



eLife's transparent reporting form

We encourage authors to provide detailed information *within their submission* to facilitate the interpretation and replication of experiments. Authors can upload supporting documentation to indicate the use of appropriate reporting guidelines for health-related research (see [EQUATOR Network](#)), life science research (see the [BioSharing Information Resource](#)), or the [ARRIVE guidelines](#) for reporting work involving animal research. Where applicable, authors should refer to any relevant reporting standards documents in this form.

If you have any questions, please consult our Journal Policies and/or contact us: editorial@elifesciences.org.

Sample-size estimation

- You should state whether an appropriate sample size was computed when the study was being designed
- You should state the statistical method of sample size computation and any required assumptions
- If no explicit power analysis was used, you should describe how you decided what sample (replicate) size (number) to use

Please outline where this information can be found within the submission (e.g., sections or figure legends), or explain why this information doesn't apply to your submission:

Immunoprecipitation and subsequent mass spectrometry analysis has been performed using a starting material of HEK-293 derived from two confluent 15 cm plates. For the primary hepatocyte experiment livers from 2-3 animals were analyzed for each biological replicate.

Replicates

- You should report how often each experiment was performed
- You should include a definition of biological versus technical replication
- The data obtained should be provided and sufficient information should be provided to indicate the number of independent biological and/or technical replicates
- If you encountered any outliers, you should describe how these were handled
- Criteria for exclusion/inclusion of data should be clearly stated
- High-throughput sequence data should be uploaded before submission, with a private link for reviewers provided (these are available from both GEO and ArrayExpress)

Please outline where this information can be found within the submission (e.g., sections or figure legends), or explain why this information doesn't apply to your submission:

The number of replicates for the individual experiments (1) immunoprecipitation data, (2) qRT-PCR data, (3) MTT assays, (4) relative protein abundance determined by immunoblotting has been $n \geq 3$ unless otherwise stated in the respective Figure Legend. Relative protein abundance of primary murine hepatocytes has been determined from two biological replicates and is stated in the respective Figure Legend.

Statistical reporting



- Statistical analysis methods should be described and justified
- Raw data should be presented in figures whenever informative to do so (typically when N per group is less than 10)
- For each experiment, you should identify the statistical tests used, exact values of N, definitions of center, methods of multiple test correction, and dispersion and precision measures (e.g., mean, median, SD, SEM, confidence intervals; and, for the major substantive results, a measure of effect size (e.g., Pearson's r, Cohen's d)
- Report exact p-values wherever possible alongside the summary statistics and 95% confidence intervals. These should be reported for all key questions and not only when the p-value is less than 0.05.

Please outline where this information can be found within the submission (e.g., sections or figure legends), or explain why this information doesn't apply to your submission:

All data are presented as mean \pm s.e.m. and subjected to statistical analysis using the GraphPad Prism software. Statistical significance was assessed for (1) relative protein abundance determined by immunoblotting, (2) mRNA expression determined by qRT-PCR and (3) proliferation rates determined by MTT assays using a paired Student's t-test with a confidence level of 95%. Experiments have been performed $n \geq 3$ unless otherwise indicated in the respective Figure Legend. For the AP-MS experiments the raw data (spectral counts) was analyzed using the SAINT (Choi et al., 2011) scoring tool implemented within the Contaminant Repository for Affinity Purification ([CRAPome](#)) platform (Mellacheruvu et al., 2013). For calculation of SAINT scores, we followed workflow 3 using the following parameters: FC-A; user, default, FC-B; all controls, stringent. For enrichment analysis, the peptide spectrum matches for each protein were ranked by fold change versus the respective control immunoprecipitation. The sample size for each immunoprecipitation was $n \geq 2$ unless otherwise stated.

(For large datasets, or papers with a very large number of statistical tests, you may upload a single table file with tests, Ns, etc., with reference to sections in the manuscript.)

Group allocation

- Indicate how samples were allocated into experimental groups (in the case of clinical studies, please specify allocation to treatment method); if randomization was used, please also state if restricted randomization was applied
- Indicate if masking was used during group allocation, data collection and/or data analysis

Please outline where this information can be found within the submission (e.g., sections or figure legends), or explain why this information doesn't apply to your submission:

Additional data files ("source data")

- We encourage you to upload relevant additional data files, such as numerical data that are represented as a graph in a figure, or as a summary table
- Where provided, these should be in the most useful format, and they can be uploaded as "Source data" files linked to a main figure or table
- Include model definition files including the full list of parameters used
- Include code used for data analysis (e.g., R, MatLab)



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- Avoid stating that data files are “available upon request”

Please indicate the figures or tables for which source data files have been provided:

Source data contain the complete mass spectrometry lists analyzed by SAINT or by spectral peptide matches as stated in the Material & Methods.