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ABSTRACT

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Little is known about the excess mortality caused by multidrug-resistant (MDR) bacterial infection in low- and middle-income countries (LMICs). We retrospectively obtained microbiology laboratory and hospital databases of nine public hospitals in northeast Thailand from 2004 to 2010, and linked these with the national death registry to obtain the 30-day mortality outcome. The 30-day mortality in those with MDR community-acquired bacteraemia, healthcare-associated bacteraemia, and hospital-acquired bacteraemia were 35% (549/1,555), 49% (247/500), and 53% (640/1,198), respectively. We estimate that 19,122 of 45,209 (43%) deaths in patients with hospital-acquired infection due to MDR bacteria in Thailand in 2010 represented excess mortality caused by MDR. We demonstrate that national statistics on epidemiology and burden of MDR in LMICs could be improved by integrating information from readily available databases. The prevalence and mortality attributable to MDR in Thailand are high. This is likely to reflect the situation in other LMICs.

INTRODUCTION

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The emergence of antimicrobial resistance (AMR) is of major medical concern, particularly in low- and middle-income countries (LMICs). 1,2 In LMICs, antibiotic use is increasing with rising incomes, affordable antimicrobials and the lack of stewardship in hospital and poor control of over-the-counter sales. This is driving the emergence and spread of multidrug-resistant (MDR) pathogens in community and hospital settings. Hospital data from LMICs suggest that the cumulative incidence of community-acquired Extended-Spectrum Beta-Lactamase (ESBL) producing Escherichia coli and Klebsiella pneumoniae infections are increasing over time. 3,4 A recent report from the International Nosocomial Infection Control Consortium (INICC) also showed that the prevalence of AMR organisms causing hospital-acquired infections (HAI) in ICUs in LMICs is much higher than those in the United States (US).⁵ Attributable mortality, generally defined as the difference in mortality between those with and without the condition of interest, is an important parameter used to estimate the burden of AMR. In the US, it is estimated that mortality from infection attributable to AMR is 6.5%, 6 leading to an estimate of 23,000 deaths attributable to AMR each year. In the European Union, it is estimated that the number of deaths attributable to selected antibiotic-resistant bacteria is about

25,000 each year.8 There is limited information on mortality attributable to AMR in LMICs. The

mortality attributable to ventilator-associated pneumonia in ICUs in Colombia, Peru, and Argentina is estimated to be 17%, 25%, and 35%, and is associated with a high percentage of AMR organisms. The mortality attributable to ESBL and methicillin-resistance Staphylococcus aureus (MRSA) is estimated to be 27% and 34% in neonatal sepsis in Tanzania, which has been used to postulate an estimate that 58,319 deaths could be attributable to ESBL and MRSA in India alone. In an effort to harmonize the surveillance systems of AMR, a joint initiative between the European Centre for Disease Prevention and Control (ECDC) and the Centres for Disease Prevention and Control (CDC) have developed standard definitions of multidrug-resistance (MDR).

We recently combined large data sets from multiple sources including microbiology databases, hospital admission databases, and the national death registry from a sample of ten public hospitals in northeast Thailand from 2004 to 2010.^{3,14} We defined community-acquired bacteraemia (CAB) as the isolation of a pathogenic bacterium from blood taken in the first 2 days of admission and without a hospital stay in the 30 days prior to admission, healthcare-associated bacteraemia (HCAB) as the isolation of a pathogenic bacterium from blood taken in the first 2 days of admission and with a hospital stay within 30 days prior to the admission, and hospital-acquired bacteraemia (HAB) as the isolation of a pathogenic bacterium from blood

taken after the first 2 days of admission.^{3,14} We reported an increase in the incidence of CAB, HCAB and HAB over the study period, and that bacteraemia was associated with high case fatality rates (37.5%, 41.8% and 45.5%, respectively).^{3,14} Here, we apply ECDC/CDC standard definitions of MDR to this large data set to evaluate the prevalence, trends, and mortality attributable to MDR bacteria isolated from the blood. We then estimate the number of deaths attributable to MDR in Thailand nationwide.

RESULTS

We contacted all 20 provincial hospitals in Northeast Thailand to participate in the study. All provincial hospitals were equipped with all basic medical specialties and intensive care units (ICUs). Agreement was obtained from 15 (75%) hospitals, of which ten had hospital databases and microbiological laboratory databases as electronic files in a readily accessible format. Of these ten hospitals, nine had databases of antimicrobial susceptibility testing results as electronic files for the study (Figure 1). The median bed number for the nine hospitals included in the analysis was 450 beds (range 300 to 1,000 beds). Of these, three had data available for the period 2004-2010, two between 2007-2010, three between 2008-2010 and one between 2009-2010. Overall, 1,803,506 admission records from 1,255,571 patients were evaluated. A total of 20,803 (1.2%) admission records had at least one blood culture positive for pathogenic

organisms during admission. Of 10,022 patients with first episodes of bacteraemia caused by *S. aureus, Enterococcus* spp, *E. coli, K. pneumoniae, P. aeruginosa* and *Acinetobacter* spp., 226 patients (2%) were excluded because the causative organisms were tested for susceptibility to fewer than three antimicrobial categories. Therefore, a total of 9,796 first episodes of bacteraemia caused by *S. aureus* (n=1,881), *Enterococcus* spp (n=342), *E. coli* (n=4,279), *K. pneumoniae* (n=1,661), *P aeruginosa* (n=568), and *Acinetobacter* spp. (n=1,065) were evaluated in the analysis. The proportion of bacteria being MDR was highest in HAB and lowest in CAB for all organisms (all p<0.001 except for *Enterococcus* spp., Table 1).

Staphylococcus aureus

Of CAB, HCAB and HAB caused by *S. aureus*, 8%, 28%, and 50% were caused by MDR *S. aureus*, respectively (p<0.001). Almost all MDR *S. aureus* were MRSA (92% [357/389], Table 2). We did not observe a trend in the proportion of *S. aureus* bacteremia being caused by MRSA (Figure 2). Vancomycin non-susceptible *S. aureus* was found in <1% of tested isolates (6/1,380).

Enterococcus species

MDR *Enterococcus* spp. were not found in CAB (0/176) and HCAB (0/49), while 3% (4/117) of *Enterococcus* spp. causing HAB were MDR. Of CAB caused by *Enterococcus* spp., 15% (20/134) and 23% (35/153) was non-susceptible to ampicillin and gentamicin, respectively (Table3), while 42% (34/81) and 62% (63/101) of HAB caused by *Enterococcus* spp. were non-susceptible to those agents, respectively (both p<0.001). Vancomycin non-susceptible *Enterococcus* spp. was found in 4% of tested isolates (15/338).

Escherichia coli

Of CAB, HCAB and HAB caused by *E. coli*, 35%, 58% and 63% were caused by MDR *E. coli*, respectively (p<0.001). Of *E. coli* causing CAB, 79% (2,246/2,843), 16% (501/3,076), 24% (728/3,000), 58% (1,738/3,007), and 17% (559/3,346) were non-susceptible to commonly-used antimicrobials for community-acquired infections such as ampicillin, cefotaxime, ciprofloxacin, trimethoprim-sulphamethoxazole, and gentamicin, respectively (Table 4). From 2004 to 2010, the proportions of community-acquired *E. coli* bacteraemia being caused by *E. coli* non-susceptible to extended-spectrum cephalosporins rose from 5% (9/169) to 23% (186/815) (p=0.04) (Figure 3). The proportions of healthcare-associated and hospital-acquired *E. coli* bacteraemia being caused by *E. coli* non-susceptible to extended-spectrum cephalosporins

was not observed (p=0.18 and p=0.63, respectively). Carbapenem non-susceptible *E. coli* was found in <1% of tested isolates (12/3,838).

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Klebsiella pneumoniae

Of CAB, HCAB and HAB caused by K. pneumoniae, 14%, 36%, and 66% were caused by MDR K. pneumoniae, respectively (p<0.001). Of K. pneumoniae causing CAB, 16% (146/902), 16% (143/894), 23% (198/876), and 9% (94/999) were non-susceptible to cefotaxime, ciprofloxacin, trimethoprim-sulphamethoxazole and gentamicin, respectively (Table 5). From 2004 to 2010, the proportions of community-acquired K. pneumoniae bacteraemia being caused by K. pneumoniae non-susceptible to extended-spectrum cephalosporins rose from 12% (6/50) to 24% (64/263) (p=0.04) (Figure 4). The proportions of healthcare-associated and hospitalacquired K. pneumoniae bacteraemia being caused by K. pneumoniae non-susceptible to extended-spectrum cephalosporins were also high (40% [71/177] and 71% [304/431], respectively), but a significant trend over time was not observed (p=0.16 and p=0.35, respectively). Carbapenem non-susceptible K. pneumoniae was found in <1% of tested isolates (11/1,555).

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Pseudomonas aeruginosa

Of CAB, HCAB and HAB caused by *P. aeruginosa*, 5%, 10%, and 25% were caused by MDR *P. aeruginosa*, respectively (p<0.001). Of *P. aeruginosa* causing HAB, 38% (68/179), 27% (48/177), 23% (39/169) and 26% (42/164) were non-susceptible to commonly-used antimicrobials for HAI such as ceftazidime, amikacin, ciprofloxacin and carbapenems, respectively (Table 6). We did not observe a trend in the proportions of *P. aeruginosa* being caused by *P. aeruginosa* that were non-susceptible to any specific antibiotic group (Figure 5).

Acinetobacter species.

Of CAB, HCAB and HAB caused by *Acinetobacter* spp., 28%, 50%, and 75% were caused by MDR *Acinetobacter* spp., respectively (p<0.001). Of *Acinetobacter* spp. causing HAB, 75% (377/500), 63% (310/495), 67% (322/481) and 64% (315/490) were non-susceptible to ceftazidime, amikacin, ciprofloxacin and carbapenems, respectively (Table 7). There was borderline evidence that the proportion of hospital-acquired *Acinetobacter spp.* bacteremia being caused by *Acinetobacter spp.* non-susceptible to carbapenem rose from 49% (19/39) in 2004 to 65% (70/108) in 2010 (p=0.10) (Figure 6). Non-susceptibility to colistin was observed in 3% of tested isolates (2/63).

Mortality attributable to MDR

The 30-day mortality in patients with CAB, HCAB and HAB caused by MDR bacteria was 35% (549/1,555), 49% (247/500), and 53% (640/1,198), compared with 32% (1,595/4,924), 37% (264/716), and 42% (383/903) in CAB, HCAB, and HAB caused by non-MDR bacteria, respectively (Figure 7). In the final multivariable logistic regression model, gender, age group, year of admission and time to bacteraemia (for HAB) were included (Supplementary file 2).

If excess mortality in patients infected with MDR bacteria after adjusting for confounding factors in the final multivariable model is assumed to be caused by MDR, the mortality attributable to MDR was 7% (95%CI 4% to 10%, p<0.001) in CAB, 15% (95%CI 5% to 24%, p<0.001) in HCAB and 15% (95%CI 2% to 27%, p<0.001) in HAB (Figure 2). Heterogeneity between different organisms was clearly observed in HAB (p<0.001), and borderline evidence of heterogeneity was observed in HCAB (p=0.09). The heterogeneity observed in HCAB and HAB was largely caused by MDR *Acinetobacter* spp. (Figure 7B and 7C). Mortality attributed to MDR was highest for hospital-acquired MDR *Acinetobacter* bacteraemia (41%).

Using our estimated mortality attributed to MDR bacteraemia (Figure 7C) and national statistics of HAI caused by MDR bacteria, we further estimated that 19,122 of 45,209 (43%) deaths in patients with HAI due to MDR bacteria in Thailand in 2010 represented excess mortality caused

by MDR (Table 8). All parameters used to estimate the number of excess deaths in Thailand are shown in Supplementary file 2.

DISCUSSION

This study presents detailed antimicrobial susceptibility data on common pathogenic bacteria, the association of MDR with infection acquisition (community-acquired, healthcare-associated and hospital-acquired), and excess mortality from MDR in a developing country. Our estimate of excess deaths caused by MDR in HAI patients in Thailand (19,122 deaths per year in a country of about 66 million population in 2010) is large compared to those estimated in USA (23,000 death per year in a country of 316 million population in 2013)⁷ and the European Union (25,000 deaths per year in EU of about 500 million population in 2007).⁸ Our study highlights the need for public health officials and international health organizations to improve systems to track and reduce the burden of AMR in LMICs. Our estimated mortality for those with MDR HAI (45,209, Table 2) is higher than those previously published by Pumart et al. (38,481)¹⁵, probably because we used 30-day mortality rather than in-hospital mortality.

Acinetobacter spp. is increasingly recognized as an important cause of HAI,^{16,17} and our study confirms the importance of this species as a leading cause of hospital-acquired MDR infection in

a developing tropical country. ^{14,18} The high mortality observed in MDR *Acinetobacter* spp. bacteraemia is because treatment options are limited and those available are associated with toxicity. ¹⁹ The high proportions of *S. aureus*, *E. coli* and *K. pneumoniae* bactaeremia being caused by MRSA and *E. coli* and *K. pneumoniae* non-susceptible to extended-spectrum cephalosporins, respectively, are consistent with previous reports from other tropical countries. ⁹⁻¹¹ The rising proportions of community-acquired *E. coli* and *K. pneumoniae* bacteremia being caused by *E. coli* and *K. pneumoniae* non-susceptible to extended-spectrum cephalosporins, and the rising proportion of hospital-acquired *Acinetobacter* bacteraemia being causing *Acinetobacter* non-susceptible to carbapenem suggest that the burden of AMR in Thailand is deteriorating over time.

A limitation of this study is that more complete clinical data were not available. Mortality attributable to MDR could be overestimated if MDR infection was associated with more severely ill patients in ICUs. However, our estimated attributable mortality is comparable to the previous reports. For example, our estimated mortality attributable to MDR *Acinetobacter* bacteraemia (40.6%) is comparable to the mortality attributable to imipenem resistant *Acinetobacter* bacteraemia reported by Kwon *et al.* in Korea (41.1%), which was adjusted by severity of illness.^{20,21} In addition, data on hospitalization in other hospitals not participating in the study (for

example, a smaller community hospital or a private hospital in the province) were not available, which could have resulted in a misclassification of CAB, HCAB and HAB in some cases. We also note that data on attributable mortality from different countries is difficult to compare because of the differing study designs. For example, our mortality outcome is the overall 30-day mortality, including both directly and indirectly contributed to MDR, while an EU study only considered directly attributable deaths.⁸ The p values for trends were generated by the stratification method; therefore, the analysis was not biased towards the increasing availability of the hospital data over the study period. Nonetheless, the trends could be affected by an increasing use of blood culture, changes in antimicrobial agents tested for susceptibility, and greater standardization of laboratory methodologies over time.²³ It is likely that burdens of MDR similar to that observed in our study are present in many secondary and tertiary hospitals in tropical LMICs, particularly where extended-spectrum cephalosporins and carbapenem are widely used. Nonetheless, resources for diagnostics, methodologies used in the laboratories, and study designs need to be carefully considered when performing a comparison between different settings.

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Despite the increasing global focus on AMR in LMICs, considerable gaps remain in our understanding of the scale of the problem. We have demonstrated that the integration of information from readily available routinely collected databases can provide valuable information on the trends and mortality attributable to AMR in Thailand. The methodology used in our study could be applied to explore the burden of AMR in other LMICs where microbiological facilities and hospital admission database are available.

MATERIALS AND METHODS

Study Population

From 2004 to 2010, Thailand was classified as a lower-middle income country with an average income of \$4,782 per person per year in 2010.²² Northeast Thailand consists of 20 provinces covering 170,226 km² and had an estimated population of 21.4 million in 2010. A large proportion of the population in this area lives in rural settings, with most adults engaging in agriculture with an emphasis on rice farming. Healthcare in Thailand is mainly provided by government-owned hospitals. Each province has a provincial hospital, which provides services and care to individuals within its catchment area. Additionally, provincial hospitals act as referral hospitals for smaller community hospitals for severely ill patients. All provincial hospitals receive

comparable resources, which are proportional to the respective populations of the provinces.

Provincial hospitals, unlike smaller community hospitals, are equipped with a microbiology laboratory capable of performing bacterial culture using standard methodologies for bacterial identification and susceptibility testing provided by the Bureau of Laboratory Quality and Standards, Ministry of Public Health (MoPH), Thailand.²³ During the study period, antimicrobial susceptibility was determined in all study hospitals using the disc diffusion method according to Clinical and Laboratory Standards Institute (CLSI).²⁴

Study Design

We conducted a retrospective, multicentre surveillance study of all provincial hospitals in Northeast Thailand. From the hospitals that agreed to participate, data were collected from microbiology and hospital databases between January 2004 and December 2010. Hospital number (HN) and admission number (AN) were used as a record linkage between the two databases and to identify individuals who had repeat admissions. The death registry for Northeast Thailand was obtained from the Ministry of Interior (MoI), Thailand, and used to identify patients who were discharged from hospital but died at home shortly after, which is a common practice in Thailand.^{3,14} Ethical permission for this study was obtained from the Ethical and Scientific Review Committees of the Faculty of Tropical Medicine, Mahidol University, and

of the MoPH, Thailand. Written consent was given by the directors of the hospitals to use their routine hospital database for research. Consent was not sought from the patients as this was a retrospective study, and the Ethical and Scientific Review Committees approved the process.

Data Collection

The microbiology laboratory data collected included hospital number (HN), admission number (AN), specimen type, specimen date, culture result, and antibiotic susceptibility profile (antibiogram). We consulted with study sites when the results of antimicrobial susceptibility testing were unclear. Hospital admission data were collected from the routine in-patient discharge report, which is regularly completed by attending physicians and reported to the MoPH, Thailand, as part of the national morbidity and mortality reporting system. The data collected included HN, AN, national identification 13-digit number, admission date, and discharge date. Date of death was also extracted from this record. Data collected from the national death registry obtained from the Mol included the national identification 13-digit number and the date of death.

Definitions

Bacteraemia was classified as CAB, HAB and HCAB as described previously.^{3,14} Polymicrobial infection was defined in patients who had more than one species of pathogenic organisms isolated from the blood during the same episode, and was excluded from the analysis.

Information on the incidence of CAB, HCAB and HAB from all pathogenic organisms has been published previously.^{3,14}

The 30-day mortality of CAB and HCAB was defined as death within 30 days of the admission date. The 30-day mortality of HAB was determined on the basis of a record of death within 30 days of the positive blood culture taken as recorded in the routine hospital database or by a record of death in the national death registry. In the event that a patient had more than one episode of bacteraemia, only the first episode was included in the study.

The standard definition of MDR proposed by ECDC/CDC was used.¹³ In brief, MDR was defined as non-susceptibility to at least one agent in three or more antimicrobial categories. Additionally, methicillin-resistant *Staphylococcus aureus* (MRSA) were automatically described as MDR.¹³

Statistical Analysis

Pearson's chi-squared test and Fisher's Exact test were used to compared categorical variables. A non-parametric test for trends was used to assess change in prevalence of antimicrobial resistance over time stratified by hospital (using the npt s command in STATA).

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Mortality of patients with a first episode of HAB, HCAB and HAB caused by S. aureus, Enterococcus spp., E. coli, K. pneumoniae, Pseudomonas aeruginosa, and Acinetobacter spp. were evaluated in relation to MDR. We selected these organisms based on guidelines for MDR proposed by ECDC/CDC, ¹³ and the fact that *E. coli* and *K. pneumoniae* were the most common causes of bacteraemia caused by Enterobacteriaceae in our setting. 3,14 Isolates tested for less than three antimicrobial categories were excluded from the analysis because they were not applicable to ECDC/CDC standard definitions of MDR. To examine the association between MDR and mortality, we performed a multivariable logistic regression analysis adjusting for a priori selected baseline confounders. To take account of the fact that patients with CAB, HCAB, and HAB were different populations with different definitions of 30-day mortality, we applied models to each group (CAB, HCAB and HAB) separately. Multivariable logistic regression models were developed using a purposeful selection method.²⁵ Potential confounding variables evaluated included age, gender and admission year. In the model for HAB, time to bacteremia was also evaluated as a potential confounder because there was evidence suggesting that time to HAI was associated with mortality from HAI.^{26,27} Time to bacteremia was defined as the duration between hospital admission and the date positive blood culture was taken. All models were stratified by hospital.

The mortality attributable to MDR was calculated using adjusted odds ratios (aORs) estimated by the final multivariable logistic regression models. If X was the observed mortality in patients with MDR infection, the estimated odds of mortality if they were infected with non-MDR organisms (O) would be $(1/aOR)^*(X/(1-X))$. Assuming that excess mortality was due to MDR, then the mortality attributable to MDR would be the absolute difference between mortality in patients with MDR infection (X) and the predicted mortality if they were infected with non-MDR organisms (O/(1+O)), which would be X - (O/(1+O)). Heterogeneity between different organisms within each group of patients (CAB, HCAB, and HAB) was assessed using the chisquared test, and the percentage of variation due to heterogeneity (I-square) was calculated.

The number of deaths attributable to MDR in Thailand was determined using methodology described previously.⁸ Data used included our estimated mortality attributable to MDR bacteraemia and cumulative incidence of HAI bacteraemia, lower respiratory track infection (LRTI), urinary tract infection (UTI), skin and soft tissue infection (SSTI), and other sites of

infection caused by MDR *S. aureus, E. coli, K. pneumoniae*, *P. aeruginosa*, and *Acinetobacter* spp. in Thailand in 2010, which have been described previously. Death attributable to MDR *Enterococcus* spp. was not included as the cumulative incidence of MDR *Enterococcus* infection in Thailand was not available. Attributable mortality by site of infection (LRTI, UTI, SSTI and other site) was estimated by applying correction factors corresponding to the relative mortality from infections of those sites compared to bacteraemia. All analyses were performed using STATA version 14.0 (StataCorp LP, College station, Texas, USA).

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Competing Interests

We declare that no competing interests exist.

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References

- 382 1. World Health Organization. Antimicrobial resistance: global report on surveillance 2014.
- 383 2014. http://apps.who.int/iris/bitstream/10665/112642/1/9789241564748 eng.pdf?ua=1
- 384 (accessed December 9, 2015).
- 2. Laxminarayan R, Duse A, Wattal C, et al. Antibiotic resistance-the need for global
- 386 solutions. Lancet Infect Dis 2013; **13**(12): 1057-98.
- 387 3. Kanoksil M, Jatapai A, Peacock SJ, Limmathurotsakul D. Epidemiology, microbiology
- and mortality associated with community-acquired bacteremia in northeast Thailand: a
- multicenter surveillance study. *PLoS One* 2013; **8**(1): e54714.
- 390 4. Ansari S, Nepal HP, Gautam R, et al. Community acquired multi-drug resistant clinical
- isolates of Escherichia coli in a tertiary care center of Nepal. Antimicrob Resist Infect Control
- 392 **2015**; **4**: 15.
- 393 5. Rosenthal VD, Maki DG, Mehta Y, et al. International Nosocomial Infection Control
- Consortium (INICC) report, data summary of 43 countries for 2007-2012. Device-associated
- 395 module. Am J Infect Control 2014; **42**(9): 942-56.
- Roberts RR, Hota B, Ahmad I, et al. Hospital and societal costs of antimicrobial-resistant
- infections in a Chicago teaching hospital: implications for antibiotic stewardship. *Clin Infect Dis*
- 398 2009; **49**(8): 1175-84.

- 399 7. Center for Disease Controls and Prevention, U.S. Department of Health and Human
- Services. Antibiotic resistance threats in the United States, 2013. 2013.
- 401 http://www.cdc.gov/drugresistance/pdf/ar-threats-2013-508.pdf (accessed February 4, 2016).
- 402 8. European Centre for Disease Prevention and Control and European Medicines Agency.
- 403 ECDC/EMEA Joint Technical Report The bacterial challenge: time to react, 2009. 2009.
- 404 http://ecdc.europa.eu/en/publications/Publications/0909 TER The Bacterial Challenge Time t
- 405 <u>o React.pdf</u> (accessed January 13, 2016).
- 406 9. Moreno CA, Rosenthal VD, Olarte N, et al. Device-associated infection rate and mortality
- in intensive care units of 9 Colombian hospitals: findings of the International Nosocomial
- Infection Control Consortium. *Infect Control Hosp Epidemiol* 2006; **27**(4): 349-56.
- 409 10. Cuellar LE, Fernandez-Maldonado E, Rosenthal VD, et al. Device-associated infection
- 410 rates and mortality in intensive care units of Peruvian hospitals: findings of the International
- Nosocomial Infection Control Consortium. Rev Panam Salud Publica 2008; **24**(1): 16-24.
- 11. Rosenthal VD, Guzman S, Orellano PW. Nosocomial infections in medical-surgical
- intensive care units in Argentina: attributable mortality and length of stay. *Am J Infect Control*
- 414 2003; **31**(5): 291-5.

- 12. Kayange N, Kamugisha E, Mwizamholya DL, Jeremiah S, Mshana SE. Predictors of
- positive blood culture and deaths among neonates with suspected neonatal sepsis in a tertiary
- 417 hospital, Mwanza-Tanzania. BMC Pediatr 2010; 10: 39.
- 418 13. Magiorakos AP, Srinivasan A, Carey RB, et al. Multidrug-resistant, extensively drug-
- resistant and pandrug-resistant bacteria: an international expert proposal for interim standard
- definitions for acquired resistance. Clin Microbiol infect 2012; **18**(3): 268-81.
- 421 14. Hongsuwan M, Srisamang P, Kanoksil M, et al. Increasing incidence of hospital-acquired
- and healthcare-associated bacteremia in northeast Thailand: a multicenter surveillance study.
- 423 PLoS One 2014; **9**(10): e109324.
- 424 15. Pumart P, Phodha T, Thamlikitkul V, Riewpaiboon A, Prakongsai P, Limwattananon
- S. Health and economic impacts of antimicrobial resistance in Thailand. *J Health Services*
- 426 Research Policy 2012; **9**(6): 352-60.
- 427 16. Munoz-Price LS, Weinstein RA. *Acinetobacter* infection. *N Engl J Med* 2008; **358**(12):
- 428 1271-81.
- 429 17. Peleg AY, Hooper DC. Hospital-Acquired Infections Due to Gram-Negative Bacteria. N
- 430 Engl J Med 2010; **362**(19): 1804-13.
- 431 18. Nhu NT, Lan NP, Campbell JI, et al. Emergence of carbapenem-resistant *Acinetobacter*
- baumannii as the major cause of ventilator-associated pneumonia in intensive care unit patients

- at an infectious disease hospital in southern Vietnam. J Med Microbiol 2014; 63(Pt 10): 1386-
- 434 94.
- 435 19. Fishbain J, Peleg AY. Treatment of *Acinetobacter* infections. *Clin Infect Dis* 2010; **51**(1):
- 436 79-84.
- 437 20. Kwon KT, Oh WS, Song JH, et al. Impact of imipenem resistance on mortality in patients
- with Acinetobacter bacteraemia. J Antimicrob Chemother 2007; **59**(3): 525-30.
- 439 21. Falagas ME, Rafailidis PI. Attributable mortality of *Acinetobacter baumannii*: no longer a
- 440 controversial issue. *Crit care* 2007; **11**(3): 134.
- 441 22. WorldBank. Thailand Overview. 2015.
- http://www.worldbank.org/en/country/thailand/overview (accessed 12/09/2015).
- 443 23. Opartkiattikul N, Bejrachandra S. The external quality assessment schemes in Thailand.
- 444 Rinsho Byori 2002; **50**(2): 121-5.
- 445 24. National Committee for Clinical Laboratory Standards. Performance Standards for
- 446 Antimicrobial disk susceptibility tests. NCCLS document M100-514. Wayne Pa: National
- 447 Committee for Clinical Laboratory Standards; 2004.
- 448 25. Bursac Z, Gauss CH, Williams DK, Hosmer DW. Purposeful selection of variables in
- logistic regression. Source Code Biol Med 2008; **3**: 17.

- 450 26. Moine P, Timsit JF, De Lassence A, et al. Mortality associated with late-onset
- pneumonia in the intensive care unit: results of a multi-center cohort study. *Intensive Care Med*
- 452 2002; **28**(2): 154-63.
- 453 27. Nguile-Makao M, Zahar JR, Français A, et al. Attributable mortality of ventilator-
- 454 associated pneumonia: respective impact of main characteristics at ICU admission and VAP
- onset using conditional logistic regression and multi-state models. *Intensive Care Med* 2010;
- **36**(5): 781-9.

- 457 28. Benichou J. A review of adjusted estimators of attributable risk. Statistical methods in
- 458 *medical research* 2001; **10**(3): 195-216.
- 459 29. Greenland S, Robins JM. Conceptual problems in the definition and interpretation of
- attributable fractions. *Am J Epidemiol* 1988; **128**(6): 1185-97.
- 461 30. Martone WJ, Jarvis WR, Edwards JR, Culver DH, Haley RW. Incidence and nature of
- endemic and epidemic nosocomial infections. In: Bennett JV & Brachman PS (eds) Hospital
- Infections, 4th ed Philadelphia, PA, USA: Lippincott-Raven; 1998: pp. 461-76.

465 Figure legends 466 Figure 1. Location of participating hospitals. These were situated in (1) Nong Khai, (2) Udon 467 468 Thani, (3) Nakhon Phanom, (4) Chaiyaphum, (5) Mukdahan, (6) Yasothon, (7) Burirum, (8) 469 Sisaket, and (9) Ubon Ratchathani provinces. 470 471 Figure 2. Trends in proportions of Staphylococcus aureus bacteremia being caused by 472 MRSA in Northeast Thailand. (A) community-acquired, (B) healthcare-associated and (C) 473 hospital-acquired Staphylococcus aureus bacteremia 474 475 Figure 3. Trends in proportions of Escherichia coli bacteremia being caused by E. coli 476 non-susceptible to extended-spectrum cephalosporins in Northeast Thailand. (A) 477 community-acquired, (B) healthcare-associated and (C) hospital-acquired E. coli bacteremia. 478 479 Figure 4. Trends in proportions of Klebsiella pneumoniae bacteremia being caused by K. 480 pneumoniae non-susceptible to extended-spectrum cephalosporins in Northeast 481 **Thailand.** (A) community-acquired, (B) healthcare-associated and (C) hospital-acquired K. 482 pneumoniae bacteremia.

| 483 | |
|-----|---|
| 484 | Figure 5. Trends in proportions of <i>Pseudomonas aeruginosa</i> bacteremia being caused by |
| 485 | P. aeruginosa non-susceptible to carbapenem in Northeast Thailand. (A) community- |
| 486 | acquired, (B) healthcare-associated and (C) hospital-acquired Pseudomonas aeruginosa |
| 487 | bacteremia. |
| 488 | |
| 489 | Figure 6. Trends in proportions of Acinetobacter spp bacteremia being caused by |
| 490 | Acinetobacter spp non-susceptible to carbapenem in Northeast Thailand. (A) community- |
| 491 | acquired, (B) healthcare-associated and (C) hospital-acquired Acinetobacter spp bacteremia. |
| 492 | |
| 493 | Figure 7. Forest plot of mortality in patients with MDR bacteremia compared with non- |
| 494 | MDR bacteremia in Northeast Thailand. (A) Community-acquired bacteraemia. (B) |
| 495 | Healthcare-associated bacteraemia. (C) Hospital-acquired bacteraemia. |
| 496 | |
| 497 | Figure 7 – source data 1 |

Table 1. Proportions of bacteraemias being caused by multidrug-resistant (MDR) variants of those

499 bacteria.

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| Pathogens | Community-acquired | Healthcare-associated | Hospital-acquired | P values |
|---------------------------|--------------------|-----------------------|-------------------|----------|
| | bacteraemia | bacteraemia | bacteraemia | |
| | (CAB) | (HCAB) | (HAB) | |
| MDR Staphylococcus aureus | 94/1176 (8%) | 73/259 (28%) | 222/446 (50%) | <0.001 |
| MDR Enterococcus spp | 0/176 (0%) | 0/49 (0%) | 4/117 (3%) | 0.02 |
| MDR Escherichia coli | 1177/3382 (35%) | 288/494 (58%) | 252/403 (63%) | <0.001 |
| MDR Klebsiella pneumoniae | 146/1010 (14%) | 71/196 (36%) | 301/455 (66%) | <0.001 |
| MDR Pseudomonas | 13/286 (5%) | 10/103 (10%) | 45/179 (25%) | <0.001 |
| aeruginosa | | | | |
| MDR Acinetobacter spp | 125/449 (28%) | 58/115 (50%) | 374/501 (75%) | <0.001 |

Table 2. Antibiogram of *S. aureus* causing bacteremia in Northeast Thailand.

| A 111 1 1 | A | CAR | LICAR | LIAD | |
|---------------------|-------------------|-------------------|------------------|------------------|--------|
| Antibiotic category | Antibiotic agents | CAB | HCAB | HAB | Ρ. |
| | | (n=1176 patients) | (n=259 patients) | (n=446 patients) | values |
| Aminoglycosides | Gentamicin | 24/484 (5%) | 16/84 (19%) | 66/151 (44%) | <0.001 |
| Ansamycins | Rifampin | 2/129 (2%) | 1/19 (5%) | 0/38 (0%) | 0.37 |
| Anti-MRSA | Ceftaroline | NA | NA | NA | - |
| cephalosporins | | | | | |
| Cefamycins | Oxacillin * | 80/1145 (7%) | 67/247 (27%) | 210/441 (48%) | <0.001 |
| Fluoroquinolones | Ciprofloxacin | 3/45 (7%) | 2/8 (25%) | 4/10 (40%) | 0.01 |
| | Moxifloxacin | NA | NA | NA | - |
| Folate pathway | Trimethoprim- | 99/1139 (9%) | 57/251 (23%) | 185/438 (42%) | <0.001 |
| inhibitors | sulphamethoxazole | | | | |
| Fucidanes | Fusidic acid | 33/618 (5%) | 4/170 (2%) | 12/291 (4%) | 0.26 |
| Glycopeptides | Vancomycin ** | 4/833 (0.5%) | 0/190 (0%) | 2/357 (1%) | 0.86 |
| | Teicoplanin | 2/66 (3%) | 1/17 (6%) | 0/17 (0%) | 0.72 |
| | Telavancin | NA | NA | NA | - |
| Glycylcyclines | Tigecycline | NA | NA | NA | - |
| Lincosamides | Clindamycin | 118/1147 (10%) | 77/251 (31%) | 202/438 (46%) | <0.001 |
| Lipopeptides | Daptomycin | NA | NA | NA | - |
| Macrolides | Erythromycin | 138/1116 (12%) | 76/240 (32%) | 222/429 (52%) | <0.001 |
| Oxazolidinones | Linezolid | 0/81 (0%) | 0/16 (0%) | 0/32 (0%) | - |
| Phenicols | Chloramphenicol | 6/86 (7%) | 4/24 (17%) | 2/14 (14%) | 0.21 |
| Phosphonic acids | Fosfomycin | 14/361 (4%) | 10/66 (15%) | 24/141 (17%) | <0.001 |
| Streptogramins | Quinupristin- | NA | NA | NA | - |
| | dalfopristin | | | | |
| Tetracyclines | Tetracycline | NA | NA | NA | - |
| | Doxycycline | NA | NA | NA | - |
| | Minocycline | NA | NA | NA | - |
| MDR | | 94/1176 (8%) | 73/259 (28%) | 222/446 (50%) | <0.001 |
| | | · · · | ·· | ·· | |

NOTE: Data are number of isolates demonstrating non-susceptible to the antimicrobial over the total number of isolates tested (%). CAB=Community-acquired bacteraemia, HCAB=Healthcare-associated bacteraemia, HAB= Hospital-acquired bacteraemia, and NA=Not available. The first isolate of each patient was used. MDR (one or more of these have to apply): (i) an MRSA is always considered MDR by virtue of being an MRSA (ii) non-susceptible to ≥ 1 agent in ≥ 3 antimicrobial categories.

^{*} Defined by using a 30 µg cefoxitin disc and an inhibition zone diameter of <21 mm.

^{**} Defined by using a 30 µg vancomycin disc and an inhibition zone diameter of <15 mm.

Table 3. Antibiogram of Enterococcus spp. causing bacteraemia in Northeast Thailand.

| Antibiotic category | Antibiotic agents | CAB | HCAB | HAB | P values |
|---------------------|-------------------------------|------------------|-----------------|------------------|----------|
| | | (n=176 patients) | (n=49 patients) | (n=117 patients) | |
| Aminoglycosides | Gentamicin (high level) | 35/153 (23%) | 24/45 (53%) | 63/101 (62%) | <0.001 |
| Streptomycin | Streptomycin (high level) | NA | NA | NA | - |
| Carbapenems* | Imipenem | NA | NA | NA | - |
| | Meropenem | 1/1 (100%) | NA | 3/5 (60%) | >0.99 |
| | Doripenem | NA | NA | NA | - |
| Fluoroquinolones | Ciprofloxacin | 37/44 (84%) | 9/10 (90%) | 31/37 (84%) | >0.99 |
| | Levofloxacin | 5/18 (28%) | 1/6 (17%) | 11/15 (73%) | 0.01 |
| | Moxifloxacin | NA | NA | NA | - |
| Glycopeptides | Vancomycin | 9/176 (5%) | 0/49 (0%) | 6/113 (5%) | 0.27 |
| | Teicoplanin | 0/11 (0%) | 0/4 (0%) | 0/10 (0%) | - |
| Glycylcyclines | Tigecycline | NA | NA | NA | - |
| Lipopeptides | Daptomycin | NA | NA | NA | - |
| Oxazolidinones | Linezolid | 0/8 (0%) | 0/2 (0%) | 0/4 (0%) | - |
| Penicillins | Ampicillin | 20/134 (15%) | 6/37 (16%) | 34/81 (42%) | <0.001 |
| Streptogramins* | Quinupristin- dalfopristin | NA | NA | NA | - |
| Tetracycline | Doxycycline | NA | NA | NA | - |
| | Minocycline | NA | NA | NA | - |
| MDR | | 0/176 (0%) | 0/49 (0%) | 4/117 (3%) | 0.02 |

NOTE: Data are number of isolates demonstrating non-susceptible to the antimicrobial over the total number of isolates tested (%). CAB=Community-acquired bacteraemia, HCAB=Healthcare-associated bacteraemia, HAB= Hospital-acquired bacteraemia, and NA=Not available. The first isolate of each patient was used. MDR: non-susceptible to ≥1 agent in ≥3 antimicrobial categories.
*Intrinsic resistance in *E. faecium* against carbapenems and in *E. faecalis* against streptogramins. When a species has intrinsic resistance to an antimicrobial category, that category is removed prior to applying the criteria for the MDR definition and is not counted when calculating the number of categories to which the bacterial isolate is non-susceptible.

Table 4. Antibiogram of *E. coli* causing bacteraemia in Northeast Thailand.

| Antibiotic category | Antibiotic agents | CAB (n= 3382 patients) | HCAB (n= 494 patients) | HAB (n= 403 patients) | P values |
|--|---|---------------------------|---------------------------|--------------------------|-------------|
| Aminoglycosides | Gentamicin | 559/3346 (17%) | 166/484 (34%) | 178/398 (45%) | <0.001 |
| | Tobramycin | NA | NA | NA | - |
| | Amikacin | 72/2685 (3%) | 26/397 (7%) | 32/326 (10%) | <0.001 |
| | Netilmicin | 68/1394 (5%) | 25/259 (10%) | 42/254 (17%) | <0.001 |
| Anti-MRSA cephalosporins | Ceftaroline | NA | NA | NA | - |
| Antipseudomonal | Ticarcillin- | NA | NA | NA | - |
| penicillins + β lactamase inhibitors | clauvanic acid Piperacillin- tazobactam | 23/511 (5%) | 10/103 (10%) | 15/89 (17%) | <0.001 |
| Carbapenems | Ertapenem | 4/1325 (<1%) | 1/235 (<1%) | 4/205 (2%) | 0.02 |
| | Imipenem | 3/2449 (<1%) | 0/386 (0%) | 3/344 (1%) | 0.04 |
| | Meropenem | 0/1988 (0%) | 1/314 (<1%) | 1/244 (<1%) | 0.05 |
| Non-extended | Cefazolin | 468/1095 (43%) | 115/174 (66%) | 80/102 (78%) | <0.001 |
| spectrum cephalosporins | Cefuroxime | 219/1438 (15%) | 96/226 (42%) | 102/202 (50%) | <0.001 |
| Extended- | Cefotaxime | 501/3076 (16%) | 199/455 (44%) | 185/361 (51%) | <0.001 |
| spectrum | Ceftazidime | 392/3020 (13%) | 165/446 (37%) | 164/351 (47%) | <0.001 |
| cephalosporins | Cefepime | 30/293 (10%) | 12/42 (29%) | 18/53 (34%) | <0.001 |
| Cephamycins | Cefoxitin | 36/1200 (3%) | 16/215 (7%) | 16/195 (8%) | <0.001 |
| | Cefotetan | NA | NA | NA | - |
| Fluoroquinolones | Ciprofloxacin | 728/3000 (24%) | 221/452 (49%) | 171/384 (45%) | <0.001 |
| Folate pathway inhibitors | Trimethoprim- sulphamethoxazol e | 1738/3007 (58%) | 294/442 (67%) | 225/350 (64%) | <0.001 |
| Glycylcyclines | Tigecycline | 0/7 (0%) | NA | 0/1 (0%) | - |
| Monobactams | Aztreonam | NA | NA | NA | - |
| Penicillins | Ampicillin | 2246/2843 (79%) | 371/420 (88%) | 301/342 (88%) | <0.001 |
| Penicillins + β lactamase | Amoxicillin- clavulanic acid | 790/3074 (26%) | 191/463 (41%) | 158/373 (42%) | <0.001 |
| inhibitors | Ampicillin- sulbactam | 83/296 (28%) | 18/48 (38%) | 12/25 (48%) | 0.06 |
| Phenicols | Chloramphenicol | 14/63 (22%) | 1/4 (25%) | 3/5 (60%) | 0.14 |
| Phosphonic acids | Fosfomycin | NA | NA | NA | - |
| Polymyxins | Colistin * | 2/34 (6%) | 0/6 (0%) | 1/6 (17%) | 0.61 |
| MDR | | 1177/3382 (35%) | 288/494 (58%) | 252/403 (63%) | <0.001 |

NOTE: Data are number of isolates demonstrating non-susceptible to the antimicrobial over the total number of isolates tested (%). CAB=Community-acquired bacteraemia, HCAB=Healthcare-associated

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- bacteraemia, HAB= Hospital-acquired bacteraemia, and NA=Not available. The first isolate of each
- 525 patient was used. MDR: non-susceptible to ≥1 agent in ≥3 antimicrobial categories.
- * Defined by using an inhibition zone of <11 mm.

528 Table 5. Antibiogram of *K. pneumoniae* causing bacteraemia in Northeast Thailand.

| Antibiotic category | Antibiotic agents | CAB (n= 1010 patients) | HCAB (n= 196 patients) | HAB (n=455 patients) | P values |
|--|---|------------------------|---------------------------|-------------------------|-------------|
| Aminoglycosides | Gentamicin | 94/999 (9%) | 53/193 (27%) | 265/444 (60%) | <0.001 |
| | Tobramycin | NA | NA | NA | - |
| | Amikacin | 17/815 (2%) | 12/157 (8%) | 109/398 (27%) | <0.001 |
| | Netilmicin | 20/450 (4%) | 23/112 (21%) | 124/320 (39%) | <0.001 |
| Anti-MRSA cephalosporins | Ceftaroline | NA | NA | NA | - |
| Antipseudomonal | Ticarcillin- | NA | NA | NA | - |
| penicillins + β lactamase inhibitors | clauvanic acid Piperacillin- tazobactam | 24/166 (14%) | 14/32 (44%) | 73/121 (60%) | <0.001 |
| Carbapenems | Ertapenem | 2/432 (0%) | 1/100 (1%) | 5/264 (2%) | 0.17 |
| | Imipenem | 1/778 (0%) | 1/164 (1%) | 2/408 (0%) | 0.24 |
| | Meropenem | 0/583 (0%) | 1/113 (1%) | 2/317 (1%) | 0.10 |
| Non-extended | Cefazolin | 76/319 (24%) | 30/60 (50%) | 101/127 (80%) | <0.001 |
| spectrum cephalosporins | Cefuroxime | 81/478 (17%) | 35/98 (36%) | 161/231 (70%) | <0.001 |
| Extended- | Cefotaxime | 146/902 (16%) | 71/173 (41%) | 298/424 (70%) | <0.001 |
| spectrum cephalosporins | Ceftazidime | 124/927 (13%) | 63/176 (36%) | 295/430 (69%) | <0.001 |
| серниюзроння | Cefepime | 5/100 (5%) | 8/22 (36%) | 25/51 (49%) | <0.001 |
| Cephamycins | Cefoxitin | 15/396 (4%) | 10/95 (11%) | 14/230 (6%) | 0.03 |
| | Cefotetan | NA | NA | NA | - |
| Fluoroquinolones | Ciprofloxacin | 143/894 (16%) | 66/176 (38%) | 187/430 (43%) | <0.001 |
| Folate pathway inhibitors | Trimethoprim- sulphamethoxazol e | 198/876 (23%) | 69/171 (40%) | 219/407 (54%) | <0.001 |
| Glycylcyclines | Tigecycline | NA | NA | NA | - |
| Monobactams | Aztreonam | NA | NA | NA | - |
| Penicillins + β lactamase | Amoxicillin- clavulanic acid | 131/945 (14%) | 68/183 (37%) | 291/443 (66%) | <0.001 |
| inhibitors | Ampicillin- sulbactam | 20/105 (19%) | 9/17 (53%) | 23/38 (61%) | <0.001 |
| Phenicols | Chloramphenicol | 4/19 (21%) | 0/2 (0%) | 0/3 (0%) | >0.99 |
| Phosphonic acids | Fosfomycin | NA | NA | NA | - |
| Polymyxins | Colistin * | 0/6 (0%) | 0/2 (0%) | 0/5 (0%) | - |
| MDR | | 146/1010 (14%) | 71/196 (36%) | 301/455 (66%) | <0.001 |

NOTE: Data are number of isolates demonstrating non-susceptible to the antimicrobial over the total number of isolates tested (%). CAB=Community-acquired bacteraemia, HCAB=Healthcare-associated

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bacteraemia, HAB= Hospital-acquired bacteraemia, and NA=Not available. The first isolate of each
 patient was used. MDR: non-susceptible to ≥1 agent in ≥3 antimicrobial categories.
 * Defined by using an inhibition zone of <11 mm.
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Table 6. Antibiogram of *P. aeruginosa* causing bacteraemia in Northeast Thailand.

| Antibiotic category | Antibiotic | CAB | HCAB | НАВ | P values |
|------------------------------|----------------|------------------|------------------|------------------|----------|
| | agents | (n=286 patients) | (n=103 patients) | (n=179 patients) | |
| Aminoglycosides | Gentamicin | 29/235 (12%) | 13/88 (15%) | 60/140 (43%) | <0.001 |
| | Tobramycin | NA | NA | NA | - |
| | Amikacin | 27/284 (10%) | 13/100 (13%) | 48/177 (27%) | <0.001 |
| | Netilmicin | 8/155 (5%) | 5/67 (7%) | 34/120 (28%) | <0.001 |
| Antipseudomonal | Imipenem | 14/238 (6%) | 6/86 (7%) | 37/154 (24%) | <0.001 |
| carbapenems | Meropenem | 9/163 (6%) | 8/73 (11%) | 24/125 (19%) | 0.001 |
| | Doripenem | 2/17 (12%) | 0/3 (0%) | 2/2 (100%) | 0.04 |
| Antipseudomonal | Ceftazidime | 29/280 (10%) | 16/103 (16%) | 68/179 (38%) | <0.001 |
| cephalosporins | Cefepime | 2/36 (6%) | 2/18 (11%) | 10/28 (36%) | 0.01 |
| Antipseudomonal | Ciprofloxacin | 24/275 (9%) | 12/101 (12%) | 39/169 (23%) | <0.001 |
| fluoroquinolones | Levofloxacin | 0/1 (0%) | 1/1 (100%) | 1/1 (100%) | >0.99 |
| Antipseudo | Ticarcillin- | NA | NA | NA | - |
| $monal\ penicillins + \beta$ | clauvanic acid | | | | |
| lactamase inhibitors | Piperacillin- | 8/85 (9%) | 6/38 (16%) | 8/46 (17%) | 0.37 |
| | tazobactam | | | | |
| Monobactams | Aztreonam | NA | NA | NA | - |
| Phosphonic acids | Fosfomycin | 1/1 (100%) | NA | NA | - |
| Polymyxins | Colistin | 0/7 (0%) | 0/3 (0%) | 1/7 (14%) | >0.99 |
| | Polymyxin B | NA | NA | NA | - |
| MDR | | 13/286 (5%) | 10/103 (10%) | 45/179 (25%) | <0.001 |

NOTE: Data are number of isolates demonstrating non-susceptible to the antimicrobial over the total number of isolates tested (%). CAB=Community-acquired bacteraemia, HCAB=Healthcare-associated bacteraemia, HAB= Hospital-acquired bacteraemia, and NA=Not available. The first isolate of each patient was used. MDR: non-susceptible to ≥ 1 agent in ≥ 3 antimicrobial categories.

Table 7. Antibiogram of *Acinetobacter* spp. causing bacteraemia in Northeast Thailand.

| Austilatia aasta aasu | Antibiatia aganta | CAD | LICAD | LIAD | |
|-----------------------|-------------------|------------------|------------------|------------------|--------|
| Antibiotic category | Antibiotic agents | CAB | HCAB | HAB | P |
| | | (n=449 patients) | (n=115 patients) | (n=501 patients) | values |
| Aminoglycosides | Gentamicin | 112/390 (29%) | 45/105 (43%) | 310/455 (68%) | <0.001 |
| | Tobramycin | NA | NA | NA | - |
| | Amikacin | 123/442 (28%) | 45/112 (40%) | 310/495 (63%) | <0.001 |
| | Netilmicin | 44/203 (22%) | 24/64 (38%) | 224/381 (59%) | <0.001 |
| Antipseudomonal | Imipenem | 87/397 (22%) | 37/102 (36%) | 293/459 (64%) | <0.001 |
| carbapenems | Meropenem | 65/284 (23%) | 32/81 (40%) | 229/348 (66%) | <0.001 |
| | Doripenem | 16/45 (36%) | 9/10 (90%) | 6/7 (86%) | 0.001 |
| Antipseudomonal | Ciprofloxacin | 84/413 (20%) | 53/106 (50%) | 322/481 (67%) | <0.001 |
| fluoroquinolones | Levofloxacin | 2/5 (40%) | 2/2 (100%) | 8/9 (89%) | 0.11 |
| Antipseudomonal | Ticarcillin- | NA | NA | NA | - |
| penicillins + β | clauvanic acid | | | | |
| lactamase | Piperacillin- | 22/98 (22%) | 13/28 (46%) | 74/106 (70%) | <0.001 |
| inhibitors | tazobactam | | | | |
| Extended- | Cefotaxime | 242/291 (83%) | 89/94 (95%) | 407/420 (97%) | <0.001 |
| spectrum | Ceftazidime | 133/448 (30%) | 61/114 (54%) | 377/500 (75%) | <0.001 |
| cephalosporins | Cefepime | 18/53 (34%) | 10/22 (45%) | 95/133 (71%) | <0.001 |
| Folate pathway | Trimethopri- | 119/356 (33%) | 55/99 (56%) | 333/435 (77%) | <0.001 |
| inhibitor | sulphamethoxazole | | | | |
| Penicillins + β | Ampicillin- | 43/134 (32%) | 16/29 (55%) | 79/115 (69%) | <0.001 |
| lactamase | sulbactam | | | | |
| inhibitors | | | | | |
| Polymyxins | Colistin * | 2/16 (13%) | 0/14 (0%) | 0/33 (0%) | 0.11 |
| | Polymyxin B | NA | NA | NA | - |
| Tetracyclines | Tetracycline | NA | NA | NA | - |
| | Doxycycline | NA | NA | NA | - |
| | Minocycline | NA | NA | NA | - |
| MDR | | 125/449 (28%) | 58/115 (50%) | 374/501 (75%) | <0.002 |

NOTE: Data are number of isolates demonstrating non-susceptible to the antimicrobial over the total number of isolates tested (%). CAB=Community-acquired bacteraemia, HCAB=Healthcare-associated bacteraemia, HAB= Hospital-acquired bacteraemia, and NA=Not available. The first isolate of each patient was used. MDR: non-susceptible to ≥ 1 agent in ≥ 3 antimicrobial categories.

^{*} Defined by using an inhibition zone of <11 mm.

Table 8. Estimates of mortality attributable to multidrug-resistance (MDR) in hospital-acquired infection (HAI) in Thailand

| Pathogens | No of patients ^a | Estimated | Estimated mortality if the | Estimated excess mortality caused |
|----------------------------|-----------------------------|----------------------------|--------------------------------|-----------------------------------|
| | | mortality (%) ^b | infections were caused by non- | by MDR (%) ^{b, c} |
| | | | MDR organisms (%) b, c | |
| MDR Staphylococcus aureus | 18,725 | 8,262 (44%) | 5,463 (29%) | 2,799 (15%) |
| MDR Escherichia coli | 11,116 | 2,163 (19%) | 1,566 (14%) | 597 (5%) |
| MDR Klebsiella pneumoniae | 15,239 | 5,267 (35%) | 4,979 (33%) | 288 (2%) |
| MDR Pseudomonas aeruginosa | 6,118 | 3,966 (65%) | 3,696 (60%) | 270 (4%) |
| MDR Acinetobacter spp | 36,553 | 25,551 (70%) | 10,383 (28%) | 15,168 (41%) |
| Total | 87,751 | 45,209 (52%) | 26,087 (30%) | 19,122 (22%) |

^a Cumulative incidence of antimicrobial resistant HAI in Thailand 2010 estimated by Pumart et al. ¹⁵

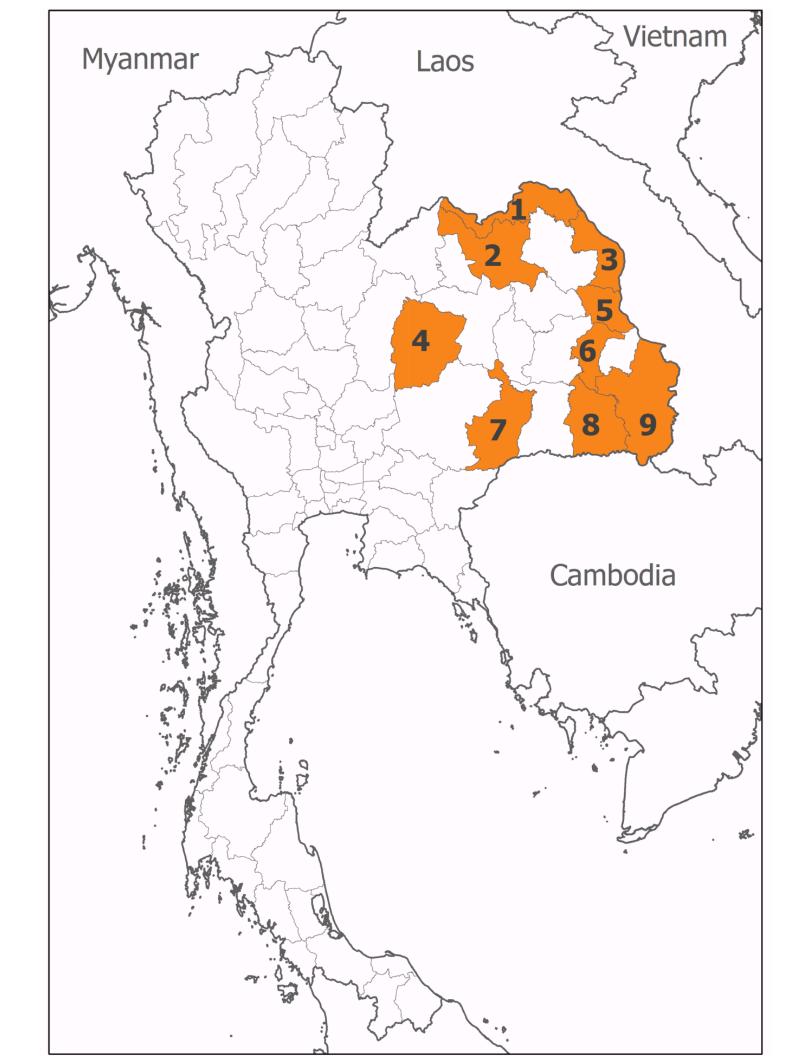
551

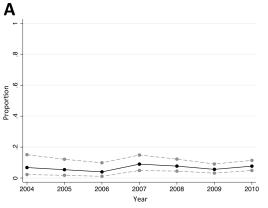
552

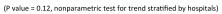
553

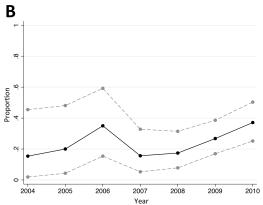
^b All parameters used to estimate the mortality and excess mortality are shown in Supplementary File 2.

⁵⁵⁴ ^c Excess mortality caused by MDR (mortality attributable to MDR) was defined as the difference in mortality of patients with MDR infection and their mortality if they were infected with non-MDR infections.

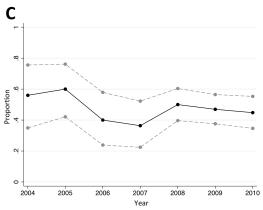




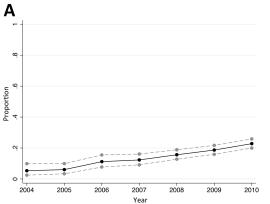




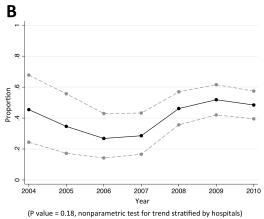
(P value = 0.77, nonparametric test for trend stratified by hospitals)

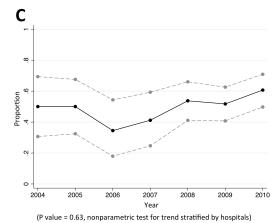


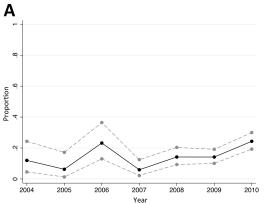
(P value = 0.61, nonparametric test for trend stratified by hospitals)



(P value = 0.04, nonparametric test for trend stratified by hospitals)







(P value = 0.04, nonparametric test for trend stratified by hospitals)

