

Figures and figure supplements

The transcriptional regulator BZR1 mediates trade-off between plant innate immunity and growth

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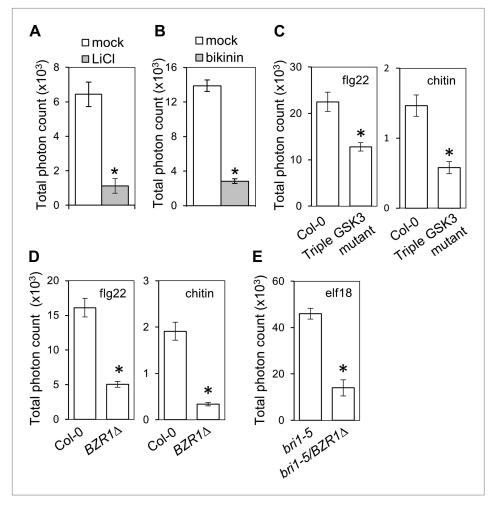


Figure 1. Activation of BZR1 is sufficient to inhibit the PAMP-triggered ROS burst. (**A**) and (**B**) Flg22-triggered ROS burst after LiCl (**A**) or bikinin (**B**) treatment. Leaf discs were pre-treated with a 10 mM LiCl solution for 5 hr or with a 50 μM bikinin solution for 16 hr. (**C**) Flg22- or chitin-triggered ROS burst in Col-0 and the triple GSK3 mutant plants. (**D**) Flg22- or chitin-induced ROS burst in Col-0 and $BZR1\Delta$ plants. (**E**) Elf18-triggered ROS burst in *bri1-5* and *bri1-5*/BZR1Δ plants. In all cases, bars represent SE of n = 28 rosette leaf discs. Asterisks indicate a statistically significant difference compared to the corresponding control (mock treatment [**A** and **B**], Col-0 [**C** and **D**] or *bri1-5* [**E**]), according to a Student's t-test (p<0.05). Leaf discs of four- to five-week-old Arabidopsis plants were used in these assays. Flg22 and elf18 were used at a concentration of 50 nM; chitin was used at a concentration of 1 mg/ml. Total photon counts were integrated between minutes two and 40 after PAMP treatment. All experiments were repeated at least three times with similar results.



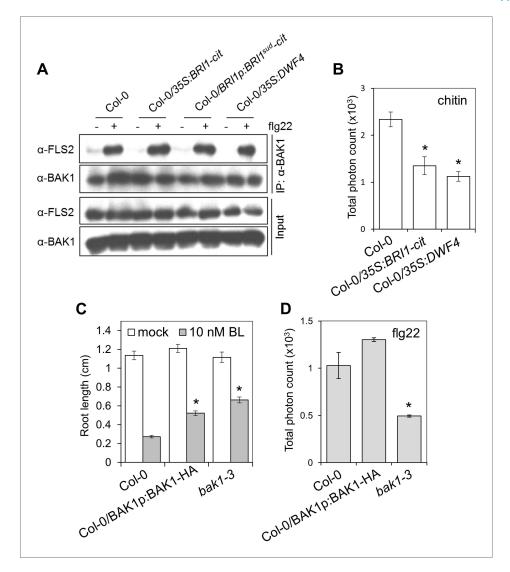


Figure 1—figure supplement 1. The BR-mediated suppression of PTI can be triggered independently of a competition for BAK1. (**A**) Co-IP of BAK1 and FLS2 in Col-0, 35S:BR11-cit, $35S:BR11^{sud}-cit$ and 35S:DWF4 seedlings treated with 1 μM flg22 for 10 min. Coimmunoprecipitated proteins were analyzed by using anti-FLS2 or anti-BAK1 antibodies. (**B**) Chitin-triggered ROS burst in Col-0, 35S:BR11-cit and 35S:DWF4 plants. Chitin was used at a concentration of 1 mg/ml. Total photon counts were integrated between minutes two and 40 after PAMP treatment. Bars represent SE of n = 28 rosette leaf discs. (**C**) Root length of seven-day-old Col-0, BAK1p:BAK1-HA (in Col-0 WT background) or bak1-3 seedlings grown on medium supplemented or not with 10 nM BL. Bars represent SE of $12 \le n \le 17$. Asterisks indicate a statistically significant difference between treatments according to a Student's t-test (p<0.05). (**D**) Flg22-triggered ROS burst in Col-0, BAK1p:BAK1-HA (in Col-0 WT background) or bak1-3 plants. Leaf discs of four- to five-week-old Arabidopsis plants were used in these assays. Flg22 was used at a concentration of 50 nM. Total photon counts were integrated between minutes two and 40 after PAMP treatment. Bars represent SE of n = 28 rosette leaf discs. All experiments were repeated at least twice with similar results. DOI: 10.7554/eLife.00983.004



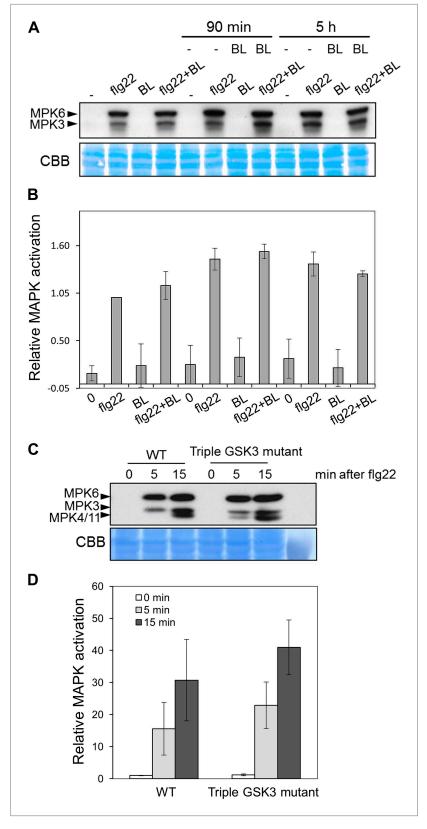


Figure 1—figure supplement 2. PAMP-triggered MAPK activation is not impaired upon activation of BR signaling. **(A)** MAPK activation in Col-0 seedlings upon treatment with 1 μ M flg22 (**F**) and/or epiBL (**B**) for 10 min (with or without a 90-min or 5-hr BL pre-treatment). **(B)** Quantification of total MAPK activation in the experiment shown in (**A**), Figure 1—figure supplement 2. Continued on next page



Figure 1—figure supplement 2. Continued

measured as pixel intensity using ImageJ. Results are the average of two independent blots, corresponding to two independent biological replicated. (\mathbf{C}) MAPK activation in Col-0 and Triple GSK3 mutant seedlings upon treatment with 1 μ M flg22. (\mathbf{D}) Quantification of total MAPK activation in the experiment shown in (\mathbf{C}), measured as pixel intensity using ImageJ. Results are the average of two independent blots, corresponding to two independent biological replicated. Proteins were separated in a 10% acrylamide gel and transferred to PVDF membranes. Membranes were blotted with phospho-p44/42 MAPK (Erk1/2; Thr202/Tyr204) rabbit monoclonal antibodies. Bars represent SD. DOI: 10.7554/eLife.00983.005

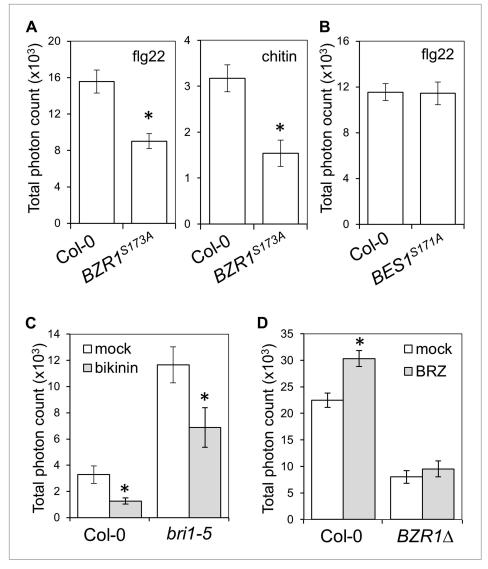


Figure 1—figure supplement 3. Activation of BZR1, but not BES1, is sufficient to inhibit the PAMP-triggered ROS burst. (A) Flg22- or chitin-triggered ROS burst in $BZR1^{5173A}$ plants. (B) Flg22-triggered ROS burst in $BES1^{5171A}$ plants. (C) Flg22-triggered ROS burst in mock- or bikinin-treated Col-0 or bri1-5 plants. Leaf discs were pre-treated with a 50 μM bikinin solution for 16 hr. (D) Flg22-triggered ROS burst in mock- or BRZ-treated Col-0 or $BZR1\Delta$ plants. Leaf discs were pre-treated with a 2.5 μM BRZ solution for 16 hr. In all cases, bars represent SE of 21 ≤ n ≤ 28. Asterisks indicate a statistically significant difference compared to Col-0 (A and B) or mock-treatment (C and D) according to a Student's t-test (p<0.05). Flg22 was used at a concentration of 50 nM; chitin was used at a concentration of 1 mg/ml. Total photon counts were integrated between minutes two and 40 after PAMP treatment.



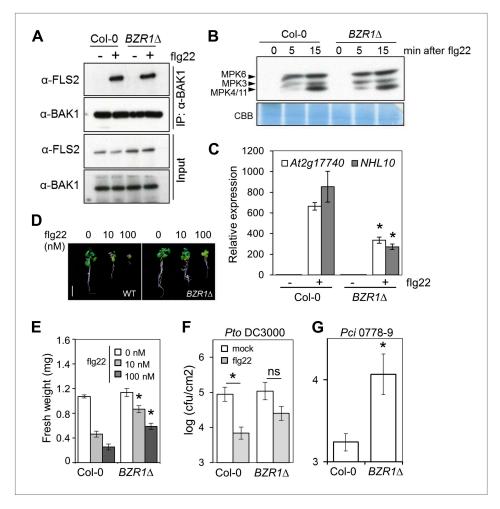


Figure 2. Activation of BZR1 results in the suppression of specific PTI outputs. (A) Co-immunoprecipitation (Co-IP) of BAK1 and FLS2 in Col-0 and $BZR1\Delta$ seedlings after 10 min mock (−) or 1 μM flg22 (+) treatment. Proteins were separated in a 10% acrylamide gel and transferred to PVDF membranes. Membranes were blotted with anti-FLS2 or anti-BAK1 antibodies. (B) MAPK activation in Col-0 and $BZR1\Delta$ seedlings upon 1 μM flg22 treatment. Proteins were separated in a 10% acrylamide gel and transferred to PVDF membranes. Membranes were blotted with phospho-p44/42 MAPK (Erk1/2; Thr202/Tyr204) rabbit monoclonal antibodies. CBB: Coomassie brilliant blue. (C) Marker gene (At2g17700 and NHL10) expression in Col-0 and $BZR1\Delta$ seedlings after 1 hr mock (−) or 1 μM flg22 (+) treatment, as determined by qPCR. Bars represent SE of n = 3. (D) and (E) Seedling growth inhibition of 10-day-old Col-0 or $BZR1\Delta$ seedlings induced by increasing concentrations of flg22, as indicated. Scale bar (D), 1 cm. Bars (E) represent SE of 8 ≤ n ≤ 16. (F) Flg22-induced resistance to *P. syringae* pv. tomato DC3000 in Col-0 and $BZR1\Delta$ plants. Plants were pre-treated with 1 μM flg22 or water 24 hr prior to bacterial infiltration. Bars represent SE of n = 4. This experiment was repeated seven times with similar results. (G) Susceptibility of Col-0 and $BZR1\Delta$ plants to *P. syringae* pv. cilantro 0788-9. Bars represent SE of n = 4. Asterisks indicate a statistically significant difference compared to Col-0 according to a Student's t-test (p<0.05); ns = not significant. All experiments were repeated at least twice with similar results unless otherwise stated.



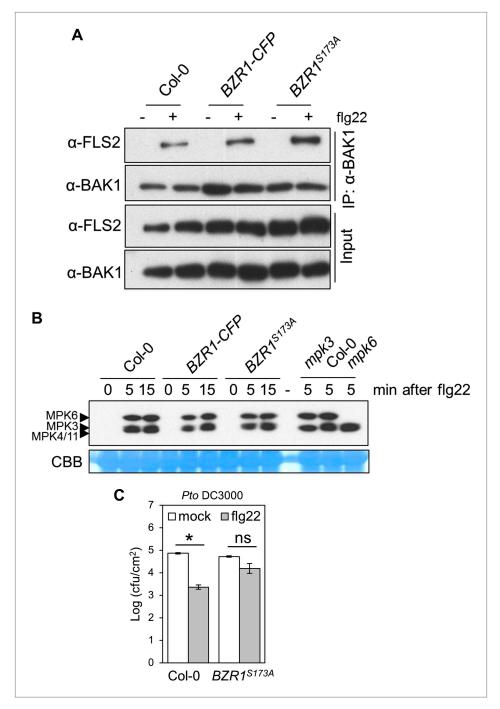


Figure 2—figure supplement 1. Expression of the constitutively active BZR1^{5173A} results in the suppression of specific PTI outputs. (**A**) Co-IP of BAK1 and FLS2 in Col-0 and $BZR1^{5173A}$ seedlings treated with 1 μ M flg22 for 10 min. Co-immunoprecipitated proteins were analyzed by using anti-FLS2 or anti-BAK1 antibodies. (**B**) MAPK activation in Col-0 and $BZR1^{5173A}$ seedlings upon treatment with 1 μ M flg22. Membranes were blotted with phospho-p44/42 MAPK (Erk1/2; Thr202/Tyr204) rabbit monoclonal antibodies. (**C**) Flg22-induced resistance to Pto DC3000 in $BZR1^{5173A}$ plants. Plants were pre-treated with 1 μ M flg22 or water 24 hr prior to bacterial inoculation. Bars represent SE of n = 4. Asterisks indicate a statistically significant difference compared to mock-treated plants according to a Student's t-test (p<0.05); ns = not significant. All experiments were repeated at least twice with similar results.



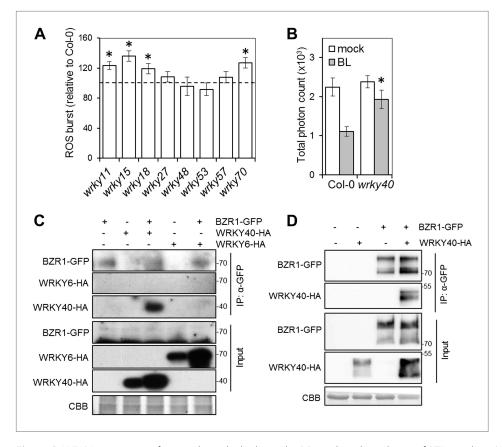


Figure 3. WRKY transcription factors play a dual role on the BR-mediated regulation of PTI signaling. (A) Flg22triggered ROS burst in mutants in each BR-induced BZR1-targeted WRKY. Leaf discs of four- to five-week-old Arabidopsis plants were used in these assays. Flg22 was used at a concentration of 50 nM. Total photon counts were integrated between minutes two and 40 after PAMP treatment. Bars represent SE of n = 28. Asterisks indicate a statistically significant difference compared to Col-0 according to a Student's t-test (p<0.05). (B) Flg22-triggered ROS burst in epiBL (BL)- or mock- pre-treated wrky40 mutant or wild-type plants. Leaf discs of four- to five-week-old plants were pre-treated with a 1 µM BL solution or mock solution for 8 hr. Flg22 was used at a concentration of 50 nM. Total photon counts were integrated between minutes two and 40 after PAMP treatment. Bars represent SE of n = 21. Asterisks indicate a statistically significant difference compared to Col-0 according to a Student's t-test (p<0.05). (C) Co-IP of BZR1-GFP transiently expressed in N. benthamiana, alone or together with WRKY40-HA or WRKY6-HA. BZR1-GFP was immunoprecipitated with an anti-GFP antibody. Immuniprecipitated or total proteins were separated in a 10% acrylamide gel and transferred to PVDF membranes. Membranes were blotted with anti-HA or anti-GFP antibodies. CBB: Coomassie brilliant blue. (D) Co-IP of BZR1-GFP transiently expressed in Arabidopsis protoplasts, alone or together with WRKY40-HA. BZR1-GFP was immunoprecipitated with an anti-GFP antibody. Immuniprecipitated or total proteins were separated in a 10% acrylamide gel and transferred to PVDF membranes. Membranes were blotted with anti-HA or anti-GFP antibodies. CBB: Coomassie brilliant blue. All experiments were repeated at least twice with similar results.



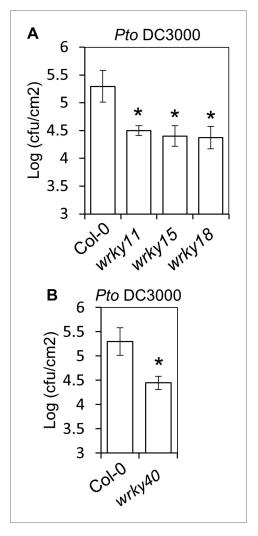


Figure 3—figure supplement 1. Mutants in WRKY11, WRKY15, WRKY18 and WRKY40 are more resistant to Pto DC3000. (A) and (B) Pto DC3000 infections in Col-0, wrky11, wrky15, and wrky18 (A) and in Col-0 and wrky40 (B) plants. Bars represent SE of n = 4. Asterisks indicate a statistically significant difference compared to Col-0 plants according to a Student's t-test (p<0.05); ns = not significant. All experiments were repeated at least three times with similar results.



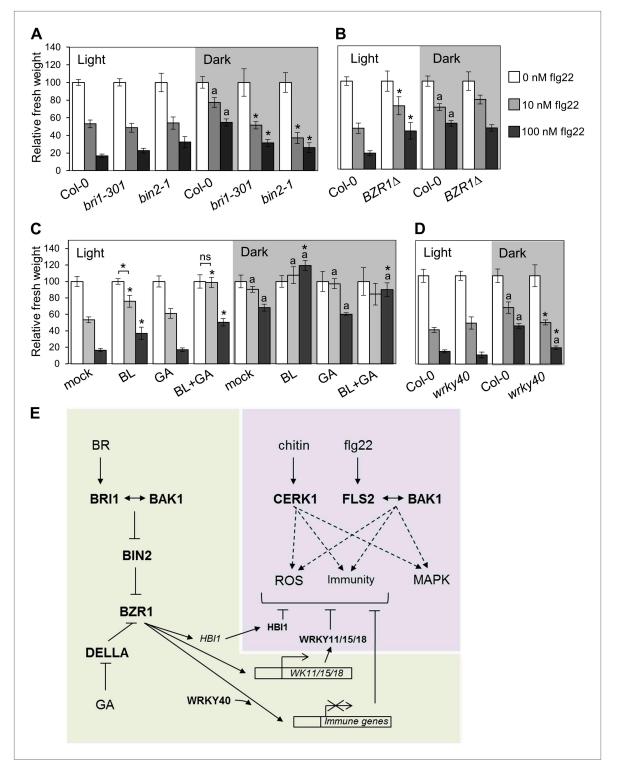


Figure 4. Activation of BR signaling and BZR1 prioritizes growth over immunity in the dark. (**A**) and (**B**) Relative seedling growth inhibition of 10-day-old (**A**) Col-0, bri1-301 and bin2-1 or (**B**) Col-0 and $BZR1\Delta$ seedlings induced by increasing concentrations of flg22 in either light or dark. (**C**) Relative seedling growth inhibition of 10-day-old Col-0 seedlings grown on medium supplemented or not with BL (1 μM), GA (1 μM), BL+GA (1 μM + 1 μM) or mock solution in light or dark. (**D**) Relative seedling growth inhibition of Col-0 or wrky40 seedlings induced by increasing concentrations of flg22 in either light or dark. Bars represent SE of n = 16 (**A**, **B** and **D**) or n = 8 (**C**) Asterisks indicate a statistically significant difference compared to Col-0 in the same condition (light or dark and same concentration of flg22), according to a Student's t-test (p<0.05); 'a' indicates a statistically significant difference compared to the same genotype/treatment and concentration of flg22 in light, according to a Student's t-test (p<0.05). All experiments were repeated at Figure 4. Continued on next page



Figure 4. Continued

least three times with similar results. Values are relative to Col-0 (A, B and D) or mock-treated seedlings (C) (set to 100). Absolute values of these experiments are shown in *Figure 4—figure supplement 3*. (E) Schematic model depicting the BZR1-mediated inhibition of PTI. Upon BR- and DELLA-dependent activation, BZR1 induces the expression of negative regulators of PTI, such as *WRKY11*, *WRKY15*, *WRKY18*, or *HBI1*. In addition, BZR1 also inhibits the expression of immune genes, acting cooperatively with WRKY40 and possibly other WRKYs. Ultimately, the BZR1-mediated changes in transcription would lead to the suppression of PTI signaling. The PTI signaling pathway is shadowed in violet; the BR signaling pathway is shadowed in green.



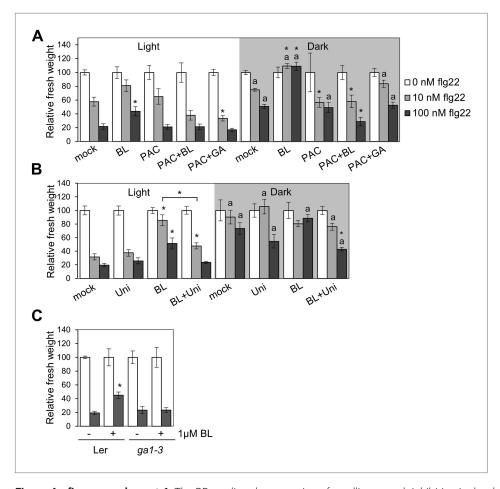


Figure 4—figure supplement 1. The BR-mediated suppression of seedling growth inhibition in the dark requires GA synthesis. Seedling growth inhibition of 10-day-old Col-0 seedlings grown on medium supplemented or not with (A) BL (1 μ M), paclobutrazol (PAC; 1 μ M), BL+PAC (1 μ M + 1 μ M) and PAC+GA (1 μ M + 1 μ M), or (B) uniconazole (Uni; 100 μ M), BL (1 μ M) and Uni+BL (100 μ M + 1 μ M) induced by increasing concentrations of flg22 in light or dark. (C) Seedling growth inhibition of 10-day-old Ler and ga1-3 seedlings grown on medium supplemented or not with BL (1 μ M). Bars represent SE of n = 8. Asterisks indicate a statistically significant difference compared to Col-0 in the same condition (light or dark and same concentration of flg22), according to a Student's t-test (p<0.05); 'a' indicates a statistically significant difference compared to the same genotype/treatment and concentration of flg22 in light, according to a Student's t-test (p<0.05). Values are relative to mock-treated seedlings (set to 100). All experiments were repeated at least twice with similar results.



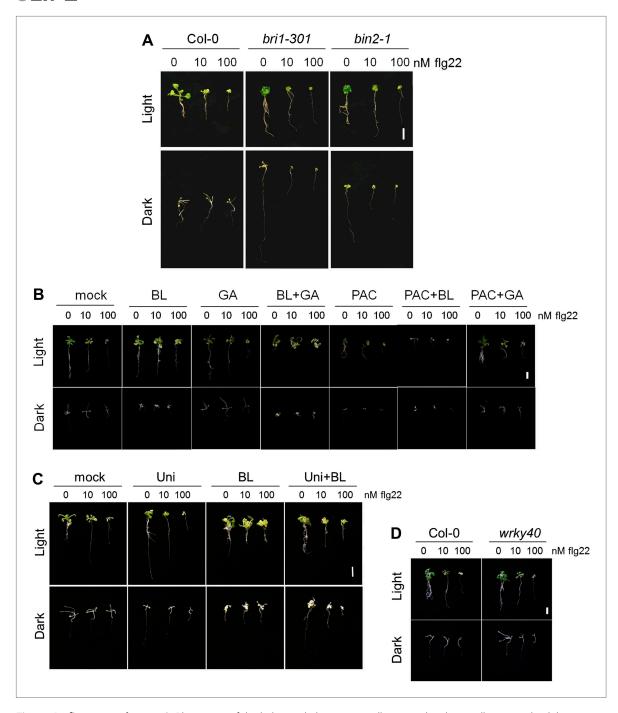


Figure 4—figure supplement 2. Phenotype of the light- or dark-grown seedlings used in the seedling growth inhibition assays (Figure 4 and Figure 4—figure supplement 1). Representative seedlings of the seedling growth inhibition experiments depicted in: (A) Figure 4A; (B) Figure 4—figure supplement 1A; (C) Figure 4-figure supplement 1B; (D) Figure 4D. Scale bar, 1 cm.

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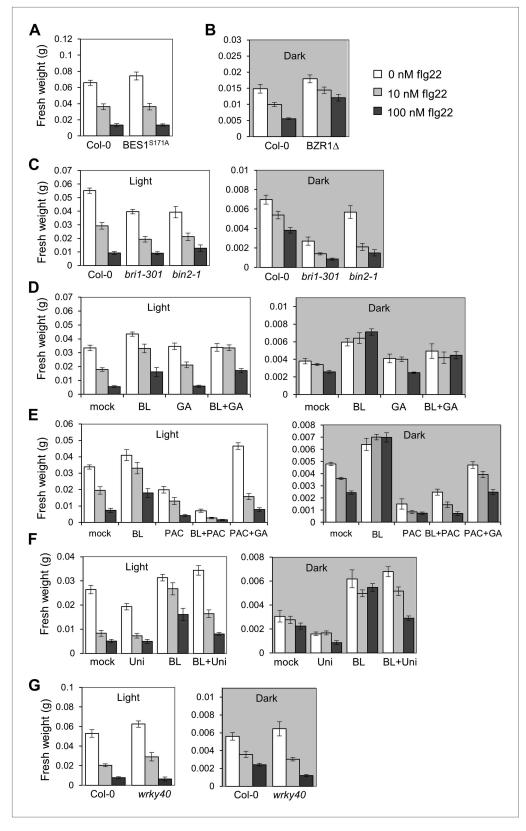


Figure 4—figure supplement 3. Absolute fresh weight values of seedling growth inhibition assays. **(A)** Seedling growth inhibition of 10-day-old Col-0 or *BES1*^{5171A} seedlings induced by increasing concentrations of flg22. **(B)** Figure 4—figure supplement 3. Continued on next page



Figure 4—figure supplement 3. Continued

Absolute fresh weight values of the seedling growth inhibition assay depicted in *Figure 4B*, dark. (C) Absolute fresh weight values of the seedling growth inhibition assay depicted in *Figure 4A*. (D) Absolute fresh weight values of the seedling growth inhibition assay depicted in *Figure 4C*. (E) Absolute fresh weight values of the seedling growth inhibition assay depicted in *Figure 4—figure supplement 1A*. (F) Absolute fresh weight values of the seedling growth inhibition assay depicted in *Figure 4—figure supplement 1B*. (G) Absolute fresh weight values of the seedling growth inhibition assay depicted in *Figure 4D*. Error bars represent SE as indicated in *Figure 4*, *Figure 4—figure supplement 1*.