

Supplementary File 5: Full description of all the cell-cell interaction given in main text. All interactions are referenced in Appendix 1. Melanocytes, xanthophores, loose iridophores, dense iridophores and xanthoblasts are denoted M , X , I^l , I^d , X^b respectively. C1, C2 stands for cell 1, cell 2, where C1 = X , X^b is the target cell and cell 2 is the signalling cell with corresponding signal range (R) that is either short (S), up to 0.04mm, or long (L), 0.12mm. This signal generates action (A) of cell type C1 of: movement (M), differentiation (D), proliferation (P) or survival (S) of type (T). For action (D) or (S), type (T) is denoted '+' if the resultant action is promotion of action A and '-' if the resultant action is inhibition of action (A). For action (M), type (T) is denoted '+' if the resultant action is attraction towards cell type C1 and '-' if the resultant action is repulsion away from cell type C1. *Melanocytes can also differentiate randomly, independent of any other cell type.

| C1 | C2 | A | R | T | Basis of assumption | Details | Ref. |
|-------|-------|---|---|---|--|--|--------|
| I^l | M | S | - | - | Adult <i>pfe</i> , <i>shd</i> , <i>nac</i> and WT fish. | I^l are typically spaced at a distance from each other. | [1] |
| X | M | S | + | + | WT fish development. | Arrival of I^d leads to compaction of X overlaying this region. | [2] |
| I^d | X | M | S | + | Adult <i>pfe</i> , <i>shd</i> , <i>nac</i> and WT fish. | X and I^d are highly associated with each other. | [1] |
| M | M | S | S | - | Long term imaging of WT fish | M appear to migrate to dense I^d free zones. | [3] |
| I^d | M | S | S | - | Adult <i>pfe</i> , <i>shd</i> , <i>nac</i> and WT fish. | I^d are typically not associated with M . | [1] |
| X | M | S | S | + | Cultured WT M and X . | <i>In vitro</i> , X extended pseudopodia towards M which they then followed towards M . | [4] |
| M | X | S | S | - | Regeneration experiments performed <i>in vivo</i> on WT fish | When X in close proximity to M in the pale interstripes, M move out of interstripe, versus when the X in close proximity to M are ablated. Then M are less likely to leave the interstripe region. | [5] |
| M | X^b | M | L | + | Long term imaging of WT fish | X^b in stripes exhibit airnemes which attach to M activating Delta-Notch signalling and resulting in the consolidation of M into stripes. | [6, 7] |
| I^d | M | D | S | - | Observations of WT fish. | M typically differentiate in I^d free zones. | [3] |
| I^d | M | D | L | + | Observations of <i>pfe</i> mutants and WT | M differentiation is associated with one stripe width distance away from the central strip of I^d . | [1] |
| X | M | D | S | - | Regeneration experiments <i>in vivo</i> on WT fish. | M and X compete to differentiate in the S range. | [4] |
| X | M | S | L | + | Regeneration experiments <i>in vivo</i> on WT fish. | Increased M death when X ablated from neighbouring pale interstripes. | [4, 8] |
| X | M | S | S | - | Regeneration experiments <i>in vivo</i> on WT fish. | When X in close proximity to M , some M die versus when X in close proximity to M are ablated, no M die. | [5, 9] |
| X | M | D | L | + | Regeneration experiments <i>in vivo</i> on WT fish. | M regeneration proportional to neighbouring pale interstripe X number. | [4, 8] |

| C1 | C2 | A | R | T | Basis of assumption | Details | Ref. |
|-------|-------|---|---|---|---|---|------------|
| M | M | D | L | - | Regeneration experiments <i>in vivo</i> on WT fish. | M regeneration negatively proportional to M at half a stripe width distance. | [4] |
| M | M | S | L | - | Regeneration experiments <i>in vivo</i> on WT fish. | M regeneration negatively proportional to M at half a stripe width distance (when X ablated from neighbouring inter-stripes). | [4] |
| I^l | M | S | S | + | Observations of <i>pfe</i> fish. | We predict that loose S-iridophores promote M survival as this would account for the aggregates of M s observed in <i>pfe</i> . | [10, 1] |
| * | M | D | | | Observation of double mutant <i>nac;pfe</i> fish. | <i>nac;pfe</i> are unable to generate X s or iridophores, yet readily generate M . | [1] |
| X^b | I^d | D | S | + | S-iridophore expression studies. | I^d express xanthogenic Colony stimulating factor-1 (Csf1) which is essential for X^b differentiation. | [3, 1, 11] |
| X^b | X | P | S | + | Regeneration experiments <i>in vivo</i> on WT fish. | M and X compete to differentiate in the short range. | [4] |
| X^b | M | D | S | - | Regeneration experiments <i>in vivo</i> on WT fish. | M and X compete to differentiate in the short range. | [4] |
| X | I^d | P | S | + | S-iridophore expression studies. | I^d express xanthogenic Colony stimulating factor-1 (Csf1) which is essential for X proliferation and survival. | [3, 1, 11] |
| X | M | P | S | - | Regeneration experiments <i>in vivo</i> on WT fish. | M and X compete to differentiate in the short range. | [4] |
| X | X | P | S | + | Regeneration experiments <i>in vivo</i> on WT fish. | M and X compete to differentiate in the short range. | [4] |
| X | | P | | | Clonal analyses of X lineage. | X and X^b observed to proliferate at a rate of approximately 1/week. | [2] |
| X^b | | P | | | Clonal analyses of X lineage. | X and X^b observed to proliferate at a rate of approximately 1/week. | [2] |

1 **References**

- 2 [1] H. G. Frohnhof, J. Krauss, H.-M. Maischein, and C. Nüsslein-Volhard.
3 Iridophores and their interactions with other chromatophores are required
4 for stripe formation in zebrafish. *Development*, 140(14):2997–3007, 2013.
- 5 [2] P. Mahalwar, B. Walderich, A. P. Singh, and C. Nüsslein-Volhard. Local
6 reorganization of xanthophores fine-tunes and colors the striped pattern of
7 zebrafish. *Science.*, 345(6202):1362–4, 2014.
- 8 [3] L. B. Patterson and D. M. Parichy. Interactions with Iridophores and
9 the Tissue Environment Required for Patterning Melanophores and Xan-
10 thophores during Zebrafish Adult Pigment Stripe Formation. *PLoS Genet.*,
11 9(5), 2013.
- 12 [4] A. Nakamasu, G. Takahashi, A. Kanbe, and S. Kondo. Interactions between
13 zebrafish pigment cells. *PNAS*, 106(21):8429–8434, 2009.
- 14 [5] G. Takahashi and S. Kondo. Melanophores in the stripes of adult zebrafish
15 do not have the nature to gather, but disperse when they have the space
16 to move. *Pigment Cell Melanoma Res.*, 21(6):677–686, 2008.
- 17 [6] D. S. Eom, E. J. Bain, L. B. Patterson, M. E. Grout, and D. M. Parichy.
18 Long-distance communication by specialized cellular projections during pig-
19 ment pattern development and evolution. *eLife*, 4(December 2015):1–25,
20 2015.
- 21 [7] D. S. Eom and D. M. Parichy. A macrophage relay for long-distance signal-
22 ing during postembryonic tissue remodeling. *Science*, 355(6331):1317–1320,
23 2017.
- 24 [8] S. Kondo and M. Watanabe. Black, yellow, or silver: Which one leads skin
25 pattern formation? *Pigment Cell Melanoma Res.*, 28(1):2–4, 2015.
- 26 [9] H. Yamanaka and S. Kondo. In vitro analysis suggests that difference in cell
27 movement during direct interaction can generate various pigment patterns
28 in vivo. *PNAS*, 111(5):1867–1872, 2014.
- 29 [10] C. B. Kimmel, W.W. Ballard, S. R. Kimmel, B. Ullmann, and T. F.
30 Schilling. Stages of embryonic development of the zebrafish. *Dev. Dyn.*,
31 203(3):253–310, 1995.
- 32 [11] B. Walderich, A. P. Singh, P. Mahalwar, and C. Nüsslein-Volhard. Homo-
33 typic cell competition regulates proliferation and tiling of zebrafish pigment
34 cells during colour pattern formation. *Nat. Commun.*, 7:11462, 2016.