

1 **Environmental selection overturns the decay relationship of soil prokaryotic community**
2 **over geographic distance across grassland biotas**

3

4 Biao Zhang¹, Kai Xue^{123*}, Shutong Zhou⁴, Kui Wang⁴, Wenjing Liu¹, Cong Xu⁵, Lizhen Cui⁴,
5 Linfeng Li¹⁶, Qinwei Ran⁴, Zongsong Wang⁴, Ronghai Hu¹, Yanbin Hao³⁴, Xiaoyong Cui³⁴,
6 Yanfen Wang¹³

7

8 ¹College of Resources and Environment, University of Chinese Academy of Sciences, Beijing
9 101408, China,

10 ²Yanshan Earth Critical Zone National Research Station, University of Chinese Academy of
11 Sciences, Beijing 101408, China,

12 ³Key Laboratory of Adaptation and Evolution of Plateau Biota, Chinese Academy of Sciences,
13 Xining 810001, China

14 ⁴College of Life Sciences, University of Chinese Academy of Sciences, Beijing 101408, China,

15 ⁵State Key Laboratory of Remote Sensing Science, Aerospace Information Research Institute,
16 Chinese Academy of Sciences, Beijing 100101, China,

17 ⁶Environmental Futures Research Institute, School of Environment and Science, Griffith
18 University, Nathan QLD 4111, Australia

19

20

21

22

23 **Abstract**

24 Though being fundamental to global diversity distribution, little is known about the geographic
25 pattern of soil microorganisms across different biotas on a large scale. Here, we investigated soil
26 prokaryotic communities from Chinese northern grasslands on a scale up to 4,000 km in both
27 alpine and temperate biotas. Prokaryotic similarities increased over geographic distance after
28 tipping points of 1,760 - 1,920 km, generating a significant U-shape pattern. Such pattern was
29 likely due to decreased disparities in environmental heterogeneity over geographic distance when
30 across biotas, supported by three lines of evidences: 1) prokaryotic similarities still decreased
31 with the environmental distance, 2) environmental selection dominated prokaryotic assembly,
32 and 3) short-term environmental heterogeneity followed the U-shape pattern spatially, especially
33 attributed to dissolved nutrients. In sum, these results demonstrate that environmental selection
34 overwhelmed the geographic “distance” effect when across biotas, overturning the previously
35 well-accepted geographic pattern for microbes on a large scale.

36

37

38

39

40

41

42

43

44

45 **Introduction**

46 To clarify the spatial pattern of biodiversity is one of primary aims in ecology and biogeography
47 (1, 2). In past decades, intensive biogeographic studies have been conducted for
48 macro-organisms, including plants (3-8), insects (9, 10) and vertebrates (11, 12). With the
49 emergence and development of next-generation sequencing, increasing attention has been paid
50 recently to the spatial pattern of microorganisms. The similarity of microbial communities has
51 been observed to decrease over geographic distance as that of macro-organisms does, so-called
52 distance-decay relationship, in different habits (e.g., forest (13, 14) grassland (15), desert (16)
53 and agriculture soils (17-19)) for bacteria (15, 20, 21), archaea (18, 22), fungi (23-26) and
54 specific microbial functional groups (e.g. ammonia-oxidizing archaea, ammonia-oxidizing and
55 sulfate-reducing bacteria (27-30)). To certain extent, the reported distance-decay relationship has
56 been regarded as a principal generalization in nature (31).

57

58 However, no consensus has been reached so far on underlying mechanisms of the distance-decay
59 relationship for soil microbial communities. A few mechanisms for biodiversity maintenance (20)
60 have been proposed to be responsible for such relationship as well, including the environmental
61 heterogeneity, dispersal limitation and stochastic processes (4, 32, 33). Specifically,
62 environmental heterogeneity tends to increase over geographic distance, responsible for the
63 distance-decay relationship (2, 18). Microorganisms have been observed to have the dispersal

64 limitation as macro-organisms do (2, 34, 35), crucial in biodiversity maintenance and evolution
65 (33, 36). Spatial configurations and the nature of landscapes influence the dispersal rate of
66 organisms among sites (32), and communities tend to be more similar in open and
67 topographically homogeneous settings than in heterogeneous landscapes. Moreover,
68 communities are expected to become increasingly different along with geographic distance as
69 their species are sorted according to their niche requirements (34). Under this scenario,
70 dissimilarities among communities parallel to increasing disparities in environmental
71 heterogeneity along with geographic distance. Furthermore, stochastic processes in birth, death,
72 migration, disperse and drift may also contribute to the distance-decay relationship for soil
73 microbial communities (37-39).

74

75 The relative importance of various mechanisms is still unclear (4, 32, 33), likely being scale- and
76 biota-dependent. Environment heterogeneity has been reported to be more important in
77 influencing the spatial distribution of microorganisms at local scales up to hundreds of
78 kilometers (19, 40, 41), while dispersal limitation dominated the distance-decay relationship on
79 larger scales (42, 43). Moreover, different biotas under distinct climate conditions/latitudes may
80 have different turnover rates for community similarity over geographic distance. High
81 temperature was reported to lead to a lower turnover rate for forest soil microbial community
82 (30), likely due to accelerated biochemical reactions and increased ecological niche breadth (44).
83 Community similarity was observed to decline faster at high than low latitudes on large scales,
84 while the turnover rate was higher at low latitudes on small scales (32). However, most previous

85 studies were conducted locally or regionally within a single biota, region or climate type, and
86 surveys for the geographic pattern of soil microorganisms across different biotas are still lacking,
87 which are essential to understand the spatial pattern of microbial communities beyond these
88 scales.

89

90 Here, we collected grassland soil samples from two biotas of alpine and temperate grasslands
91 with distinct hydrothermal conditions to investigate the spatial pattern of prokaryotic
92 communities and underlying mechanisms. A total of 258 samples were collected from the top-
93 (0-5 cm in depth) and subsoils (5-20 cm in depth) in both the alpine biota on the Qinghai-Tibet
94 Plateau and the temperate biota on the Inner Mongolia Plateau, China, on a scale up to 4,000 km.
95 Our objectives were to test the following hypotheses: (I) soil prokaryotic community similarity
96 would decrease over geographic distance within and across biotas; (II) the turnover rate of soil
97 prokaryotic community similarity over geographic distance would be higher in the temperate
98 than alpine biota, as temperate biota is with a wider temperature range; (III) the turnover rate of
99 soil prokaryotic community similarity over geographic distance would be lower in top- than
100 subsoil, since the subsoil may be less dynamic as not affected by environmental factors like UV
101 and wind.

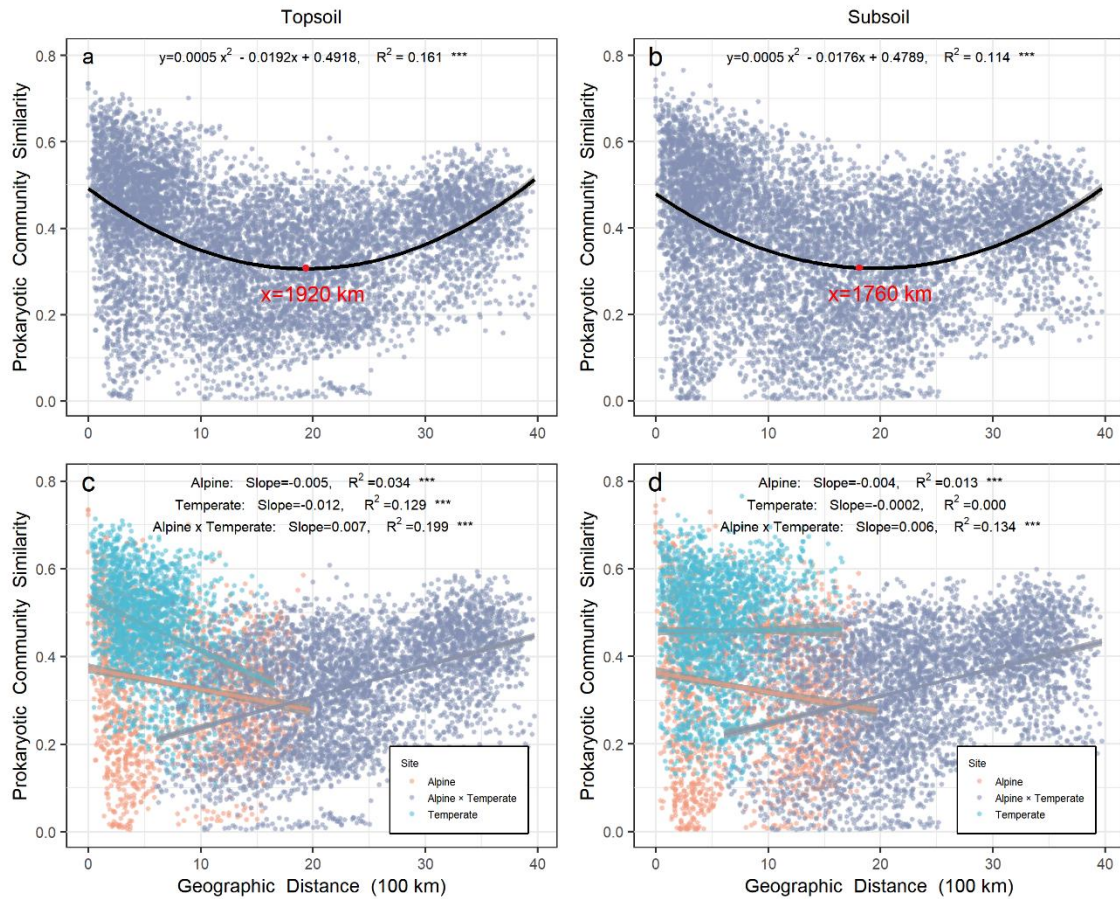
102

103 **Results**

104 **Prokaryotic and plant community similarity over geographic distance**

105 A total of 11,063 OTUs were detected from all 258 grassland soil samples in both alpine and

106 temperate biotas (Figure 1 - figure supplement 1). A significant ($P < 0.001$) binomial relationship
107 (U shape) was observed for the prokaryotic community over geographic distance in top- ($R^2 =$
108 0.161) or subsoil ($R^2 = 0.114$) from all sites on a scale up to 4,000 km. Specifically, the
109 prokaryotic community similarity in topsoil decreased over geographic distance on a scale of <
110 1,920 km mostly within either temperate or alpine biota, but increased after this tipping point
111 when across biotas (in pairwise sites between alpine and temperate biotas). Similarly, the
112 prokaryotic community similarity in subsoil decreased over geographic distance on a scale of <
113 1,760 km mostly within either temperate or alpine biota, but increased after this tipping point
114 when across biotas (Figure 1). When across biotas, the prokaryotic community similarity
115 increased significantly over geographic distance with similar slopes (turnover rates) in top-
116 (slope = 0.007, $R^2 = 0.199$) and subsoil (slope = 0.006, $R^2 = 0.134$).



117

118 **Figure 1 Relationship between prokaryotic community similarity over geographic distance**

119 **in Chinese northern grassland. Panel a and c represent the prokaryotic community in topsoil,**

120 **while panel b and d represent the prokaryotic community in subsoil. Orange and light blue points**

121 **represent pairwise sites within the alpine and temperate biota, respectively. Grey points represent**

122 **pairwise sites between the alpine biota cross temperate biota. Grey shades stand for 95%**

123 **confidence interval.**

124

125 Within the alpine biota, a valid ($P < 0.001$) distance-decay relationship was observed for the

126 prokaryotic community over geographic distance in top- ($R^2 = 0.034$) or subsoil ($R^2 = 0.013$).

127 However, within the temperate biota, the distance-decay relationship for the prokaryotic

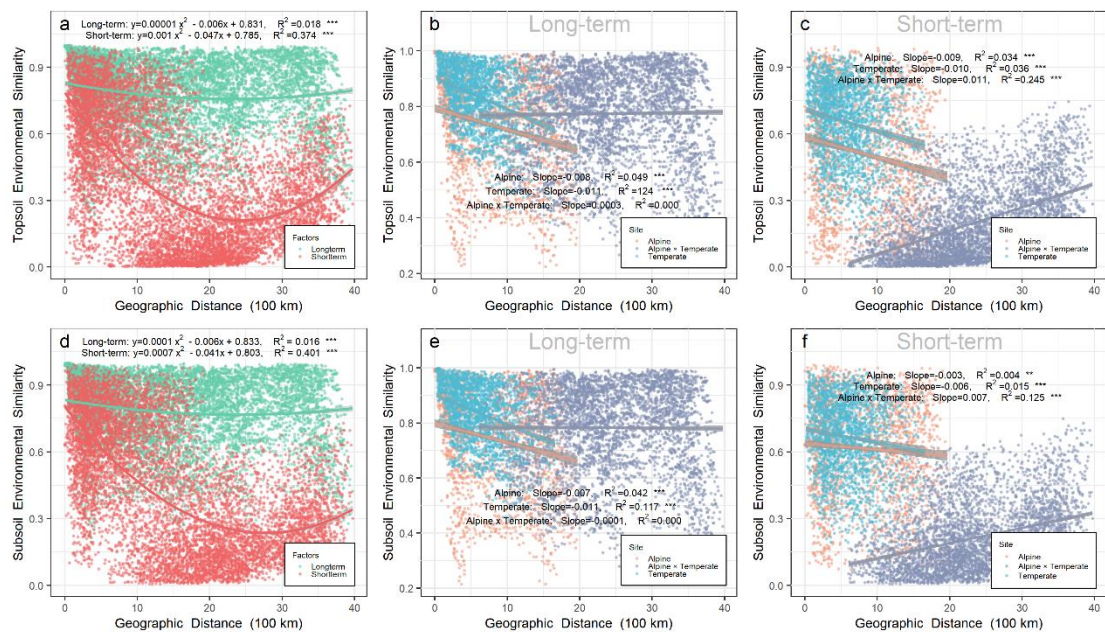
128 community occurred only in topsoil ($R^2 = 0.129$, $P < 0.001$), while no relationship was observed
 129 in subsoil. In topsoil, prokaryotic community similarity had a higher turnover rate in the
 130 temperate (- 0.012) than alpine biota (- 0.005).

131

132 Similar to the prokaryotic community, the plant community also exhibited a significant U-shape
 133 relationship ($R^2 = 0.071$, $P < 0.001$) for its similarity over geographic distance in all sites on a
 134 scale up to 4,000 km, with a tipping point of 1,858 km (Figure 1 - figure supplement 2a). A
 135 significant ($P < 0.001$) distance-decay relationship for plant community was observed within the
 136 alpine (Figure 1 - figure supplement 2b, $R^2 = 0.015$, $P < 0.001$) or temperate biota (Figure 1 -
 137 figure supplement 2b, $R^2 = 0.005$, $P < 0.01$).

138

139 **Prokaryotic community similarity over environmental distance**



140

141 **Figure 2 Relationship between environmental similarity over geographic distance in**

142 **Chinese northern grassland.** Panel a, b and c represent the environmental similarity in topsoil,
143 while panel d, e and f represent the environmental similarity in subsoil. The environmental
144 similarity was calculated by Bray-Curtis index based on relatively long-term (b, e & green points
145 in a and d) or short-term variables (c, f & red points in a and d). Relatively long-term
146 environmental variables included mean annual precipitation (MAP), mean annual temperature
147 (MAT), pH, soil organic carbon (SOC), soil total nitrogen (TN) and soil total phosphorus (TP).
148 Relatively short-term environmental variables included soil water content (SWC), soil available
149 phosphorus (AP), dissolved organic carbon (DOC), dissolved organic nitrogen (DON), NH_4^+ ,
150 and NO_3^- . Orange, grey, and light blue points represent pairwise sites within the alpine biota,
151 within temperate biota, and alpine biota cross temperate biota, respectively. Grey shades stand
152 for 95% confidence interval.

153

154 Relatively short-term environmental similarity also exhibited a U-shape pattern over geographic
155 distance in either top- or subsoil from all sites on a scale up to 4,000 km (Figure 2 a and d). In
156 contrast, relatively long-term environmental similarity, much higher than short-term
157 environmental similarity on the same scale, did not change greatly over geographical distance in
158 either top- or subsoil.

159

160 Soil prokaryotic community similarity decreased significantly ($P < 0.001$) in all sites over the
161 relatively long-term (turnover rate = - 0.291 or - 0.278 in top- or subsoil, respectively) or
162 short-term (turnover rate = - 0.193 or - 0.159 in top- or subsoil, respectively) environmental

163 distance (Figure 2 - figure supplement 1 a and e). In the topsoil, the prokaryotic community
 164 similarity decreased significantly ($P < 0.001$) over the relatively long-term (turnover rate = -
 165 0.277, $R^2 = 0.131$) or short-term (turnover rate = - 0.194, $R^2 = 0.135$) environmental distance
 166 within the alpine biota (Figure 2 - figure supplement 1 b), as well as decreased over long-term
 167 (turnover rate = - 0.339, $R^2 = 0.108$) or short-term (turnover rate = - 0.063, $R^2 = 0.011$)
 168 environmental distance within the temperate biota (Figure 2 - figure supplement 1 c). In the
 169 subsoil, the prokaryotic community similarity decreased significantly over the relatively
 170 long-term (turnover rate = - 0.279, $R^2 = 0.104$) or short-term (turnover rate = - 0.215, $R^2 = 0.093$)
 171 environmental distance within the alpine biota (Figure 2 - figure supplement 1 f), while there was
 172 no relationship within the temperate biota (Figure 2 - figure supplement 1 g). Across biotas or in
 173 pairwise sites between the alpine cross temperate biota (Figure 2 - figure supplement 1 d and h),
 174 the prokaryotic community similarity decreased over relatively long-term (turnover rate = -
 175 0.161 or - 0.130 in top- or subsoil, respectively) or short-term (turnover rate = - 0.191 or - 0.175
 176 in top- or subsoil, respectively) environmental distance.

177

178 **Table 1** Partial Mantel test for relationship between prokaryotic community similarity and
 179 relatively long-term environmental variables, relatively short-term environmental variables and
 180 geographic variables across biotas.

	Alpine × Temperate			
	Topsoil		Subsoil	
	r	p	r	p
MAP	0.060	0.101	0.047	0.165
MAT	0.000	0.498	-0.021	0.625
pH	-0.001	0.516	-0.004	0.542
SOC	0.231	0.001	0.120	0.010

TN	0.151	0.003	0.046	0.146
TP	-0.143	0.999	-0.072	0.908
Long-term environment variables	0.121	0.010	0.049	0.174
SWC	0.112	0.015	0.094	0.032
AP	0.003	0.387	0.064	0.124
DOC	0.307	0.001	0.206	0.001
DON	0.398	0.001	0.401	0.001
NH ₄ ⁺	0.223	0.001	0.023	0.350
NO ₃ ⁻	-0.135	0.990	0.040	0.234
Short-term environment variables	0.523	0.001	0.465	0.001
Latitude	-0.541	1.000	-0.433	1.000
Longitude	-0.525	1.000	-0.430	1.000
Distance*	-0.544	1.000	-0.438	1.000
Altitude	-0.672	1.000	-0.588	1.000
Geographic variables	-0.5664	1.000	-0.462	1.000

181 * Calculated by the geographic distance for paired samples based on the longitude and latitude of each sample

182

183 By Mantel test, the prokaryotic composition dissimilarity in top- or subsoil was significantly
184 correlated with geographic distance or altitude within the alpine or temperate biota and across the
185 alpine and temperate biotas (Supplementary file 1), except that the subsoil prokaryotic
186 composition dissimilarity within the temperate biota was not linked to geographic distance.
187 However, by partial Mantel, the prokaryotic composition dissimilarity in top- or subsoil was not
188 correlated with geographic variables when across biotas (Table 1).

189

190 As revealed by Partial Mantel (Table 1 and Supplementary file 2) test, the significant decay
191 relationship between topsoil prokaryotic community similarity and relatively short-term
192 environmental distance across biotas was mainly driven by soil water content (SWC, $r = 0.112$, p
193 $= 0.015$), dissolved organic carbon (DOC, $r = 0.307$, $p = 0.001$), dissolved organic nitrogen

194 (DON, $r = 0.398$, $p = 0.001$) and NH_4^+ ($r = 0.223$, $p = 0.001$). Similar short-term environmental
195 variables (except NH_4^+) were responsible for the significant decay relationship in the subsoil.

196

197 Within the alpine biota, the significant distance-decay relationship between topsoil prokaryotic
198 community similarity and relatively long-term environmental distance was driven by mean
199 annual precipitation (MAP, $r = 0.235$, $p = 0.002$). Except the relatively long-term variable of
200 MAP ($r = 0.199$, $p = 0.007$, Table S2), the short-term environmental variable of DON ($r = 0.211$,
201 $p = 0.029$) was also responsible for the significant distance-decay relationship of prokaryotic
202 community similarity in the subsoil within the alpine biota. Within the temperate biota, the
203 significant distance-decay relationship between topsoil prokaryotic community similarity and
204 relatively long-term environmental distance was driven by MAP ($r = 0.310$, $p = 0.001$).

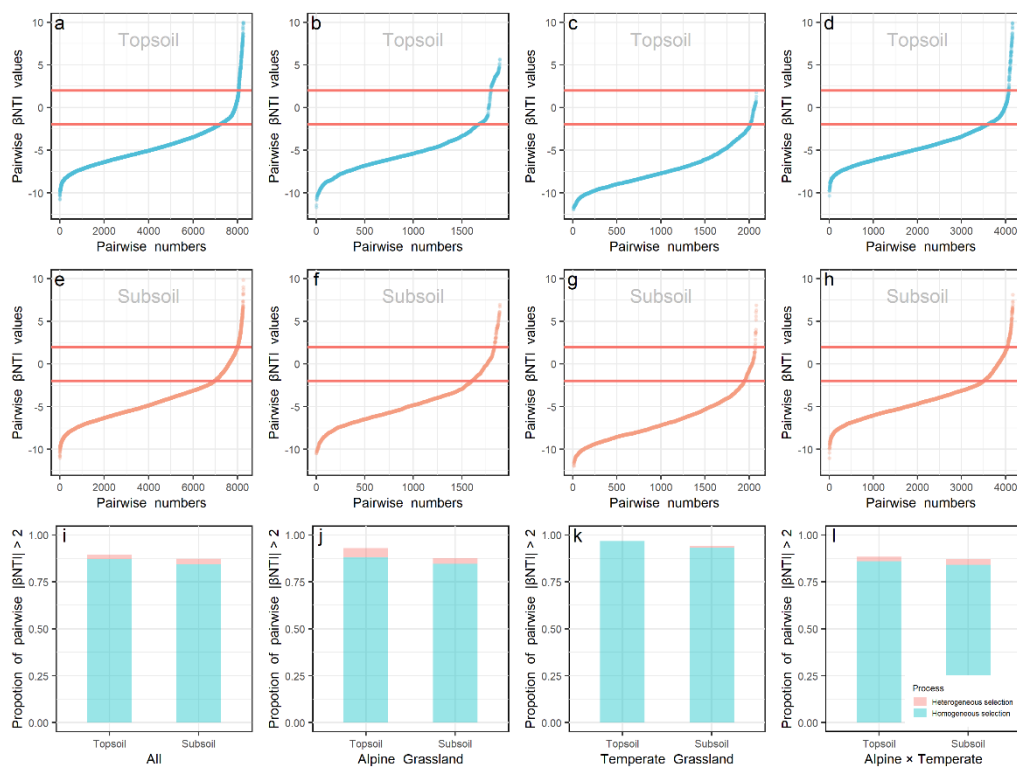
205

206 **Deterministic and stochastic processes in prokaryotic community assembly**

207 Null model and β NTI (β -nearest taxon index) analyses were used to explore the roles of
208 deterministic and stochastic processes on community assembly. The similarity of observed
209 communities in all samples (Figure 3 - figure supplement 1c) or samples in each biota (Figure 3 -
210 figure supplement 1a and b) was significantly ($P < 0.001$) higher than that of the permuted
211 communities generated by the null model, revealing the important role of deterministic processes
212 in determining the community assembly. As shown in Figure 3, the range of $|\beta\text{NTI}| > 2$ also
213 confirmed the importance of deterministic processes in prokaryotic community assembly, and
214 identified the dominant role (>84%) of homogeneous environmental selection, which means

215 selection under homogeneous abiotic and biotic environmental conditions leading to more
 216 similar structures among communities (45). Specifically, the contribution of deterministic
 217 processes was relatively lower in the alpine (Figure 3j; 92.91% and 87.47% in top- and subsoils,
 218 respectively) than temperate (96.63% and 94.04% in top- and subsoils, respectively) biota, and
 219 higher in the topsoil than subsoil in all sites. Moreover, most β NTI values were less than -2
 220 either in the top- (86.01% to 96.63%) or subsoil (84.09% to 93.37%) from all sites, indicating
 221 that prokaryotic communities were assembled mainly by homogeneous selection, in
 222 deterministic processes.

223

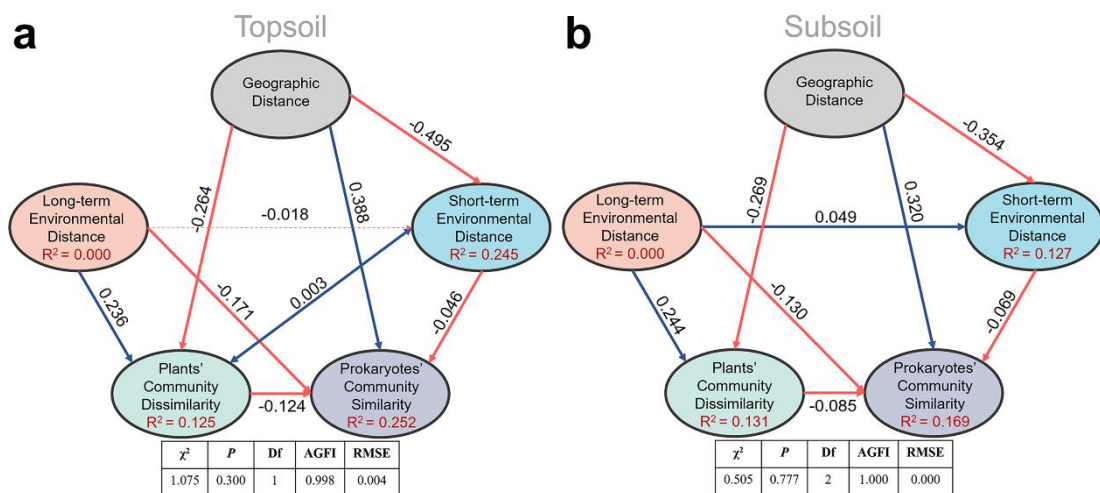


224

225 **Figure 3 β NTI values.** β NTI values of prokaryote in the top- (a, b, c and d) and subsoil (e, f, g
 226 and h) from all sites (a and e), within alpine (b and f) or temperate biota (c and g), and pairwise
 227 sites in the alpine cross temperate biotas (d and h) were presented. The proportion of $|\beta$ NTI| > 2

228 (deterministic processes) in the top- and subsoils from all sites (i), within alpine biota (j), within
 229 temperate biota (k) and pairwise sites in the alpine cross temperate biotas (l) were presented.
 230 Light blue and orange colors stand for β NTI values and their proportions in the top- and subsoils,
 231 respectively.
 232
 233 We further compared the immigration rates (m) of prokaryotes (Figure 3 - figure supplement 2)
 234 based on the algorithm developed by Hubbell for the neutral theory (49). Prokaryotic
 235 immigration rates were significantly lower in the alpine (0.159 ± 0.008 and 0.146 ± 0.008 in top-
 236 and subsoils, respectively, $P < 0.01$) than temperate biota (0.261 ± 0.010 and 0.246 ± 0.009 in top-
 237 and subsoils, respectively, $P < 0.01$) in the same soil layer. Moreover, immigration rates were
 238 higher in the top- (0.159 ± 0.008 and 0.261 ± 0.010 in the alpine and temperate biotas,
 239 respectively, $P < 0.01$) than subsoil (0.146 ± 0.008 and 0.246 ± 0.009 in the alpine and temperate
 240 biotas, respectively, $P < 0.01$) in the same biota.

241



242

243 **Figure 4 Structure equation model to quantify effects of geographic distance, relatively**

244 **long-term and short-term environmental distance, and plant community dissimilarity on**
245 **soil prokaryotic community similarity in the pairwise sites between alpine cross temperate**
246 **biotas, either in top-(a) or subsoil (b).** Red and blue lines stand for negative and positive
247 correlations, respectively; bold lines stand for significance at $p = 0.05$ level.

248

249 In the structural equation model (SEM), plant community dissimilarity (1-similarity) rather than
250 its similarity was used to parallel to geographical distance and environmental distance. As is
251 shown in Figure 4 and S8, SEM results showed that long-term environmental variables,
252 including soil properties (SOC, TN, TP, and pH) and climate factors (MAT and MAP), had a
253 strong influence on the plant community; while the short-term environmental variables (i.e.,
254 SWC, DOC, DON, NH_4^+ , NO_3^- and AP) hadn't. In the pairwise sites between alpine cross
255 temperate biotas, SEM (Figure 4) showed prokaryotic community similarity was mainly affected
256 by geographic distance ($r = 0.388$ and 0.320 in top- and subsoils, respectively), relatively
257 long-term environmental distance ($r = -0.171$ and -0.130 in top- and subsoils, respectively),
258 plant community dissimilarity ($r = -0.124$ and -0.065 in top- and subsoils, respectively), and
259 relatively short-term environmental distance ($r = -0.046$ and -0.069 in top- and subsoils,
260 respectively).

261

262 Within each biota, geographic distance only had a direct effect on topsoil prokaryotic community
263 similarity ($r = -0.275$) in the temperate grassland, while its effect in topsoil of the alpine biota
264 was only indirect through relatively short-term ($r = 0.140$) and long-term environmental

265 distances ($r = 0.222$), as well as plant community dissimilarity ($r = 0.232$). In the alpine biota,
266 increases in plant community dissimilarity, relatively long-term and short-term environmental
267 distances directly decreased the similarity of soil prokaryotic community. The explained
268 variances of soil prokaryotic community similarity were 26.8% and 21.3% in the top- and
269 subsoils, respectively (Figure 4 - figure supplement 1). In the temperate biota, other than the
270 direct effect of geographic distance, plant community dissimilarity, relatively long-term and
271 short-term environmental distances also affected topsoil prokaryotic community similarity
272 directly. In the subsoil of the temperate biota, prokaryotic community similarity was not
273 significantly correlated with any factors and the explained variance was merely 0.2%.

274

275 **Discussion**

276 “Everything is related to everything else, but near things are more related to each other” is
277 termed as "the first law of geography" (46). Ecologists and biogeographers refer it to the
278 negative relationship between community similarity and distance as a geographical
279 distance-decay relationship (31, 47). Though being regarded as a principal generalization, the
280 geographic distance-decay relationship was denied and overturned in this study as prokaryotic
281 community similarity increased over geographic distance after tipping points of 1,920 - 1,760 km
282 when across biotas. This finding is contradictory with most previous studies, including a report
283 that was conducted even at the similar scale of 4,000 km to ours but within a single biota
284 (temperate biota) (15). Consistently, when within a single biota of alpine or temperate grassland,
285 the distance-decay relationship was still valid in this study as well.

286

287 The observed U pattern of prokaryotic community similarity increased over geographic distance
288 in this study was likely due to decreased disparities in environmental heterogeneity over
289 geographic distance when across biotas, supported by three lines of evidences. Firstly, over
290 environmental distance, prokaryotic community similarity showed significant decay
291 relationships (Figure 2 - figure supplement 1) in all sites on a scale up to 4,000 km in this study,
292 consistent with previous reports (48, 49). "Everything is everywhere, but environment selects"
293 (EIE) predicts a distance-decay relationship because environmental selection would lead to a
294 larger community dissimilarity with larger geographic distance, which usually implies higher
295 environmental heterogeneity. However, the EIE hypothesis denies dispersal limitation, whose
296 importance has been supported by some evidences of endemic or non-cosmopolitan distribution
297 of certain microorganisms in certain habitats, e.g. the cyanobacterial genus *Synechococcus* in
298 mats of hot springs (50, 51). Nevertheless, in this study, we found that the similarity of soil
299 prokaryotic community decreased with environmental distance rather than geographic distance
300 when across biotas or after tipping points of 1,760 - 1,920 km. Together with other studies
301 discovering the cosmopolitan distribution (52, 53) of prokaryotes especially at higher taxonomic
302 levels, the observation of this study strongly supports the EIE hypothesis as indicating that all or
303 most prokaryotic groups surveyed in this study reached all or most of their potentially suitable
304 habitats.

305

306 Secondly, null model and β NTI analyses demonstrated that the prokaryotic community assembly

307 was dominantly determined by deterministic processes, especially homogeneous selection
308 referring that similar habitats (environment) harbor similar prokaryotic communities (54). The
309 role of environment filtering, including biotic and abiotic factors, on microbial community
310 assembly has been widely reported at the scale of hundreds to thousands of kilometers (19, 40,
311 41). This line of evidence also confirms the importance of environmental heterogeneity in
312 determining the observed geographic pattern of soil prokaryotic community. In this study,
313 prokaryotic composition dissimilarity in top or subsoil across the alpine and temperate biotas
314 was not correlated with geographic variables by partial mantel test, but with certain
315 environmental variables. These phenomena indicate that influences of these environmental
316 variables overwhelmed the geographic “distance” effect on shaping the microbial community
317 composition when across biotas. Consistently, even clear biogeographic patterns of soil microbial
318 communities over space (e.g. Taxa-area relationship) reported in previous studies were also
319 mainly attributed to environmental heterogeneity (55, 56) rather than geographic distance (55).

320

321 Thirdly, the similarity of relatively short-term environmental variables exhibited a U shape
322 pattern over geographic distance in all sites on a scale up to 4,000 km, while that of relatively
323 long-term environmental variables did not, indicating that relatively short-term environmental
324 variables may be responsible for the homogeneous selection shaping the U shape pattern of
325 prokaryotic community over geographic distance. Environmental variables measured in this
326 study were separated into relatively long-term and short-term environmental variables, judged by
327 the dynamic time (57). Consistently, SEM analyses also revealed a significant direct effect of

328 relatively short-term environmental distance on prokaryotic community similarity when across
329 biotas. The Partial Mantel test further demonstrated that soil water content and dissolved
330 nutrients (DOC and DON) were the primary short-term environmental variables in the significant
331 relationship between prokaryotic community similarity and geographic distance. The effects of
332 water on the microbial community have been widely reported (58, 59), as soil water content
333 could determine soil texture, bulk density, oxygen availability and connectivity within soils
334 (60-62), which can vitally influence soil microbial community composition (63) and microbial
335 basal respiration in the semi-arid area (64). Soil water content also influences microbial
336 communities through changing nutrient availability. Plenty of studies found that DOC and DON
337 affected the distribution pattern of soil microbes (41, 65-68). Compared to other nutrients, DOC
338 and DON can be utilized by microbes more directly and easily to provide energy and nutrients
339 for supporting their growth (69, 70).

340

341 The sampling transects we used in this study had also been adopted, though within a single biota,
342 region or climate type, in previous studies (15, 41). Precipitation (MAP) in our sampling sites
343 tends to decrease from northeast to southwest within the temperate biota, while it tends to
344 decrease from southeast to northwest within the alpine biota. Temperature (MAT) in our
345 sampling sites tends to decrease from southwest to northeast within the temperate biota, while it
346 tends to decrease from southeast to northwest within the alpine biota. However, in the topsoil
347 within the temperate biota or both top- and subsoils within the alpine biota, MAP rather than
348 MAT was responsible for the significant decay relationship between prokaryotic community

349 similarity and long-term environmental distance. These phenomena indicated that the spatial
350 pattern of soil prokaryotic community was controlled by long-term precipitation condition rather
351 than temperature within the alpine or temperate biota. Interestingly, though there was no
352 significant correlation between prokaryotic community similarity and MAT or MAP when across
353 biotas, the importance of soil water content and dissolved nutrients were discovered.

354

355 In addition to abiotic factors, plant community attributes (71-75), especially plant species
356 identity (48, 76, 77), may also be important for the U-shape pattern of prokaryotic community
357 similarity. Our results demonstrated that soil prokaryotic community similarity decreased over
358 the plant community dissimilarity in all sites on a scale up to 4,000 km. Moreover, SEM also
359 revealed the significantly direct effect of plant community dissimilarity on prokaryotic
360 community similarity in pairwise sites between alpine cross temperate biotas. Plant community
361 composition and diversity can affect soil prokaryotic communities through altering the quality
362 and quantity of organic matter input to soils by the forms of litterfall and root exudates (78).
363 Plants exude a substantial proportion (11 - 40%) of photosynthesis-derived carbon (79),
364 including sugars, amino acids, organic acids, fatty acids and secondary metabolites (79-81).
365 Their compounds in exudation can attract beneficial microorganisms deliberately and influence
366 the assembly of soil microbiomes to promote plants' adaptation to the surrounding environment
367 (82-87).

368

369 In the topsoil, the turnover rate of prokaryote was higher in temperate than alpine biota (Figure

370 1). We found that the immigration rate (*m*) of topsoil prokaryotes was higher in the temperate
371 biota than that of the alpine biota (Figure 3 - figure supplement 2), indicating a weakened
372 dispersal limitation that may be responsible for the higher similarity of prokaryotic community in
373 the temperate biota (2, 88). The effects of dispersal limitation on microbial communities (43, 55,
374 89) were dependent on ecosystems or environmental habitats (90-92). Harsh environments (88,
375 93, 94) with low temperature, high UV and complex mountain terrain in the alpine biota on the
376 Tibet Plateau would not be conducive for soil prokaryote to disperse. In contrast, the temperate
377 biota has benign temperature, low UV and better landscape connectivity to promote the spatial
378 dispersal of microorganisms.

379

380 Notably, within the temperate biota across 1,661 km, prokaryotic community similarity did not
381 change over geographic distance in the subsoil. Moreover, subsoil prokaryotic community
382 similarity was not linked with plant community dissimilarity (Figure 1 - figure supplement 3)
383 and long-term environmental distance (Figure 2 - figure supplement 1). It was correlated weakly
384 with the short-term environmental distance based on the correlation test ($R^2 = 0.011$) and SEM (r
385 $= - 0.002$). These phenomena denied the possibility that prokaryote community assembly was
386 dependent on plant community or environment heterogeneity in subsoil of the temperate biota.
387 Cases of no distance-decay relationship for microorganisms have also been reported previously
388 (29, 95, 96), possibly explained by factors like the paleogeographic history (95, 96) and
389 plant-dependent (96).

390

391 **Conclusion**

392 This study provides a systematical analysis of the spatial pattern of soil prokaryotic communities
393 in the northern grassland of China. Soil prokaryotic similarity exhibited a U-shape distribution
394 pattern over geographic distance at a scale of up to 4,000 km. This finding overturns the
395 well-accepted geographic distance-decay relationship, which was still valid in the top- or
396 subsoils within the alpine biota and only in the topsoil within the temperate biota. Despite
397 different climate and ecosystem types in the alpine and temperate biotas, habitats far more apart
398 when across biotas were more similarly as revealed by the U-shape pattern for short-term
399 environmental variables over geographic distance. Consistently, deterministic processes were
400 found to dominate the soil prokaryotic community assembly by null model and β NTI analyses,
401 and further partial Mantel analysis revealed that soil water content and dissolved nutrients (DOC
402 and DON) may be responsible for the decay relationship of prokaryotic community similarity
403 over environmental distance, overturning the geographic distance-decay relationship.

404

405 **Materials and Methods**

406 **Study sites and field sampling**

407 The alpine grassland in the region of Qinghai-Tibet Plateau belongs to the Qinghai Tibet Plateau
408 biota, and the temperate grassland in the region of Inner Mongolia belongs to Eurasian grassland
409 biota, according to the Chinese floristic geography (97). A total of 129 sites and 258 samples
410 were collected from the northern grassland of China. Among them, 128 samples from 64 sites
411 (red dots in Figure 1 - figure supplement 1) were collected from July 29 to August 14, 2014 in

412 the Qinghai-Tibet Plateau alpine biota, China. Alpine biota sample sites covered a variety of
413 alpine ecosystems, including alpine meadow, alpine steppe, alpine desert, and shrub. 130
414 samples from 65 sites (blue dots in Figure 1 - figure supplement 1) were collected from
415 September 10 to 24, 2015 in the temperate biota, in Inner Mongolia, China. Temperate biota
416 sample sites covered three types of temperate grassland ecosystem, namely temperate meadow,
417 temperate steppe, and temperate desert. The distance between each two adjacent sample sites was
418 no less than 60 km and removed from potential human interference such as towns, villages, and
419 roads.

420

421 A GPS (global positioning system) was used to record the geographic coordinates and altitudes
422 of each sample site. Five plots (1×1m²) were selected randomly at each site and the distance
423 between adjacent plots was more than 10 m. In each plot, plant species were identified and their
424 aboveground parts were collected separately by species. After being delivered to the laboratory,
425 the aboveground plant part of each species was dried at 65 °C for 72 hours and weighted as the
426 aboveground biomass (AGB, g/ m²). The community AGB in each plot was calculated by the
427 sum of AGBs for all collected species in that plot. After removing plant aboveground biomass
428 and litter, three topsoil (0-5 cm in depth) and subsoil (5-20 cm in depth) cores (7 cm in diameter)
429 were randomly sampled within each plot.

430

431 Topsoil or subsoil samples from each plot were pooled and then sieved through a 2 mm mesh,
432 and the roots were selected as belowground biomass (BGB). Sieved topsoil or subsoil samples

433 were divided into two subsamples. One part was stored at room temperature and dried in the
434 shade for measuring physical and chemical properties. The other part was stored at
435 approximately 4 °C in the field by a mobile refrigerator, delivered with dry ice to the laboratory
436 in Beijing, and finally frozen at -80 °C in a freezer before DNA extraction.

437

438 **Soil properties**

439 Soil pH was measured by pH meter (STARTER3100, Ohaus Instruments Co., Ltd., Shanghai,
440 China) with a 1:5 of soil water ratio (5 g soil: 25 mL ddH₂O). SWC was measured by ovening
441 fresh soil samples at 105°C for 24 h. SOC was measured by a TOC analyzer (Liqui TOC II;
442 Elementar Analysensysteme GmbH, Hanau, Germany). Soil TN was measured on an
443 auto-analyzer (SEAL Analytical GmbH, Norderstedt, Germany). Soil TP and AP were measured
444 by a UV-VIS spectrophotometer (UV2700, SHIMADZU, Japan). Nitrate-N (NO₃⁻) and
445 ammonium-N (NH₄⁺) were extracted with 2 M KCl (soil mass to solution ratio of 1:5) and then
446 analyzed on a continuous-flow ion auto-analyzer (SEAL Analytical GmbH, Norderstedt,
447 Germany). Soil DOC and DON were measured on a TOC Analyser (Liqui TOC II; Elementar
448 Analysensysteme GmbH, Hanau, Germany). Plant aboveground biomass (AGB) and
449 belowground biomass (BGB) were measured after oven drying at 65 °C for 72 h. MAT
450 (1980-2014) and MAP (1980-2014) of each study site were obtained from “China
451 Meteorological Data Service Center” (CMDC: <https://data.cma.cn/>) by latitude and longitude.

452

453 **Microbial analysis**

454 Soil genomic DNA was extracted from 0.25 g frozen soil three times at each soil layer at each
455 site and then mixed into one DNA sample using PowerSoil™ DNA Isolation Kits (MO BIO
456 Laboratories, Carlsbad, CA, USA). The quality of extracted DNA was assessed based on OD
457 260/280 nm and 260/230 nm absorbance ratios by NanoDrop (2000) spectrophotometer
458 (NanoDrop Technologies inc., Wilmington, DE, USA). Primer pair 515F
459 (5'-GTGYCAGCMGCCGCGGTA-3') and 909R (5'-CCCCGYCAATTCMTTTRAGT-3') was
460 selected to amplify the V4-V5 region of 16S rRNA and the target fragment length was 374 bp,
461 and the 12bp barcode was added at the end of 5' of 515F. A 50 µL PCR reaction system was
462 configured in 0.2 mL tube, including 2 µL template DNA diluent, 4 µL dNTP, 4 µL Mg₂⁺, 5 µL
463 Buffer, 0.5 L Ex Taq™ enzyme, 1 µL forward primer, 1 µL reverse primer, 32.5 L ddH₂O. The
464 PCR procedure was performed as follows: predenaturation at 95°C for 10 min, 30 PCR cycles
465 (denaturation at 94°C for 30 s, annealing at 53°C for 25 s, extension at 68°C for 45 s), and a final
466 extension at 72°C for 10 min. The PCR products were purified by 1% agarose gel using
467 GeneJET Gel Extraction Kit (Thermo, USA). The purified DNA was tested by NanoDrop (2000)
468 spectrophotometer (NanoDrop Technologies inc., Wilmington, DE, USA). All purified DNA
469 samples were mixed in 100 ng before database construction and sequencing, which was
470 performed by Illumina Miseq in Chengdu Biology Institute.

471

472 The MiSeq raw data was analyzed by UPARSE pipeline with USEARCH 8 software to obtain an
473 operational taxon units (OTU) table. The double ended sequencing was performed by the
474 Illumina Miseq platform with 2 × 250 bp V2 Kits at Chengdu Institute of Biology, Chinese

475 Academy of Sciences. Raw reads generated from the Miseq paired-end sequencing were merged
476 by Fast Length Adjustment of Short reads (FLASH) (98). The fastq sequencing files were
477 combined into one new fastq file. By USEARCH v8.0.1623 (99), barcode sequences were
478 replaced by new labels, and the forward and backward primer sequences were removed by the
479 command of -fastq_filter. The unique representative sequences were selected by the command of
480 -fastx_uniques. After discarding singletons, OTUs were clustered at the 97% threshold and
481 chimeras were removed simultaneously by the command of -cluster_otus. The command of
482 -usearch_global was then used to map the fastq file into an OTU table. Each OTU was annotated
483 by Mothur v1.27 (100) with classify.seqs command, and sliva.nr_v128.align was selected as the
484 reference database. The OTU table was resampled to the same sequence (Figure 1 - figure
485 supplement 4) before further analysis by R 3.5.0 with the resample package.

486

487 **Statistical analysis**

488 To compare the soil bacterial samples from different climate regions, we divided the soil samples
489 into alpine samples and temperate samples according to collection sites. The altitude of the
490 alpine biota sampling sites ranged from 2,796 to 4,891 m, and that of temperate biota was from
491 10 to 1,796 m. According to the sampling position in the soil layers, samples were divided into
492 topsoil (0-5 cm) samples and subsoil (5-20 cm) samples. The geographic distance (km) between
493 each two sites was calculated based on their longitude and latitude using the "distGeo" function
494 of "geosphere" package in R program, which considers spheric deviations. Plant communities
495 were classified into four functional groups (grasses, sedges, legumes, and forbs) and plant

496 communities' similarity and dissimilarity were calculated based on Bray-Curtis distance by the
497 vegan package of R. The relatively long-term and short-term environmental variables were
498 defined based on whether they remain stable within one year or not. For example, SOC is
499 believed to be stable in natural grasslands for years (101), thus being divided into the relatively
500 long-term environmental variables; while AP (102), DOC (103) and DON (104) are reported to
501 be dynamic within one year, thus being divided into the relatively short-term environmental
502 variables. This division has been used previously (57). In this study, relatively long-term
503 environmental variables included MAP, MAT, pH, SOC, TN, and TP, while relatively short-term
504 environmental variables included SWC, AP, DOC, DON, NH_4^+ , and NO_3^- .

505

506 The Bray-Curtis similarity and dissimilarity of the prokaryotic community were calculated using
507 OTU tables resampled to a minimum number of sequences from each sample (7500 in this study).
508 The Mantel test and Partial Mantel test based on a Pearson correlation were used to test the
509 relationship of soil prokaryotic similarity, geographic distance, and long-term multiple
510 environmental variables or short-term multiple environmental variables. The turnover rate was
511 estimated by the slope of the linear regression model based on the least square method. The
512 tipping point was calculated by the function of $d(Y)/d(x) = 0$ in binomial function. Pearson
513 correlation was used to test the relationship of soil prokaryotic diversity with environmental
514 variables.

515

516 Null model (105) and β NTI analyses (106) were used to distinguish different ecological

517 processes, including deterministic processes (homogenous selection and heterogeneous
518 selection), random dispersal (homogenous dispersal, dispersal limitation), drift, and
519 diversification (88). Null model analysis was performed by comparing the Bray-Curtis similarity
520 of observed communities and randomly permuted communities using R3.5.0 with package
521 *vegan* and *picante*. The $|\beta\text{NTI}| > 2$ means community was constructed by deterministic processes,
522 and $\beta\text{NTI} < -2$ means homogenous selection plays a major role, while $\beta\text{NTI} > +2$ means
523 heterogeneous is more important. The $-2 < \beta\text{NTI} < +2$ means stochastic processes determined
524 community succession (107). A βNTI analysis was performed by R 3.5.0 with the *ape* package.
525 The estimation of immigration rate (m) was calculated by TeTame 2.0 (108) based on Hubbell's
526 neutral theory of biodiversity (109). Parameter estimation was rigorously performed by
527 maximum-likelihood using the sampling formula developed by Etienne (110-113). This model is
528 seen as a potentially useful null model in ecology; in this model, the species relative abundances
529 in a guild are determined by two parameters, namely θ and m . The θ governs the appearance of a
530 new species in the regional species pool, and m governs immigration into local communities of
531 individuals from the regional species pool. We further used SEM to disentangle the causal
532 pathways through which geographic distance, short-term environmental distance, long-term
533 environmental distance, and plants' community dissimilarity influence soil prokaryotic similarity.
534 SEM in this study is implemented by AMOS software.

535

536 **Acknowledgement**

537 This work was financially supported by the Strategic Priority Research and Program A

538 (XDA20050104) of the Chinese Academy of Sciences (CAS), National Natural of China,
539 Science Foundation (42041005), The Second Tibetan Plateau Scientific Expedition and Research
540 (STEP) program (Grant No. 2019QZKK0304), Strategic Priority Research Program A
541 (XDA1907304) and Program B (XDB15010201) of CAS, CAS "Light of West China" Program,
542 and Sanjiangyuan National Park Joint Program (LHZX-2020-02-01).

543

544 **Additional information**

545 Supplementary information is available for this paper. The soil prokaryote dataset has been
546 deposited in the NCBI Sequence Read Archive under accession number: PRJNA 729210. Other
547 datasets including OTU table, plant community data and environmental variables are available at
548 <https://github.com/zhangbiao1989/Support-files.git> that can be accessed by the public.

549

550 **Competing interests**

551 The authors declare no competing financial interests.

552

553 **References**

- 554 1. Bahram M, Koljalg U, Courty P-E, Diedhiou AG, Kjoller R, Polme S, et al. The distance decay
555 of similarity in communities of ectomycorrhizal fungi in different ecosystems and scales. *J Ecol.*
556 2013;101(5):1335-44.
- 557 2. Hanson CA, Fuhrman JA, Horner-Devine MC, Martiny JBH. Beyond biogeographic patterns:
558 processes shaping the microbial landscape. *Nat Rev Microbiol.* 2012;10(7):497-506.

- 559 3. Condit R, Pitman N, Leigh EG, Chave J, Terborgh J, Foster RB, et al. Beta-diversity in tropical
560 forest trees. *Science*. 2002;295(5555):666-9.
- 561 4. Tuomisto H, Ruokolainen K, Yli-Halla M. Dispersal, environment, and floristic variation of
562 western Amazonian forests. *Science*. 2003;299(5604):241-4.
- 563 5. Gilbert B, Lechowicz MJ. Neutrality, niches, and dispersal in a temperate forest understory.
564 *Proceedings of the National Academy of Sciences of the United States of America*.
565 2004;101(20):7651-6.
- 566 6. Dexter KG, Terborgh JW, Cunningham CW. Historical effects on beta diversity and community
567 assembly in Amazonian trees. *Proceedings of the National Academy of Sciences of the United States*
568 *of America*. 2012;109(20):7787-92.
- 569 7. Zhao ZQ, Li SC, Liu JG, Peng J, Wang YL. The distance decay of similarity in climate variation
570 and vegetation dynamics. *Environ Earth Sci*. 2015;73(8):4659-70.
- 571 8. Draper FC, Baraloto C, Brodrick PG, Phillips OL, Vasquez Martinez R, Honorio Coronado EN,
572 et al. Imaging spectroscopy predicts variable distance decay across contrasting Amazonian tree
573 communities. *J Ecol*. 2019;107(2):696-710.
- 574 9. Gomez-Rodriguez C, Baselga A. Variation among European beetle taxa in patterns of distance
575 decay of similarity suggests a major role of dispersal processes. *Ecography*. 2018;41(11):1825-34.
- 576 10. van der Mescht L, Warburton EM, Khokhlova IS, Stanko M, Vinarski MV, Korralo-Vinarskaya
577 NP, et al. Biogeography of parasite abundance: latitudinal gradient and distance decay of similarity in
578 the abundance of fleas and mites, parasitic on small mammals in the Palearctic, at three spatial scales.
579 *Int J Parasitol*. 2018;48(11):857-66.

- 580 11. Warfe DM, Pettit NE, Magierowski RH, Pusey BJ, Davies PM, Douglas MM, et al.
581 Hydrological connectivity structures concordant plant and animal assemblages according to niche
582 rather than dispersal processes. *Freshwater Biol.* 2013;58(2):292-305.
- 583 12. Beisner BE, Peres PR, Lindstrom ES, Barnett A, Longhi ML. The role of environmental and
584 spatial processes in structuring lake communities from bacteria to fish. *Ecology.*
585 2006;87(12):2985-91.
- 586 13. Matsuoka S, Kawaguchi E, Osono T. Temporal distance decay of similarity of ectomycorrhizal
587 fungal community composition in a subtropical evergreen forest in Japan. *Fems Microbiol Ecol.*
588 2016;92(5).
- 589 14. Oono R, Rasmussen A, Lefevre E. Distance decay relationships in foliar fungal endophytes are
590 driven by rare taxa. *Environ Microbiol.* 2017;19(7):2794-805.
- 591 15. Wang XB, Lu XT, Yao J, Wang ZW, Deng Y, Cheng WX, et al. Habitat-specific patterns and
592 drivers of bacterial beta-diversity in China's drylands. *Isme J.* 2017;11(6):1345-58.
- 593 16. Finkel OM, Burch AY, Elad T, Huse SM, Lindow SE, Post AF, et al. Distance-Decay
594 Relationships Partially Determine Diversity Patterns of Phyllosphere Bacteria on Tamrix Trees across
595 the Sonoran Desert. *Appl Environ Microb.* 2012;78(17):6187-93.
- 596 17. Jiao S, Yang Y, Xu Y, Zhang J, Lu Y. Balance between community assembly processes mediates
597 species coexistence in agricultural soil microbiomes across eastern China. *Isme J.* 2020;14(1):202-16.
- 598 18. Jiao S, Xu Y, Zhang J, Lu Y. Environmental filtering drives distinct continental atlases of soil
599 archaea between dryland and wetland agricultural ecosystems. *Microbiome.* 2019;7.
- 600 19. Shi Y, Li YT, Xiang XJ, Sun RB, Yang T, He D, et al. Spatial scale affects the relative role of

601 stochasticity versus determinism in soil bacterial communities in wheat fields across the North China
602 Plain. *Microbiome*. 2018;6.

603 20. Zinger L, Boetius A, Ramette A. Bacterial taxa-area and distance-decay relationships in marine
604 environments. *Mol Ecol*. 2014;23(4):954-64.

605 21. Barreto DP, Conrad R, Klose M, Claus P, Enrich-Prast A. Distance-Decay and Taxa-Area
606 Relationships for Bacteria, Archaea and Methanogenic Archaea in a Tropical Lake Sediment. *Plos*
607 *One*. 2014;9(10).

608 22. Shi Y, Adams JM, Ni Y, Yang T, Jing X, Chen L, et al. The biogeography of soil archaeal
609 communities on the eastern Tibetan Plateau. *Sci Rep-Uk*. 2016;6.

610 23. Chen W, Ren K, Isabwe A, Chen H, Liu M, Yang J. Stochastic processes shape microeukaryotic
611 community assembly in a subtropical river across wet and dry seasons. *Microbiome*. 2019;7(1).

612 24. Wang J, Zhang T, Li L, Li J, Feng Y, Lu Q. The Patterns and Drivers of Bacterial and Fungal
613 beta-Diversity in a Typical Dryland Ecosystem of Northwest China. *Front Microbiol*. 2017;8.

614 25. Yang T, Adams JM, Shi Y, Sun H, Cheng L, Zhang Y, et al. Fungal community assemblages in a
615 high elevation desert environment: Absence of dispersal limitation and edaphic effects in surface soil.
616 *Soil Biol Biochem*. 2017;115:393-402.

617 26. Zhang J, Zhang B, Liu Y, Guo Y, Shi P, Wei G. Distinct large-scale biogeographic patterns of
618 fungal communities in bulk soil and soybean rhizosphere in China. *Sci Total Environ*.
619 2018;644:791-800.

620 27. Hu H-W, Zhang L-M, Yuan C-L, Zheng Y, Wang J-T, Chen D, et al. The large-scale distribution
621 of ammonia oxidizers in paddy soils is driven by soil pH, geographic distance, and climatic factors.

- 622 Front Microbiol. 2015;6.
- 623 28. Angermeyer A, Crosby SC, Huber JA. Decoupled distance-decay patterns between *dsrA* and 16S
624 rRNA genes among salt marsh sulfate-reducing bacteria. Environ Microbiol. 2016;18(1):75-86.
- 625 29. Zhou JZ, Kang S, Schadt CW, Garten CT. Spatial scaling of functional gene diversity across
626 various microbial taxa. Proceedings of the National Academy of Sciences of the United States of
627 America. 2008;105(22):7768-73.
- 628 30. Zhou J, Deng Y, Shen L, Wen C, Yan Q, Ning D, et al. Temperature mediates continental-scale
629 diversity of microbes in forest soils. Nat Commun. 2016;7.
- 630 31. Nekola JC, White PS. The distance decay of similarity in biogeography and ecology. Journal of
631 Biogeography. 1999;26(4):867-78.
- 632 32. Sojininen J, McDonald R, Hillebrand H. The distance decay of similarity in ecological
633 communities. Ecography. 2007;30(1):3-12.
- 634 33. Cottenie K. Integrating environmental and spatial processes in ecological community dynamics.
635 Ecology Letters. 2005;8(11):1175-82.
- 636 34. Bell T. Experimental tests of the bacterial distance-decay relationship. Isme J.
637 2010;4(11):1357-65.
- 638 35. Lindstrom ES, Ostman O. The Importance of Dispersal for Bacterial Community Composition
639 and Functioning. Plos One. 2011;6(10).
- 640 36. Leibold MA, Holyoak M, Mouquet N, Amarasekare P, Chase JM, Hoopes MF, et al. The
641 metacommunity concept: a framework for multi-scale community ecology. Ecology Letters.
642 2004;7(7):601-13.

- 643 37. Stegen JC, Lin X, Konopka AE, Fredrickson JK. Stochastic and deterministic assembly
644 processes in subsurface microbial communities. *Isme J.* 2012;6(9):1653-64.
- 645 38. Chase JM, Myers JA. Disentangling the importance of ecological niches from stochastic
646 processes across scales. *Philos T R Soc B.* 2011;366(1576):2351-63.
- 647 39. Hubbell SP. Neutral theory in community ecology and the hypothesis of functional equivalence.
648 *Functional Ecology.* 2005;19(1):166-72.
- 649 40. Feng M, Tripathi BM, Shi Y, Adams JM, Zhu Y-G, Chu H. Interpreting distance-decay pattern of
650 soil bacteria via quantifying the assembly processes at multiple spatial scales. *Microbiologyopen.*
651 2019;8(9).
- 652 41. Chu H, Sun H, Tripathi BM, Adams JM, Huang R, Zhang Y, et al. Bacterial community
653 dissimilarity between the surface and subsurface soils equals horizontal differences over several
654 kilometers in the western Tibetan Plateau. *Environ Microbiol.* 2016;18(5):1523-33.
- 655 42. Green JL, Holmes AJ, Westoby M, Oliver I, Briscoe D, Dangerfield M, et al. Spatial scaling of
656 microbial eukaryote diversity. *Nature.* 2004;432(7018):747-50.
- 657 43. Whitaker RJ, Grogan DW, Taylor JW. Geographic barriers isolate endemic populations of
658 hyperthermophilic archaea. *Science.* 2003;301(5635):976-8.
- 659 44. Okie JG, Van Horn DJ, Storch D, Barrett JE, Gooseff MN, Kopsova L, et al. Niche and
660 metabolic principles explain patterns of diversity and distribution: theory and a case study with soil
661 bacterial communities. *Proceedings of the Royal Society B-Biological Sciences.* 2015;282(1809).
- 662 45. MacArthur RH, O. WE. *The theory of island biogeography.* Princeton University Press, Princeton,
663 NJ. 1967.

- 664 46. Tobler WR, Mielke HW, Detwyler TR. Geobotanical distance between New-Zealand and
665 neighboring island. *Bioscience*. 1970;20(9):537-&.
- 666 47. Wang X, Wen X, Deng Y, Xia Y, Yang Y, Zhou J. Distance-Decay Relationship for Biological
667 Wastewater Treatment Plants. *Appl Environ Microb*. 2016;82(16):4860-6.
- 668 48. Leff JW, Bardgett RD, Wilkinson A, Jackson BG, Pritchard WJ, De Long JR, et al. Predicting
669 the structure of soil communities from plant community taxonomy, phylogeny, and traits. *Isme J*.
670 2018;12(7):1794-805.
- 671 49. Xing R, Gao Q-b, Zhang F-q, Wang J-l, Chen S-l. Large-scale distribution of bacterial
672 communities in the Qaidam Basin of the Qinghai-Tibet Plateau. *Microbiologyopen*. 2019;8(10).
- 673 50. Castenholz RW. The biogeography of hot spring algae through enrichment cultures. *Mitt Int Ver*
674 *Limnol* 21: 296–315. 1978.
- 675 51. Papke RT, Ramsing NB, Bateson MM, Ward DM. Geographical isolation in hot spring
676 cyanobacteria. *Environ Microbiol*. 2003;5(8):650-9.
- 677 52. Cho JC, Tiedje JM. Biogeography and degree of endemism of fluorescent *Pseudomonas* strains
678 in soil. *Appl Environ Microb*. 2000;66(12):5448-56.
- 679 53. DeLong EE, Pace NR. Environmental diversity of Bacteria and Archaea. *Systematic Biology*.
680 2001;50(4):470-8.
- 681 54. Su Y-g, Liu J, Zhang B-c, Zhao H-m, Huang G. Habitat-specific environmental factors regulate
682 spatial variability of soil bacterial communities in biocrusts across northern China's drylands. *Sci*
683 *Total Environ*. 2020;719.
- 684 55. Horner-Devine MC, Lage M, Hughes JB, Bohannan BJM. A taxa-area relationship for bacteria.

685 Nature. 2004;432(7018):750-3.

686 56. Ranjard L, Dequiedt S, Prevost-Boure NC, Thioulouse J, Saby NPA, Lelievre M, et al. Turnover
687 of soil bacterial diversity driven by wide-scale environmental heterogeneity. *Nat Commun.* 2013;4.

688 57. Zhou S, Xue K, Zhang B, Tang L, Pang Z, Wang F, et al. Spatial patterns of microbial
689 nitrogen-cycling gene abundances along a precipitation gradient in various temperate grasslands at a
690 regional scale. *Geoderma.* 2021;404.

691 58. Suseela V, Conant RT, Wallenstein MD, Dukes JS. Effects of soil moisture on the temperature
692 sensitivity of heterotrophic respiration vary seasonally in an old-field climate change experiment.
693 *Global Change Biology.* 2012;18(1):336-48.

694 59. Moyano FE, Manzoni S, Chenu C. Responses of soil heterotrophic respiration to moisture
695 availability: An exploration of processes and models. *Soil Biol Biochem.* 2013;59:72-85.

696 60. Kaye JP, Hart SC. Competition for nitrogen between plants and soil microorganisms. *Trends in*
697 *Ecology & Evolution.* 1997;12(4):139-43.

698 61. Brockett BFT, Prescott CE, Grayston SJ. Soil moisture is the major factor influencing microbial
699 community structure and enzyme activities across seven biogeoclimatic zones in western Canada.
700 *Soil Biol Biochem.* 2012;44(1):9-20.

701 62. Mansson KF, Olsson MO, Falkengren-Grerup U, Bengtsson G. Soil moisture variations affect
702 short- term plant- microbial competition for ammonium, glycine, and glutamate. *Ecol Evol.*
703 2014;4(7):1061-72.

704 63. Li Y, Adams J, Shi Y, Wang H, He J-S, Chu H. Distinct Soil Microbial Communities in habitats
705 of differing soil water balance on the Tibetan Plateau. *Sci Rep-Uk.* 2017;7.

- 706 64. Wichern F, Joergensen RG. Soil Microbial Properties Along a Precipitation Transect in Southern
707 Africa. *Arid Land Research and Management*. 2009;23(2):115-26.
- 708 65. Hu Y, Xiang D, Veresoglou SD, Chen F, Chen Y, Hao Z, et al. Soil organic carbon and soil
709 structure are driving microbial abundance and community composition across the arid and semi-arid
710 grasslands in northern China. *Soil Biol Biochem*. 2014;77:51-7.
- 711 66. Chen D, Mi J, Chu P, Cheng J, Zhang L, Pan Q, et al. Patterns and drivers of soil microbial
712 communities along a precipitation gradient on the Mongolian Plateau. *Landscape Ecol*.
713 2015;30(9):1669-82.
- 714 67. Zeng J, Liu X, Song L, Lin X, Zhang H, Shen C, et al. Nitrogen fertilization directly affects soil
715 bacterial diversity and indirectly affects bacterial community composition. *Soil Biol Biochem*.
716 2016;92:41-9.
- 717 68. He D, Xiang X, He J-S, Wang C, Cao G, Adams J, et al. Composition of the soil fungal
718 community is more sensitive to phosphorus than nitrogen addition in the alpine meadow on the
719 Qinghai-Tibetan Plateau. *Biol Fert Soils*. 2016;52(8):1059-72.
- 720 69. Alden L, Demoling F, Baath E. Rapid method of determining factors limiting bacterial growth in
721 soil. *Appl Environ Microb*. 2001;67(4):1830-8.
- 722 70. Drenovsky RE, Vo D, Graham KJ, Scow KM. Soil water content and organic carbon availability
723 are major determinants of soil microbial community composition. *Microbial Ecology*.
724 2004;48(3):424-30.
- 725 71. de Vries FT, Manning P, Tallowin JRB, Mortimer SR, Pilgrim ES, Harrison KA, et al. Abiotic
726 drivers and plant traits explain landscape-scale patterns in soil microbial communities. *Ecology*

727 Letters. 2012;15(11):1230-9.

728 72. Grayston SJ, Wang SQ, Campbell CD, Edwards AC. Selective influence of plant species on
729 microbial diversity in the rhizosphere. *Soil Biol Biochem.* 1998;30(3):369-78.

730 73. Singh BK, Millard P, Whiteley AS, Murrell JC. Unravelling rhizosphere-microbial interactions:
731 opportunities and limitations. *Trends Microbiol.* 2004;12(8):386-93.

732 74. Berg G, Smalla K. Plant species and soil type cooperatively shape the structure and function of
733 microbial communities in the rhizosphere. *Fems Microbiol Ecol.* 2009;68(1):1-13.

734 75. Hiiesalu I, Paertel M, Davison J, Gerhold P, Metsis M, Moora M, et al. Species richness of
735 arbuscular mycorrhizal fungi: associations with grassland plant richness and biomass. *New*
736 *Phytologist.* 2014;203(1):233-44.

737 76. Peay KG, Baraloto C, Fine PVA. Strong coupling of plant and fungal community structure
738 across western Amazonian rainforests. *Isme J.* 2013;7(9):1852-61.

739 77. Prober SM, Leff JW, Bates ST, Borer ET, Firn J, Harpole WS, et al. Plant diversity predicts beta
740 but not alpha diversity of soil microbes across grasslands worldwide. *Ecology Letters.*
741 2015;18(1):85-95.

742 78. van der Heijden MGA. The unseen majority: Soil microbes as drivers of plant diversity and
743 productivity in terrestrial ecosystems (vol 11, pg 296, 2008). *Ecology Letters.* 2008;11(6):651-.

744 79. Badri DV, Vivanco JM. Regulation and function of root exudates. *Plant Cell Environ.*
745 2009;32(6):666-81.

746 80. Bais HP, Weir TL, Perry LG, Gilroy S, Vivanco JM. The role of root exudates in rhizosphere
747 interactions with plants and other organisms. *Annual Review of Plant Biology.* 2006;57:233-66.

- 748 81. Baetz U, Martinoia E. Root exudates: the hidden part of plant defense. *Trends in Plant Science*.
749 2014;19(2):90-8.
- 750 82. Bulgarelli D, Schlaeppi K, Spaepen S, van Themaat EVL, Schulze-Lefert P. Structure and
751 Functions of the Bacterial Microbiota of Plants. In: Merchant SS, editor. *Annual Review of Plant*
752 *Biology*, Vol 64. *Annual Review of Plant Biology*. 642013. p. 807-38.
- 753 83. Hooper DU, Bignell DE, Brown VK, Brussaard L, Dangerfield JM, Wall DH, et al. Interactions
754 between aboveground and belowground biodiversity in terrestrial ecosystems: Patterns, mechanisms,
755 and feedbacks. *Bioscience*. 2000;50(12):1049-61.
- 756 84. Wardle DA. The influence of biotic interactions on soil biodiversity. *Ecology Letters*.
757 2006;9(7):870-86.
- 758 85. Mellado-Vazquez PG, Lange M, Bachmann D, Gockele A, Karlowy S, Milcu A, et al. Plant
759 diversity generates enhanced soil microbial access to recently photosynthesized carbon in the
760 rhizosphere. *Soil Biol Biochem*. 2016;94:122-32.
- 761 86. Jones DL. Organic acids in the rhizosphere - a critical review. *Plant and Soil*. 1998;205(1):25-44.
- 762 87. Zhalnina K, Louie KB, Hao Z, Mansoori N, da Rocha UN, Shi S, et al. Dynamic root exudate
763 chemistry and microbial substrate preferences drive patterns in rhizosphere microbial community
764 assembly. *Nature Microbiology*. 2018;3(4):470-80.
- 765 88. Zhou J, Ning D. Stochastic Community Assembly: Does It Matter in Microbial Ecology?
766 *Microbiology and Molecular Biology Reviews*. 2017;81(4).
- 767 89. Martiny JBH, Eisen JA, Penn K, Allison SD, Horner-Devine MC. Drivers of bacterial
768 beta-diversity depend on spatial scale. *Proceedings of the National Academy of Sciences of the*

769 United States of America. 2011;108(19):7850-4.

770 90. Chase JM. Drought mediates the importance of stochastic community assembly. Proceedings of
771 the National Academy of Sciences of the United States of America. 2007;104(44):17430-4.

772 91. Evans S, Martiny JBH, Allison SD. Effects of dispersal and selection on stochastic assembly in
773 microbial communities. *Isme J*. 2017;11(1):176-85.

774 92. Louca S, Jacques SMS, Pires APF, Leal JS, Srivastava DS, Parfrey LW, et al. High taxonomic
775 variability despite stable functional structure across microbial communities. *Nature Ecology &*
776 *Evolution*. 2017;1(1).

777 93. Zhang B, Xue K, Zhou S, Che R, Du J, Tang L, et al. Phosphorus mediates soil prokaryote
778 distribution pattern along a small-scale elevation gradient in Noijin Kangsang Peak, Tibetan Plateau.
779 *Fems Microbiol Ecol*. 2019;95(6).

780 94. Young JW, Locke JCW, Elowitz MB. Rate of environmental change determines stress response
781 specificity. *Proceedings of the National Academy of Sciences of the United States of America*.
782 2013;110(10):4140-5.

783 95. Cox F, Newsham KK, Bol R, Dungait JAJ, Robinson CH. Not poles apart: Antarctic soil fungal
784 communities show similarities to those of the distant Arctic. *Ecology Letters*. 2016;19(5):528-36.

785 96. Davison J, Moora M, Oepik M, Adholeya A, Ainsaar L, Ba A, et al. Global assessment of
786 arbuscular mycorrhizal fungus diversity reveals very low endemism. *Science*. 2015;349(6251):970-3.

787 97. Wu ZY, H S. The floristic geography of China. In: *Floristic and Vegetation Geography of China*
788 (ed. Chen LZ). Science Press, Beijing (in Chinese) 20q4.

789 98. Magoc T, Salzberg SL. FLASH: fast length adjustment of short reads to improve genome

790 assemblies. *Bioinformatics*. 2011;27(21):2957-63.

791 99. Edgar RC. UPARSE: highly accurate OTU sequences from microbial amplicon reads. *Nature*
792 *Methods*. 2013;10(10):996-+.

793 100. Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, et al. Introducing
794 *mothur*: Open-Source, Platform-Independent, Community-Supported Software for Describing and
795 Comparing Microbial Communities. *Appl Environ Microb*. 2009;75(23):7537-41.

796 101. Paul EA, Collins HP, Leavitt SW. Dynamics of resistant soil carbon of midwestern agricultural
797 soils measured by naturally occurring C-14 abundance. *Geoderma*. 2001;104(3-4):239-56.

798 102. Chen CR, Condon LM, Davis MR, Sherlock RR. Seasonal changes in soil phosphorus and
799 associated microbial properties under adjacent grassland and forest in New Zealand. *Forest Ecol*
800 *Manag*. 2003;177(1-3):539-57.

801 103. Don A, Kalbitz K. Amounts and degradability of dissolved organic carbon from foliar litter at
802 different decomposition stages. *Soil Biol Biochem*. 2005;37(12):2171-9.

803 104. Brogi SR, Jung JY, Ha S-Y, Hur J. Seasonal differences in dissolved organic matter properties
804 and sources in an Arctic fjord: Implications for future conditions. *Sci Total Environ*. 2019;694.

805 105. N.J. G, G.R. G. *Null models in ecology*. Smithsonian Institution Press, Washington, DC. 1996.

806 106. Fine PVA, Kembel SW. Phylogenetic community structure and phylogenetic turnover across
807 space and edaphic gradients in western Amazonian tree communities. *Ecography*. 2011;34(4):552-65.

808 107. Dini-Andreote F, Stegen JC, van Elsas JD, Salles JF. Disentangling mechanisms that mediate the
809 balance between stochastic and deterministic processes in microbial succession. *Proceedings of the*
810 *National Academy of Sciences of the United States of America*. 2015;112(11):E1326-E32.

811 108. Chave J, Jabot F. Estimation of neutral parameters by maximum likelihood. Université Paul
812 Sabatier. 2008.

813 109. Hubbell SP. The unified neutral theory of biodiversity and biogeography. Princeton University
814 Press. 2001;Princeton(NJ).

815 110. Etienne RS. A new sampling formula for neutral biodiversity. Ecology Letters.
816 2005;8(3):253-60.

817 111. Etienne RS, Alonso D. A dispersal-limited sampling theory for species and alleles. Ecology
818 Letters. 2005;8(11):1147-56.

819 112. Etienne RS, Latimer AM, Silander JA, Cowling RM. Comment on "Neutral ecological theory
820 reveals isolation and rapid speciation in a biodiversity hot spot". Science. 2006;311(5761):610B-+.

821 113. Etienne RS, Olf H. Confronting different models of community structure to species-abundance
822 data: a Bayesian model comparison. Ecology Letters. 2005;8(5):493-504.

823

824

825

826

827

828

829

830

831

832

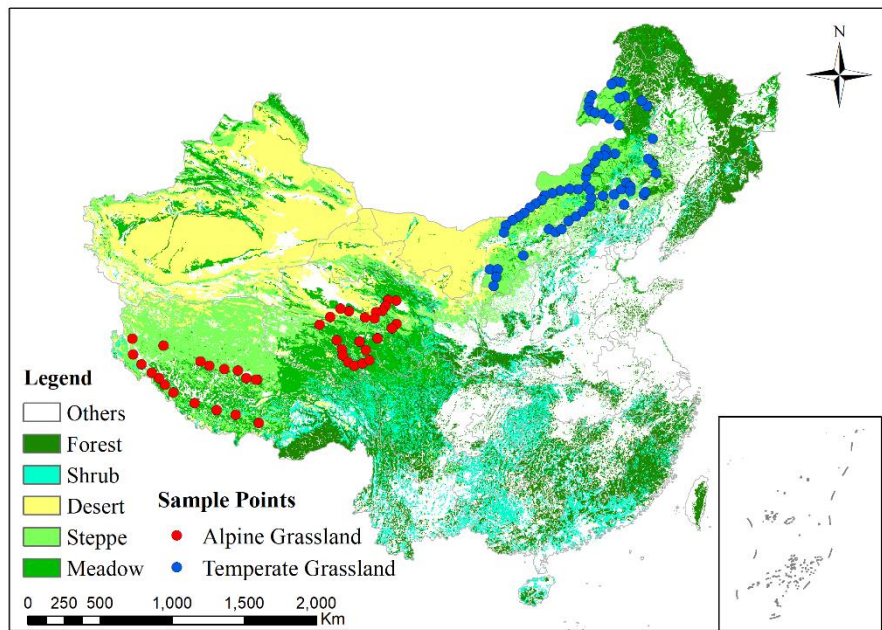
833

834

835

836

Supplementary Figures



837

838 **Figure 1 - figure supplement 1 Sampling sites across 1,921 km of the alpine grassland in**

839 **Qinghai-Tibet Plateau (in red) and 1,661 km of the temperate grassland in Inner Mongolia**

840 **(in blue).**

841

842

843

844

845

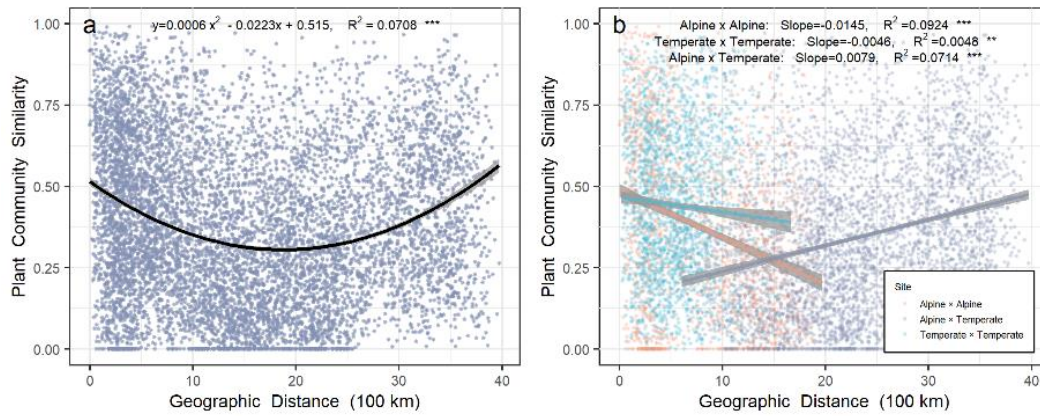
846

847

848

849

850



851

852 **Figure 1 - figure supplement 2 Relationship between plant community similarity over**

853 **geographic distance in northern grassland of China.** Light blue points are for pairwise sites in

854 the alpine grassland. Orange points are for pairwise sites in the temperate grassland. Grey points

855 are for pairwise sites between the alpine grassland cross temperate grassland. Grey shades stand

856 for 95% confidence interval.

857

858

859

860

861

862

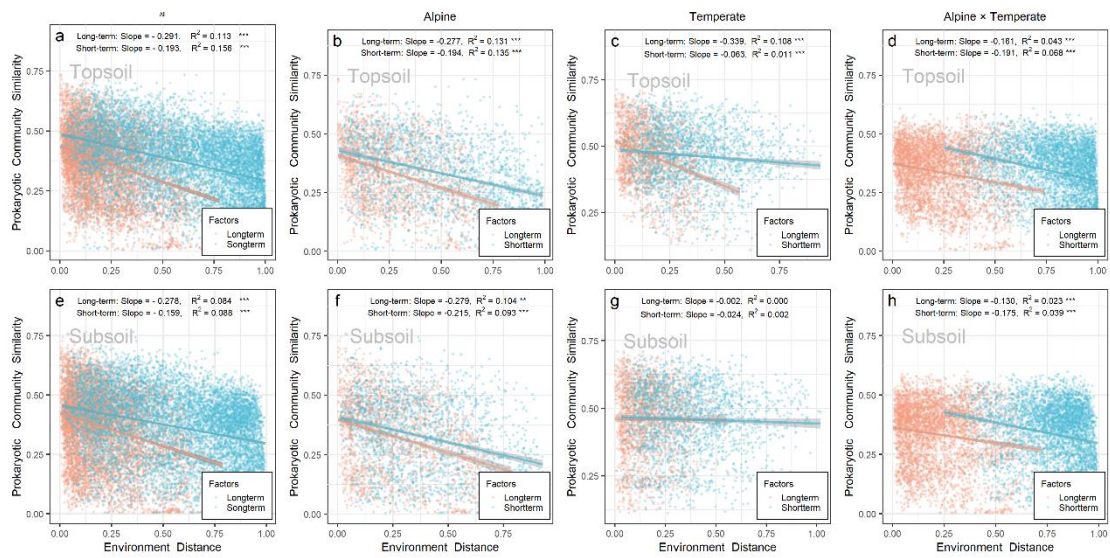
863

864

865

866

867



868

869 **Figure 2 - figure supplement 1 Distance decay relationship for prokaryotic community**

870 **similarity over relative long-term (orange points) and short-term (light blue points)**

871 **environmental factors. (a-d) topsoil; (e-h) subsoil. Shades stand for 95% confidence interval.**

872

873

874

875

876

877

878

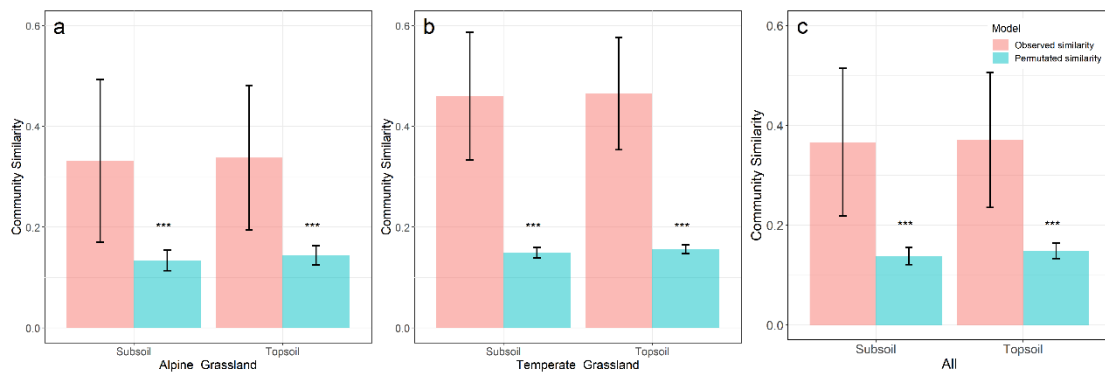
879

880

881

882

883



884

885 **Figure 3 - figure supplement 1 Null model analysis for community similarity. The null**

886 **model for soil prokaryotic community similarity from the alpine grassland (a), temperate**

887 **grassland (b) and all sites (c). The pink and blue stands for observed similarity and**

888 **permuted similarity by null model, respectively. “***” stand for the significance at 0.001**

889 **level.**

890

891

892

893

894

895

896

897

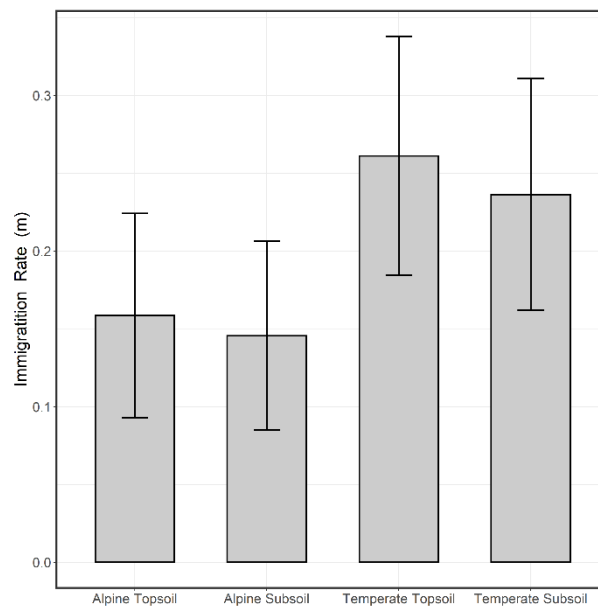
898

899

900

901

902



903

904 **Figure 3 - figure supplement 2 The immigration rate (m) of soil prokaryote community in**

905 **topsoil and subsoil of the alpine and temperate grassland biomes**

906

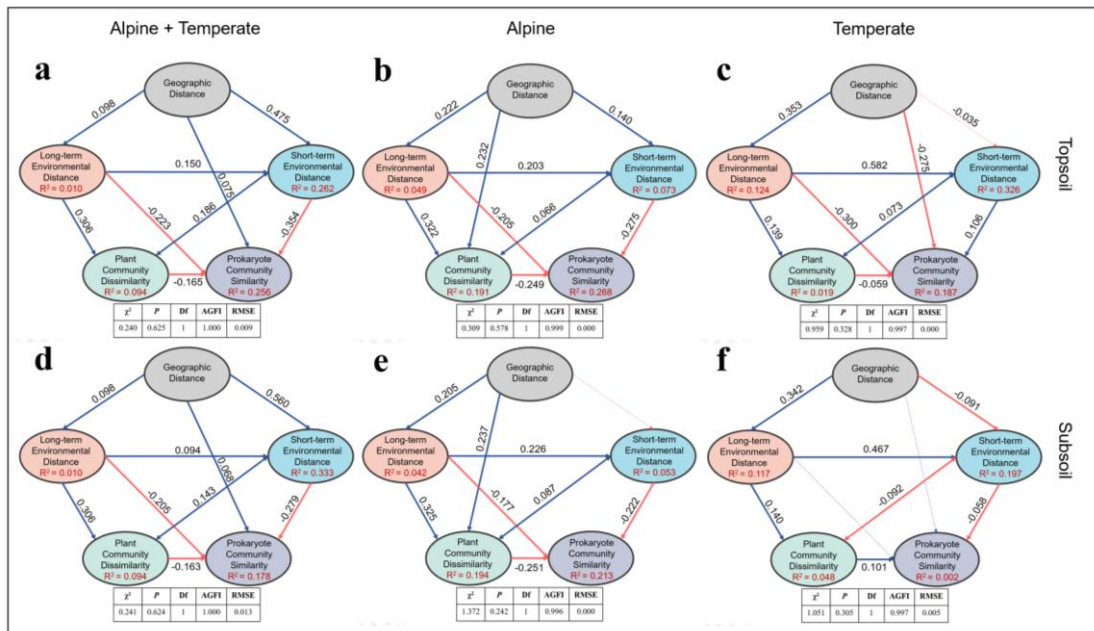
907

908

909

910

911
 912
 913
 914
 915
 916
 917



918

919 **Figure 4 - figure supplement 1 Structure equation model for geographic distance,**
 920 **long-term environmental distance, short-term environmental distance, and plant**
 921 **community dissimilarity based on Bray-Curtis distance in affecting soil prokaryote**
 922 **community similarity. (a) topsoil of the northern grassland; (b) topsoil of the alpine grassland;**
 923 **(c) topsoil of the temperate grassland; (d) subsoil of the northern grassland; (e) subsoil of the**
 924 **alpine grassland; (f) subsoil of the temperate grassland. Red lines stand for negative correlation**

925 and blue lines stand for positive correlation; Bold lines stand for significance at 0.05 level.

926

927

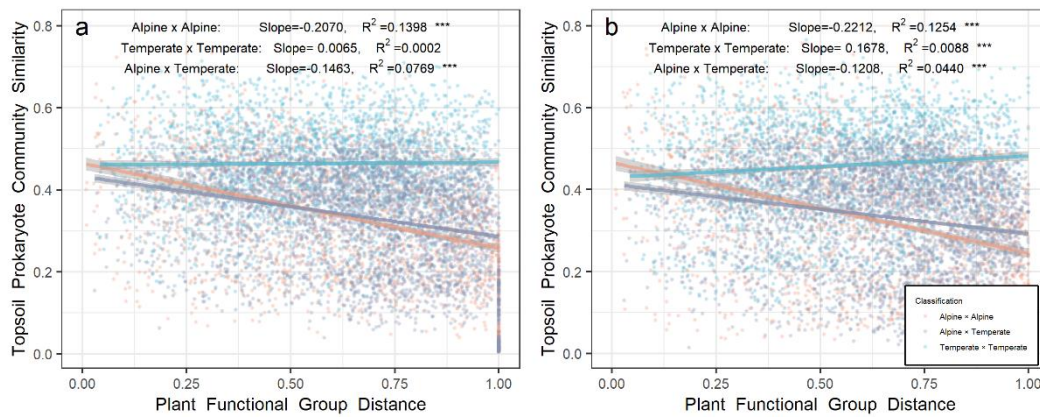
928

929

930

931

932



934

934 **Figure 1 - figure supplement 3 Relationship between prokaryote community over plant**

935 **community dissimilarity.** (a) topsoil; (b)subsoil. Light blue points are for pairwise sites in the

936 alpine grassland. Orange points are for pairwise sites in the temperate grassland. Grey points are

937 for pairwise sites between the alpine grassland cross temperate grassland. Grey shades stand for

938 95% confidence interval.

939

940

941

953

954

955

956

957

958

959

960

961

962 **Supplementary file 1** Mantel test between the prokaryotic community similarity based on

963 Bray-Curtis index and environment variables

964

965 **Supplementary file 2** Partial Mantel test between prokaryotic community similarity based on

966 Bray-Curtis index and environmental variables.

967