 days) and associated FLAG-H2BE levels measured in intact mice. Red line, best fit, **** $p < 0.0001$. Mouse ages: (C) and (D), 5 weeks; (E), 12 weeks. Scale bars for (C) and (D), 200 μm ; (E), 20 μm .

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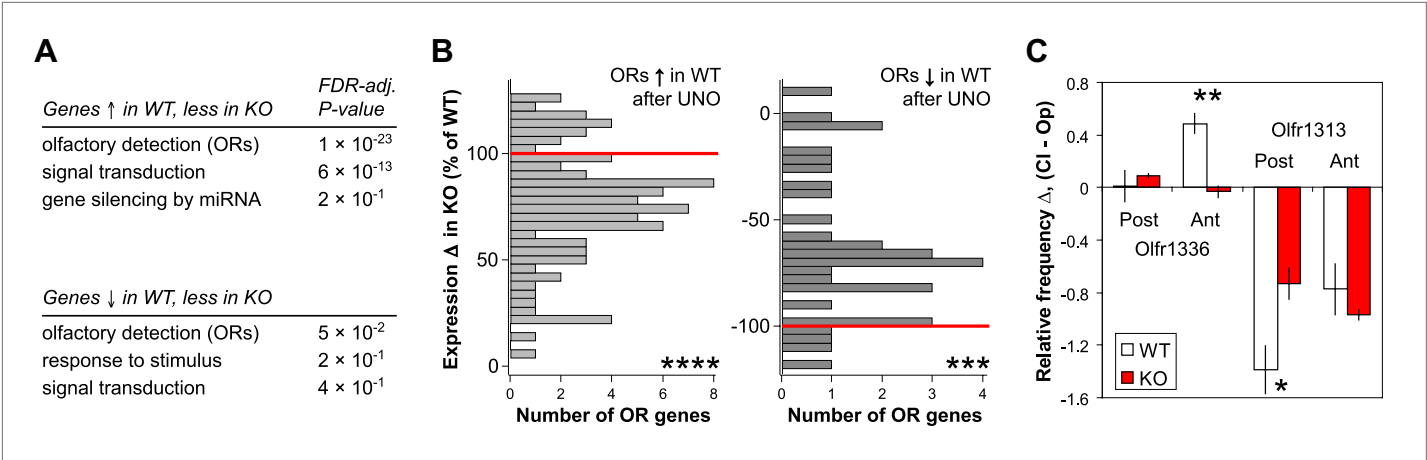


Figure 11. *H2be* affects activity-dependent gene expression. **(A)** Gene ontology (biological process) terms enriched among genes with UNO-mediated expression differences in WT mice (\log_2 fold-change > 0.3; unadjusted $p < 0.02$), but at least 20% less altered expression in *H2be*-KO compared to WT mice after UNO, based on microarray analysis of MOE halves from 5-week old WT and *H2be*-KO mice subjected to UNO (21 days; $n = 3$ samples per MOE side, four animals per sample). **(B)** Histograms of UNO-mediated OR expression differences on the closed and open sides of the MOE of *H2be*-KO mice as a percentage of the corresponding WT differences (normalized to 100% or -100%; red lines) for ORs significantly up- (left) or down-regulated (right) in WT mice after olfactory deprivation (FDR-adjusted $p < 0.05$; values from microarray data). **(C)** Comparison of UNO-altered *Olf1336* and *Olf1313* frequencies in 5-week old WT and *H2be*-KO mice subjected to UNO (21 days). Values correspond to relative OR expression frequency differences on the two sides of the MOE according to the anterior (Ant)/ posterior (Post) position ($n = 3$ mice, five sections per region per mouse). Note: UNO appears to affect OR frequencies differently in the anterior and posterior regions of the MOE. * $p < 0.05$; ** $p < 0.01$ *** $p < 0.001$; **** $p < 0.0001$.

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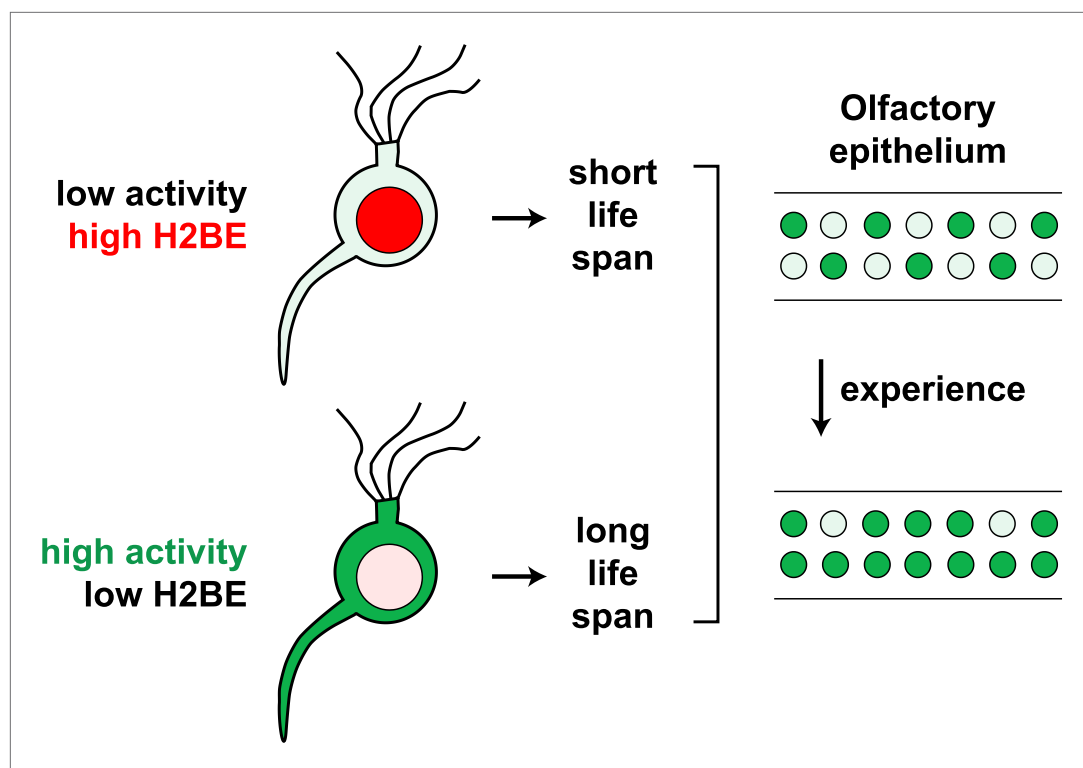


Figure 12. Model for the effects of neuronal activity on H2BE expression level, life span and resulting neuronal representation.

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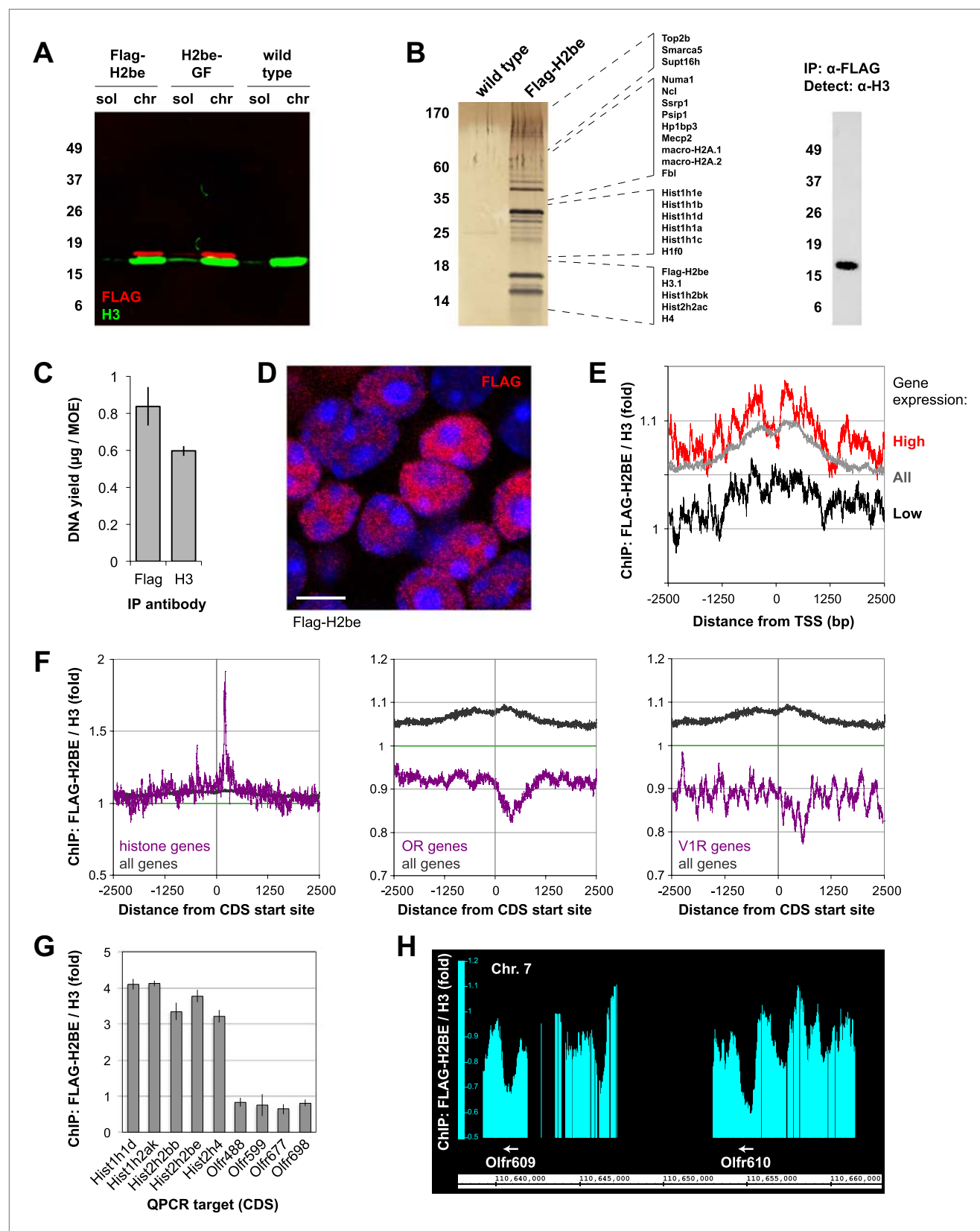


Figure 13. Chromatin incorporation and localization of FLAG-H2BE. **(A)** Two-color western analysis of FLAG-H2BE and H3 in soluble nucleoplasm (sol) and chromatin (chr) fractions of unfixed MOE cell nuclei from 16-week old Flag-H2be, H2be-GF, and WT mice. **(B)** SDS-PAGE analysis (left), mass spectrometry (right) of Flag-H2be. **(C)** Bar graph showing DNA yield (μg / MOE) for Flag and H3. **(D)** Fluorescence image of Flag-H2be. **(E)** ChIP-seq plot of FLAG-H2BE / H3 (fold) vs distance from TSS (bp). **(F)** ChIP-seq plots of FLAG-H2BE / H3 (fold) vs distance from CDS start site for histone genes, OR genes, and V1R genes. **(G)** Bar graph showing ChIP-seq of FLAG-H2BE / H3 (fold) vs QPCR target (CDS). **(H)** ChIP-seq plot of FLAG-H2BE / H3 (fold) vs distance from CDS start site for Chr. 7.

Figure 13. Continued

spectrometric identification (listed, middle), and western analysis (right) of proteins associated with immunoprecipitated FLAG-H2BE-containing native mononucleosomes from MOE tissue of Flag-H2be transgenic mice. Proteins identified by mass spectrometry are listed according to their approximate electrophoretic mobility. **(C)** Quantification of DNA immunoprecipitated from crosslinked and fragmented chromatin derived from MOE tissue of Flag-H2be transgenic mice. **(D)** Representative image of FLAG-H2BE localization within olfactory neurons of a 10-week old Flag-H2be mouse. Scale bar, 5 μ m. **(E)** Genome-wide ChIP analysis of relative FLAG-H2BE levels with respect to distance from the transcript start sites (TSS) for all mouse genes (grey), and genes expressed at high (red) and low (black) levels in olfactory neurons. **(F)** Genome-wide ChIP analysis of relative FLAG-H2BE levels with respect to distance from the CDS start sites for mouse histone (left), OR (middle), and vomeronasal type 1 receptor (V1R; right) genes in comparison to all genes. **(G)** Quantitative PCR analysis of relative FLAG-H2BE levels in the protein-coding regions of representative histone and OR genes. Analyses were performed on ligation-mediated-PCR-amplified Flag-H2BE and H3 ChIP DNA samples. **(H)** Genome-wide ChIP analysis shows depleted FLAG-H2BE levels within representative OR CDS regions.

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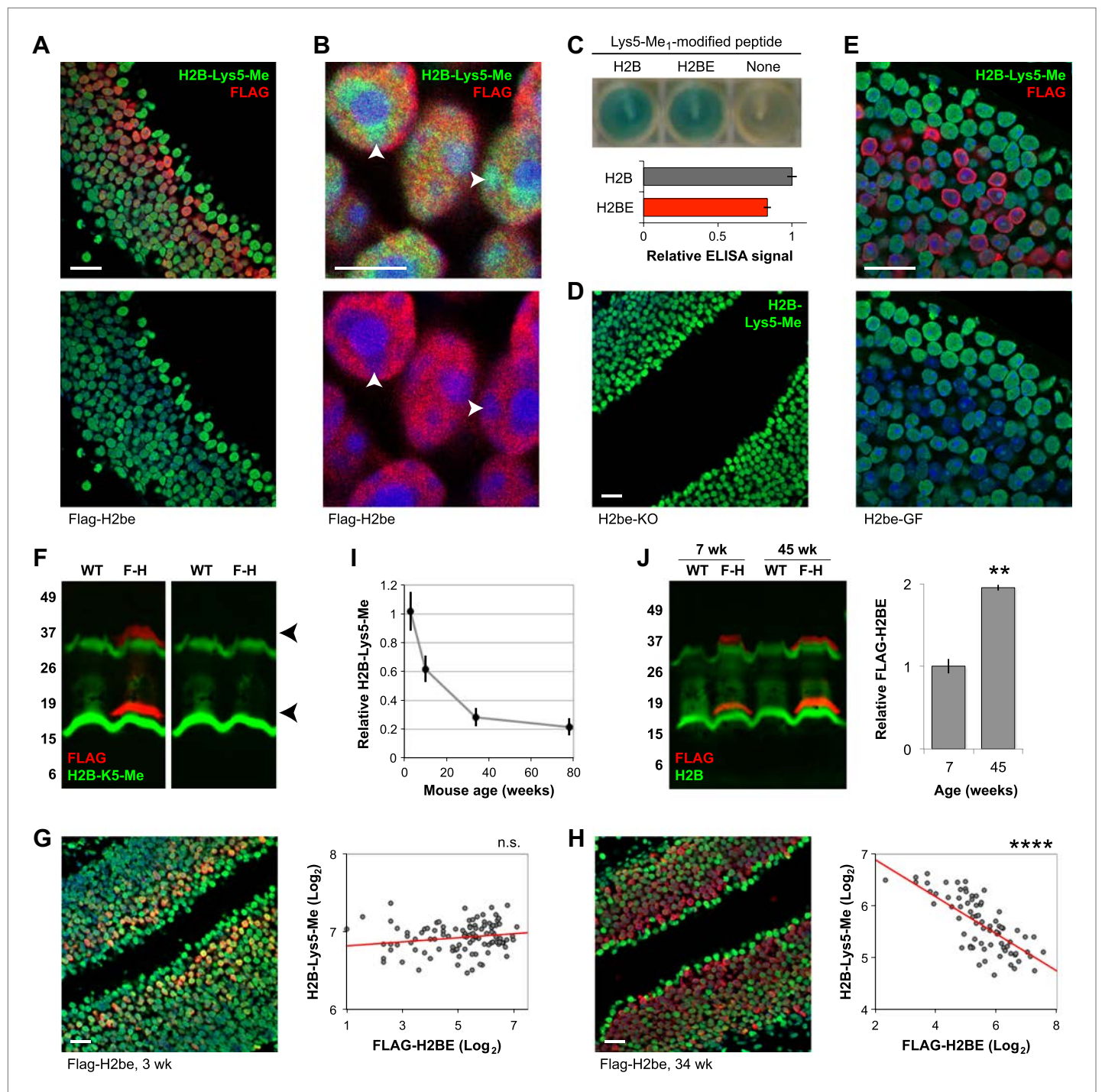


Figure 14. H2BE's post-translational modifications (PTMs) differ from those of canonical H2B. (A,B,D,E) Representative images of H2B-Lys5-Me (A, B, D, and E) and FLAG-H2BE (A, B, and E) staining in the MOE of Flag-H2be (A and B), H2be-KO (D) or H2be-GF (E) mice. (B) High-magnification image of FLAG-H2BE and H2B-Lys5-Me colocalization shows that H2BE is depleted in nuclear regions enriched for H2B-Lys5-Me (arrowheads). Mouse ages: (A), (B), and (E), 10 weeks; (D), 34 weeks. (C) Confirmation of reactivity of the anti-H2B-Lys5-Me₁ polyclonal antibody with the Lys5-Me PTM in the context of the H2BE protein sequence. Image (top) and quantification (bottom) of an ELISA assay for peptides corresponding to canonical H2B or H2BE and containing the Lys5-Me PTM. (F) Two-color fluorescent western analysis of Lys5-Me modification of FLAG-H2BE in MOE lysates from WT and Flag-H2be (F-H) mice. No detectable H2B-Lys5-Me staining of the FLAG-H2BE bands (red; arrowheads) is observed. Approximate molecular weights (kDa) are indicated (left). The bands observed at approximately 30–35 kDa likely correspond to histone dimers. (G–I) Age-dependence of H2BE accumulation. (G and H) Images (left) and quantification (right) of FLAG-H2BE and H2B-Lys5-Me co-localization in 3- (G) and 34-week old (H) mice. Red lines, best fits. (I) Figure 14. Continued on next page

Figure 14. Continued

Quantification of H2B-Lys5-Me levels in high-H2BE neurons (relative to apical sustentacular cells; $n = 20$ nuclei from two images per timepoint). (J) Two-color fluorescent western analysis (left) and quantification (right) of FLAG-H2BE relative to total H2B as a function of age in MOE lysates from WT and Flag-H2be (F-H) mice. Approximate molecular weights (kDa) are indicated (left). The bands observed at approximately 30–35 kDa likely correspond to histone dimers. ** $p < 0.01$; **** $p < 0.0001$; n.s., not significant. Scale bar for (A), (D), (E), (G), and (H), 20 μm ; (B), 5 μm .

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