Impaired astrocytic $\text{Ca}^{2+}$ signaling in awake-behaving Alzheimer’s disease transgenic mice

Knut Sindre Åbjørsbråten†, Gry H. E. Syverstad Skaaraa†, Céline Cunen³,⁴, Daniel M. Bjørnstad¹,
Kristin M. Gullestad Binder¹, Laura Bojarskaite¹,⁵, Vidar Jensen¹, Lars N.G. Nilsson⁷, Shreyas B. Rao², Wannan Tang¹,⁶, Gudmund Horn Hermansen³, Erlend A. Nagelhus⁸, Ole Petter Ottersen⁸,
Reidun Torp², Rune Enger¹*

¹GliaLab at the Letten Centre, Division of Anatomy, Department of Molecular Medicine, Institute of Basic Medical Sciences, University of Oslo, N-0317 Oslo, Norway
²Division of Anatomy, Department of Molecular Medicine, Institute of Basic Medical Sciences, University of Oslo, N-0317 Oslo, Norway
³Statistics and Data Science group, Department of Mathematics, Faculty of Mathematics and Natural Sciences, University of Oslo, 0316, Oslo, Norway
⁴Department for Statistical Analysis, Machine Learning and Image Analysis, Norwegian Computing Center, 0314 Oslo, Norway
⁵Department of Neurology, Oslo University Hospital, 0027 Oslo, Norway
⁶Department of Clinical and Molecular Medicine, Norwegian University of Science and Technology, Trondheim, Norway
⁷Department of Pharmacology, University of Oslo, N-0316 Oslo, Norway
⁸Office of the President, Karolinska Institutet, Stockholm, Sweden

† These authors have contributed equally to this work and share first authorship

* Correspondence: Rune Enger, Division of Anatomy, Department of Molecular Medicine, Institute of Basic Medical Sciences, University of Oslo, P.O. Box 1103 Blindern, N-0317 Oslo, Norway. Email: rune.enger@medisin.uio.no.

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Abstract

Increased astrocytic Ca\(^{2+}\) signaling has been shown in Alzheimer's disease mouse models, but to date no reports have characterized behaviorally induced astrocytic Ca\(^{2+}\) signaling in such mice. Here, we employ an event-based algorithm to assess astrocytic Ca\(^{2+}\) signals in the neocortex of awake-behaving tg-ArcSwe mice and non-transgenic wildtype littermates while monitoring pupil responses and behavior. We demonstrate an attenuated astrocytic Ca\(^{2+}\) response to locomotion and an uncoupling of pupil responses and astrocytic Ca\(^{2+}\) signaling in 15-months old plaque-bearing mice. Using the genetically encoded fluorescent norepinephrine sensor GRAB\(_{NE}\) we demonstrate a reduced norepinephrine signaling during spontaneous running and startle responses in the transgenic mice, providing a possible mechanistic underpinning of the observed reduced astrocytic Ca\(^{2+}\) responses. Our data points to a dysfunction in the norepinephrine-astrocyte Ca\(^{2+}\) activity-axis, which may account for some of the cognitive deficits observed in Alzheimer’s disease.

Introduction

Since astrocytic Ca\(^{2+}\) signals were first discovered in the early 1990’s they have been the object of numerous studies exploring their roles in brain physiology and pathophysiology. Importantly, such signals have been shown to occur in response to a wide array of neurotransmitters and to trigger the release of substances that affect neuronal signaling and the vasculature. A growing body of evidence suggests that astrocytic Ca\(^{2+}\) signals play important roles in higher brain functions such as memory formation and cortical processing, mediated in part through the neuromodulatory systems (Adamsky et al. 2018; Kol et al. 2020; Poskanzer and Yuste 2016, 2011; Ye et al. 2020; Paukert et al. 2014).

Astrocytic Ca\(^{2+}\) signaling in an Alzheimer's disease (AD) mouse model was first described by Kuchibhotla et al. 2009 (Kuchibhotla et al. 2009), who found pathological Ca\(^{2+}\) waves originating at amyloid plaques, and a general increase in astrocytic Ca\(^{2+}\) signaling. Later, Dekate et al. showed plaque-associated astrocytic hyperactivity mediated through activation of metabotropic purine receptors (Delekate et al. 2014; Reichenbach et al. 2018). These studies were performed under anesthesia, which severely attenuates physiological Ca\(^{2+}\) signals (Thrane et al. 2012).

The field of astrocytic Ca\(^{2+}\) signaling is undergoing a revolution as developments in optical imaging and genetically encoded fluorescent sensors now allow us to monitor these signals in awake-behaving...
mice, without the confounding effects of anesthesia (Srinivasan et al. 2015; Bojarskaite et al. 2020). Such studies have revealed exceedingly rich and complex astrocytic Ca$^{2+}$ signaling ranging from large activations of nearly all astrocytes in a field-of-view (FOV) under locomotion and startle responses due to noradrenergic activity (Ding et al. 2013; Paukert et al. 2014), to small, localized signals occurring spontaneously or as a response to local neuronal activity (Bindocci et al. 2017; Stobart et al. 2018; Srinivasan et al. 2015). New analytical tools now also enable us to accurately quantify and describe these signals (Y. Wang et al. 2019; Bjørnstad et al. 2021).

The brain noradrenergic system is crucial for mediating responses to external environmental stimuli, optimizing CNS performance and thus for arousal and cognition. Interestingly, norepinephrine is one of the main drivers of astrocytic Ca$^{2+}$ signaling activating astrocytic α1-adrenergic receptors (Ding et al. 2013; Thrane et al. 2012; Paukert et al. 2014). Notably, noradrenergic signaling in relation to locomotion and startle responses causes global increases of astrocytic Ca$^{2+}$ (Ding et al. 2013; Srinivasan et al. 2015). The main downstream effects of this astrocytic Ca$^{2+}$ activity is not yet well understood, but thought to support neurons metabolically (O'Donnell et al. 2012), or through dynamic changes of the size and composition of the extracellular fluids (F. Wang et al. 2012). Nonetheless, astrocytes are believed to be key actuators of the noradrenergic system (O'Donnell et al. 2012; Wahis and Holt 2021). Importantly, NA signaling is believed to play a pivotal role in AD pathology (Weinshenker 2018; Peterson and Li 2018; Holland, Robbins, and Rowe 2021). Profound loss of noradrenergic neurons is a hallmark of AD that occurs early in the development of the disorder, likely accounting for some of the symptoms of AD, including loss of cognitive function (Matchett et al. 2021). However, the mechanisms/signaling pathways connecting perturbed NA and cognition are only rudimentarily understood.

As astrocytes are tightly associated with NA signaling and AD is characterized by perturbed NA system, we set out to investigate whether the norepinephrine–astrocyte Ca$^{2+}$-signaling axis was perturbed in ~15 month old unanesthetized awake-behaving tg-ArcSwe mice compared to wildtype (WT) littermates. The tg-ArcSwe mice carry two mutations in the amyloid precursor protein gene, the Arctic (E693G) and Swedish (KM670/6701NL) mutations, and exhibit amyloid-β (Aβ) deposits, a hallmark of AD (Lord et al. 2006; Yang et al. 2011; Lillehaug et al. 2014; Philipson et al. 2009). The mice have a robust phenotype with emergence of Aβ plaques at 6–7 months of age both in the parenchyma and in the walls of blood vessels, that are biochemically similar to human Aβ plaques (Philipson et al. 2009). The prominent cerebral amyloid angiopathy in close proximity to astrocytic endfeet makes the model suitable for studying astrocytic signaling (Yang et al. 2011).

We demonstrate an attenuated behaviorally induced Ca$^{2+}$ signaling in cortical astrocytes during locomotion. As noradrenergic signaling is known to trigger astrocytic Ca$^{2+}$ signaling in startle
responses and locomotion, and pupil responses are regarded a faithful, although indirect, readout of noradrenergic signaling in the brain during physiological conditions (Reimer et al. 2016; Zuend et al. 2020; Costa and Rudebeck 2016), we compared the pupil responses to the astrocytic \( \text{Ca}^{2+} \) signals during running and startle and found a positive correlation in WT mice. No such correlation was present in the AD mice. Using the fluorescent extracellular norepinephrine sensor GRABNE (Feng et al. 2019) we found attenuated norepinephrine signaling in relation to locomotion and startle responses in the AD mice, likely explaining the observed reduced astrocytic \( \text{Ca}^{2+} \) signaling. Such perturbed behaviorally induced norepinephrine-linked astrocytic \( \text{Ca}^{2+} \) signaling may account for some of the cognitive deficiencies observed in AD patients.

RESULTS

Widespread reactive astrogliosis in tg-ArcSwe mice

AD transgenic mouse models with high levels of Aβ deposition exhibit reactive astrogliosis (Rodríguez-Arellano et al. 2016). As astrogliosis might affect astrocytic \( \text{Ca}^{2+} \) activity (Shigetomi et al. 2019; Sano et al., n.d.) and has not been quantified in neocortex of tg-ArcSwe mice yet, we measured GFAP expression by immunofluorescence and mRNA levels, and performed morphometrical analyses of astrocytes in tg-ArcSwe mice and WT littermates (Fig. 1).

We found a strong increase in levels of \( Gfap \) mRNA (81.58 ± 19.56 for WT; 512.13 ± 66.95 for tg-ArcSwe, \( p < 0.0001, \ n = 8 \) animals in both groups) in tg-ArcSwe compared to WT littermates (Fig. 1C). This was supported by a significantly higher GFAP labeling fraction in tg-ArcSwe animals compared to WT littermates (Fig. 1B, 24.03% ± 4.80% in tg-ArcSwe, \( n = 6 \) mice vs. 4.77% ± 1.58% in WT, \( n = 4 \) mice, \( p = 0.02 \)). Morphometric analyses displayed a significantly higher number of labelled astrocytic processes (38.63 ± 3.61 in tg-ArcSwe, \( n = 6 \) mice; 23.84 ± 3.72 in WT, \( n = 4 \) mice; \( p = 0.02 \)), as well as total length (837.9 µm ± 55.9 µm for tg-ArcSwe; 506.0 µm ± 62.5 µm for WT; \( p = 0.0095 \)) and volume of processes (990.9 µm³ ± 93.8 µm³ for tg-ArcSwe; 401.5 µm³ ± 82.9 µm³ for WT; \( p = 0.0095 \)) in tg-ArcSwe mice compared to WT littermates (Fig. 1D). In addition, astrocytes in the tg-ArcSwe mice exhibited significantly more branching points at the distance 16–32 µm from the nucleus (\( p < 0.0001 \)).

Two-photon imaging of awake-behaving tg-ArcSwe mice
To characterize astrocytic Ca\textsuperscript{2+} signaling in awake tg-ArcSwe mice and nontransgenic littermates we employed two-photon microscopy of cortical layer 1–3 astrocytes in the somatosensory cortex expressing GCaMP6f. Recombinant adeno-associated virus was used to deliver the genetic construct, and the glial fibrillary acidic protein (GFAP) promoter was used to target astrocytes (Fig 2A). Aβ plaques were visualized in vivo by methoxy-X04 delivered by intraperitoneal injection (Fig. 2A). Methoxy-X04 enters the brain and specifically stains parenchymal Aβ plaques and cerebrovascular deposits (Klunk et al. 2002), and is widely used for in vivo imaging of transgenic mice with Aβ (Delekate et al. 2014; Kuchibhotla et al. 2009; Meyer-Luehmann et al. 2008). Both tg-ArcSwe mice and littermates were injected with methoxy-X04 to rule out any confounding effects of methoxy-X04 on astrocytic Ca\textsuperscript{2+} activity. Imaging was performed at ~30 Hz frame rate to capture fast populations of astrocytic Ca\textsuperscript{2+} transients with simultaneous surveillance video recording of mouse behavior, movement of the treadmill as well as pupil diameter (Fig. 2B) to monitor the level of arousal (Reimer et al. 2016). The mice were allowed to spontaneously move on a custom-built disc shaped treadmill, and all mice exhibited both periods of quiet wakefulness (absence of locomotion), and running (Fig. 2-figure supplement 1). Astrocytic Ca\textsuperscript{2+} signals were analyzed using a newly developed event-based Ca\textsuperscript{2+} signal analysis toolkit, outlining so-called regions-of-activity (ROAs), combined with manually drawn regions-of-interests (ROIs) defining astrocytic subcompartments (Bjørnstad et al. 2021; Bojarskaite et al. 2020). This method detects Ca\textsuperscript{2+} signals in a pixel-by-pixel fashion by (1) estimating the noise level per pixel over time, (2) binarizing the data into signal/non-signal by noise level-based thresholds per pixel, and (3) connecting adjoining active pixels in space and time results in \textit{x-y-t} ROAs. For the remainder of this article, we primarily report the \textit{ROA density}, which is the fraction (in %) of the compartment analyzed with signal at any given time. See Materials and Methods for more details.

The potential differences in astrocytic Ca\textsuperscript{2+} signaling between the two genotypes during spontaneous runs and startle responses, as well as the relationship between Ca\textsuperscript{2+} signaling and pupil dilation, were investigated through statistical models. These results are presented in the following sections, and the methods are described in detail in Statistical Analyses in Materials and Methods. The Ca\textsuperscript{2+} signal responses were modeled by linear mixed effect regression models. The coefficients of primary interest were the effect of genotype, and the interaction between the effect of pupil dilation and genotype (i.e. the parameter indicating potential differences in the effect of pupil dilation on Ca\textsuperscript{2+} signaling between the two genotypes). In addition, the models included some technical and biological covariates, like the \textmu m per pixel (images acquired with two different levels of magnification), the depth of the measurements and the maximal speed of mouse locomotion in the time window studied. Further, the models include random intercepts for each mouse. The sensitivity of our result to these modeling choices were assessed by various robustness checks, primarily by iteratively removing individual
mice and trials to see to which degree this influenced p-values and estimates (see Statistical Analyses in Materials and Methods and Fig. 2-figure supplement 2)).

Astrocytes close to Aβ plaques express GCaMP6f

The tg-ArcSwe mice were imaged at ~15 months of age. At this age they present with Aβ plaques throughout the cortical mantle, and score poorly on behavioral tasks (Codita et al. 2010; Lillehaug et al. 2014; Lord et al. 2006)(Fig. 2C). Aβ plaques were characterized by loss of cells and severely perturbed tissue morphology, including autophagic vacuoles (Fig 2C). Even so, relatively normal cellular morphology was present at short distances away from Aβ plaques, and astrocytes faithfully expressed the GCaMP6f Ca\(^{2+}\) sensor 3 weeks after viral transduction (Fig. 2A,C, Fig. 2-figure supplement 3). Of note, GCaMP6f expression as judged by average GFP labeling and area fraction of GFP labeling in confocal micrographs were similar in the two groups (Fig. 2-figure supplement 3).

Astrocytic Ca\(^{2+}\) signals in quiet wakefulness are preserved in tg-ArcSwe mice

In quiet wakefulness (defined as absence of locomotion), the gross level of astrocytic Ca\(^{2+}\) signaling was similar in mutant mice and their littermates as measured by ROA frequency, ROA density (the active fraction of a compartment) as well as event size and duration in the full FOV and across the different astrocytic subcompartments (Fig. 2C and Fig. 2-figure supplement 4). We were neither able to detect a clear correlation in overall astrocytic Ca\(^{2+}\) signaling measured by ROA density and the distance from nearest Aβ plaque (in 3D) (slope = -0.00020, Fig. 2E). However, we found some examples of long-lasting pathological Ca\(^{2+}\) waves as reported previously in anesthetized mice (Delekate et al. 2014; Kuchibhotla et al. 2009). Such Ca\(^{2+}\) waves were found in 10–15% of recordings from tg-ArcSwe mice (Fig. 2F), and the number of such clear pathological events was very low compared to the overall astrocytic Ca\(^{2+}\) signaling.

Uncoupling between pupil dilation and astrocytic Ca\(^{2+}\) responses during spontaneous running in tg-ArcSwe mice

Locomotor behavior is known to be strongly correlated with astrocytic Ca\(^{2+}\) signalling (Paukert et al. 2014; Bojarskaite et al. 2020; Srinivasan et al. 2015), putatively through the activation of the noradrenergic and cholinergic neuromodulatory systems in conjunction with local network activity (Kjaerby et al. 2017). To investigate if the physiological astrocytic Ca\(^{2+}\) responses were preserved in the tg-ArcSwe mice, they were allowed to move freely on a custom built disc-shaped treadmill (Bojarskaite et al. 2020). All mice exhibited both running and behavioral quiescence, and the level of running between the two genotypes were comparable (Fig. 2-figure supplement 1). Running was
accompanied by an increase in pupil size and a brisk increase in astrocytic Ca\textsuperscript{2+} signaling typically involving most of the astrocytes in the FOV in both genotypes (Fig. 3A). Overall, astrocytic Ca\textsuperscript{2+} responses to spontaneous locomotion were somewhat reduced in tg-ArcSwe mice compared to WT littermates: When astrocytic Ca\textsuperscript{2+} signals were analyzed using a linear mixed effects regression model (Table 1), a lower ROA density rise rate was found in tg-ArcSwe mice when assessing the full FOV (0.31 in WT vs. 0.20 in tg-ArcSwe, p = 0.032), and astrocytic processes (0.32 in WT vs. 0.20 in tg-ArcSwe, p = 0.032), whereas the Ca\textsuperscript{2+} responses were not significantly different in astrocytic somata and endfeet (0.46 in WT vs. 0.39 in tg-ArcSwe, p = 0.23, and 0.35 in WT vs. 0.30 in tg-ArcSwe, p = 0.11, respectively)(Fig. 3C). Max ROA density values in WT vs. tg-ArcSwe were significantly different when assessing the full FOV (0.63 vs. 0.42, p = 0.033), near significantly different when assessing astrocytic processes and endfeet (0.72 vs. 0.53, p = 0.053 for processes and 0.69 vs. 0.51, p = 0.068 for endfeet), and not significantly different for astrocytic somata (0.81 vs. 0.71, p = 0.25)(Fig. 3C). For mean ROA density values, see Fig. 3-figure supplement 1.

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Table 1. Estimated statistical model for ROA density rise rate in the FOV during spontaneous running.

μm per pixel has a small/ no effect, but we include it for consistency. Deeper measurements tend to have lower ROA density rise rate. Given that the mice are running, a higher speed does not appear to be associated with a higher ROA density rise rate (n=109, in 10 mice).

Pupil responses are known to be a faithful indirect indicator of activity in the locus coeruleus in mice, even though also the cholinergic neuromodulatory system plays a role for sustained pupil dilation (Reimer et al. 2016). As norepinephrine is known to be a potent trigger of astrocytic Ca\textsuperscript{2+} signaling (Bekar, He, and Nedergaard 2008; Srinivasan et al. 2015; Paukert et al. 2014), one would expect to find a correlation between pupil dilations and astrocytic Ca\textsuperscript{2+} signals. This was indeed the case for WT mice (Table 2): When comparing ROA density rise rate and pupil dilation, we found a positive slope in line with previous reports (Zuend et al. 2020) (Fig 3D). In transgenic mice, this correlation was lost, or even reversed (Fig. 3D, Table 2: slope of 0.80 in WT vs. -0.35 in tg-ArcSwe, p = 0.007), demonstrating an uncoupling between pupil responses and astrocytic Ca\textsuperscript{2+} signaling in tg-ArcSwe.
mice. Similarly, when assessing max ROA density, we found a positive slope in WT, which was lost in tg-ArcSwe (Fig. 3D, 2.06 in WT vs. -0.58 in tg-ArcSwe, p = 0.00039). Similar slopes were found when comparing mean ROA density vs. pupil dilation Fig. 3-figure supplement 1, panel C.

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Table 2. Estimated statistical model for ROA density rise rate in the FOV and pupil dilation during spontaneous running. When including pupil dynamics in the model (cf. Table 1 without pupil dynamics), the effect of genotype was no longer significant. However, the relationship between pupil dilation and ROA density rise rate was different in the two genotypes, with the WT mice having a significantly higher slope. μm per pixel and depth of the recording had no significant effect on the model, but we included them for consistency. The slope for the WT group is found by adding the slope in the TG group with the interaction term (e.g. -0.3537 + 1.1562 = 0.80) (n=44, in 10 mice).

Uncoupling between pupil dilation and astrocytic Ca\textsuperscript{2+} responses during startle in tg-ArcSwe mice

Another main trigger for astrocytic Ca\textsuperscript{2+} signals are startle responses that also are mediated through an activation of the noradrenergic system (Ding et al. 2013; Srinivasan et al. 2015). Even though typically triggering running, the startle response could also trigger freezing behavior and is thought to activate different subcortical networks than spontaneous locomotor behavior (Caggiano et al. 2018; Ferreira-Pinto et al. 2018; Grillner and El Manira 2020). To investigate if the startle-evoked astrocytic Ca\textsuperscript{2+} responses were preserved in the tg-ArcSwe mice, they were subjected to 10 air puffs delivered at 10 Hz directed to the vibrissa, nasal and facial region contralaterally to the recording side once per trial at 150 s in a 300 s two-photon imaging recording. Trials in which the mouse was spontaneously running at or immediately before the air puff were excluded from the analyses. We found no signs of habituation to the stimulus in terms of behavioral response (Fig. 4–figure supplement 1). Interestingly, tg-ArcSwe mice were more prone to start running during startle responses than WT littermates (Fig. 4A vs. 4B), consistent with previous reports of enhanced startle response in other mouse models of
AD (McCool et al. 2003). The level of pupil dilation was however similar in the two genotypes (0.17 vs. 0.12 relative increase in pupil size in WT vs. tg-ArcSwe, respectively, p = 0.36, 86 trials) (Fig. 4A and 4B). Regarding astrocyte Ca^{2+} response to startle, we found no differences between the genotypes. When modeled with a mixed effects linear regression model (Table 3), for ROA density rise rate, Ca^{2+} responses in the full FOV and in all astrocytic subcompartments were statistically similar between the genotypes (FOV: 0.34 in WT vs. 0.23 in tg-ArcSwe, p = 0.09; somata: 0.25 in WT vs. 0.23 in tg-ArcSwe, p = 0.8; processes: 0.42 in WT vs. 0.31, p = 0.09; endfeet: 0.36 in WT vs. 0.25, p = 0.10 (Fig. 4C). Similarly, max ROA density was similar for the full FOV (0.36 in WT vs. 0.35 in tg-ArcSwe, p = 0.40), astrocytic somata (0.37 in WT vs. 0.35 in tg-ArcSwe, p = 0.83), astrocytic processes (0.41 in WT vs. 0.34 in tg-ArcSwe, p = 0.39), and astrocytic endfeet (0.41 in WT vs. 0.35 in tg-ArcSwe, p = 0.48) (Fig. 4C). For mean ROA density values, see (Fig. 3–figure supplement 1).

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Table 3. Estimated statistical model for ROA density rise rate in the FOV during startle. Depth from the cortical surface and maximal speed during the startle response had a significant effect in the model, whereas the μm per pixel (level of magnification of the image recording) had no significant effect (n=117 trials, in 11 mice).

However, the relationship between astrocytic Ca^{2+} responses and pupillary responses were different in the two genotypes (Fig. 4D). WT mice displayed a clear positive slope, while tg-ArcSwe exhibited a slope close to zero (0.91 vs. -0.013 p = 0.00043 for ROA density rise rate, and 1.27 vs. 0.083 p = 0.0043 and max ROA density, respectively), suggesting a potential uncoupling between pupillary and astrocytic Ca^{2+} responses during startle in tg-ArcSwe mice, similar to what was observed during spontaneous running (Fig. 3D). Similar slopes were found when comparing mean ROA density vs pupil dilation (Fig. 3–figure supplement 1).

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When including the pupil dilation in the statistical model, the two genotypes are clearly different. Moreover, depth from the cortical surface had a significant effect in the model, whereas optical zoom and maximum speed had a non-significant effect (n=95 trials, in 11 mice). The slope for the WT group is found by adding the slope in the TG group with the interaction term (e.g. -0.0129 + 0.9235 = 0.91).

**Attenuated noradrenergic signaling in tg-ArcSwe mice**

To investigate the cause of our observed attenuated astrocytic Ca\(^{2+}\) responses during locomotion and startle responses, and uncoupling of pupil responses and astrocytic Ca\(^{2+}\) responses, we took advantage of the newly developed genetically encoded fluorescent extracellular norepinephrine sensor, the GRAB\(_{NE}\) sensor (Feng et al. 2019). The sensor was delivered by rAAV transduction of the GRAB\(_{NE}\) (version 1m) construct and targeted to membranes of neurons using the hSyn1 promoter. The mice were subjected to the same imaging protocols as for Ca\(^{2+}\) imaging, and we recorded norepinephrine fluorescence, pupil dynamics and locomotor activity. Differences between the genotypes were modeled by linear mixed effects regression models as outlined for Ca\(^{2+}\) imaging (results presented in Tables 5 and 6). The coefficients modeled were the effect of genotype, and the interaction between the effect of pupil dilation and genotype (i.e. the parameter indicating potential differences in the effect of pupil dilation on norepinephrine signaling between the two genotypes). During both spontaneous running and startle responses we observed increases in norepinephrine fluorescence in WT mice (Fig. 5). This effect was less prominent in tg-ArcSwe mice, and similar to Ca\(^{2+}\) imaging, we found a decoupling between pupil responses and noradrenergic signaling. In WT mice, for spontaneous runs, the relative increase in pupil diameter explained a large proportion of change in norepinephrine signaling (Table 5). In tg-ArcSwe mice, pupil responses and norepinephrine signaling were less correlated. The results for startle responses were similar (Table 6). These findings suggest that the reduced Ca\(^{2+}\) signaling and uncoupling between pupil responses and astrocytic Ca\(^{2+}\) signaling in tg-ArcSwe mice could be due to attenuated noradrenergic release in cortex and lower correlation between pupil responses and noradrenergic signaling.

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**Table 4. Estimated statistical model for ROA density rise rate in the FOV and pupil dilation during startle.**
Table 5. Estimated statistical model for change in GRAB$_{NE}$ fluorescence and pupil dilation during spontaneous runs. The coupling between pupil responses and noradrenergic signaling is weaker in tg-ArcSwe mice compared to WT littermates ($p = 0.043$, $n=100$ running episodes, 43 trials, in 7 mice). The slope for the WT group is found by adding the slope in the TG group with the interaction term (e.g. $0.0030 + 0.043 = 0.046$).

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Table 6. Estimated statistical model for change in GRAB$_{NE}$ fluorescence and pupil dilation during startle. The coupling between pupil responses and noradrenergic signaling during startle responses is weaker in tg-ArcSwe mice compared to WT littermates ($p = 0.037$, $n=59$ trials, in 7 mice). The slope for the WT group is found by adding the slope in the TG group with the interaction term (e.g. $0.0078 + 0.037 = 0.045$).
Discussion

Astrocytic Ca\textsuperscript{2+} signaling is emerging as a key component of signal processing in the brain and figures prominently in brain state transitions and memory formation (Poskanzer and Yuste 2016, 2011; Adamsky et al. 2018; Bojarskaite et al. 2020; Vaidyanathan et al. 2021). Aberrant astrocytic Ca\textsuperscript{2+} signaling could hence be implicated in the perturbed cognition seen in dementia. Indeed, previous studies have shown increased astrocytic Ca\textsuperscript{2+} signaling and spreading pathological Ca\textsuperscript{2+} waves in AD mouse models (Kuchibhotla et al. 2009; Delekate et al. 2014; Reichenbach et al. 2018). Based on these studies in anesthetized animals and other studies (Haughey and Mattson 2003; Lim et al. 2013; Abramov, Canevari, and Duchen 2004, 2003; Verkhratsky 2019) the current concept is that aberrant signaling to some degree is spatially coupled to amyloid deposits – the pathological hallmark of Alzheimer's disease.

Methodological advances now allow astrocytic Ca\textsuperscript{2+} signals to be studied in awake animals. Benefitting from this opportunity we show that tg-ArcSwe mice sustain a pattern of behaviorally induced astrocytic Ca\textsuperscript{2+} signaling similar to that found in littermate controls. However, the signals are weaker than in controls and do not display the correlation with pupil responses typically seen in WT animals. The behaviorally induced Ca\textsuperscript{2+} signals bear no clear spatial correlation to Aβ plaques. We conclude that elimination of anesthesia unveils a new dimension of Ca\textsuperscript{2+} signaling, superimposed on the locally induced signals described in previous studies. The uniform attenuation of the behaviorally induced signals in tg-ArcSwe mice and their uncoupling from pupil responses point to a potential perturbation of neuromodulatory control.

Astrocytic Ca\textsuperscript{2+} signaling in awake behaving mice is dominated by norepinephrine-induced astrocytic Ca\textsuperscript{2+} signals across the cortical mantle in relation to locomotor or startle responses (Srinivasan et al. 2015; Ding et al. 2013). The downstream effects of these synchronized, global Ca\textsuperscript{2+} signals coupled to arousal are not fully understood, but they may play a role in altering the levels of extracellular K\textsuperscript{+} (F. Wang et al. 2012), release of gliotransmitters such as glutamate or ATP (Kjaerby et al. 2017; Haydon and Nedergaard 2014) or metabolic supply (Zuend et al. 2020). Independent of what are the exact downstream mechanisms, there are reasons to believe that astrocytes serve as actuators of the noradrenergic neuromodulatory system, presumably exerting noradrenergic effects on neural network processing and ultimately affecting cognitive function (Ye et al. 2020; Holland, Robbins, and Rowe 2021; Poskanzer and Yuste 2016).

The cause of perturbed astrocytic Ca\textsuperscript{2+} responses during locomotion responses in AD mice seems to at least partially be explained by a blunted norepinephrine increase upon startle and locomotion in the tg-ArcSwe mice as evaluated by GRAB\textsubscript{NE} fluorescence. The primary norepinephrine nucleus in the brain, locus coeruleus, is known to be affected early in AD, both in humans and in animal models (Weinshenker 2018; Jacobs et al. 2021; Braak and Del Tredici 2011), and perturbed noradrenergic
function is likely an important factor both in disease progression in AD patients and accounting for
the cognitive decline of AD patients (Weinshenker 2018; Peterson and Li 2018; Holland, Robbins,
and Rowe 2021). Restoring noradrenergic signaling in AD mice with perturbed noradrenergic
signaling pharmacologically or by pharmacogenetics have been demonstrated to be beneficial
(Holland, Robbins, and Rowe 2021). In our study we find a largely retained level of pupil response in
the AD mice during startle responses, and a similar dynamic range of pupil responses (Fig. 4-figure
supplement 2) which under physiological circumstances would suggest a functional noradrenergic
system, as a strong correlation between locus coeruleus activity and pupil responses have been
established (Reimer et al. 2016; Costa and Rudebeck 2016). The underlying connectivity that enables
the pupils to be faithful readouts of the noradrenergic system is to the best of our knowledge not fully
established, even though spinal projections from locus coeruleus have been demonstrated (Hancock
and Fougerousse 1976; Costa and Rudebeck 2016; Liu et al. 2017). Our finding of a decoupling
between norepinephrine mediated astrocytic Ca\(^{2+}\) signaling and arousal-induced pupillary dilation,
could be due to perturbed connectivity between locus coeruleus and the relevant nuclei or projections
controlling activity in the superior cervical ganglion and consequently the sympathetic projections to
the pupils.

Given the clear differences in norepinephrine signaling between the two genotypes, one would maybe
have expected even more pronounced differences in astrocytic Ca\(^{2+}\) signaling. This discrepancy could
possibly be explained by either the lack of sensitivity of the GRAB\(_{NE}\) sensor in the current
experimental paradigm to pick up small increases in cortical norepinephrine, or potentially by a
compensatory upregulation of adrenergic receptors or other receptors in reactive astrocytes in the tg-
ArcSwe mice. In line with this Reichenbach et al. demonstrated an age/disease stage-related
upregulation of P2R1-receptors in an AD mouse model that correlated with the level of reactive
astrogliosis (Reichenbach et al. 2018). Moreover, potentially the expression of \(\alpha_1\)-adrenergic
receptors in astrocytes is changed in AD, similar to reports of how \(\alpha_2\) and \(\beta_2\) receptors are
upregulated in microvessels and neurons in AD patients, respectively (R. N. Kalaria et al. 1989;
Rajesh N. Kalaria and Harik 1989). Future studies probing the expression levels and patterns of
adrenergic receptors and downstream signaling pathways in astrocytes in AD models are warranted.

Although still sparsely investigated, both attenuated and increased Ca\(^{2+}\) signaling have been
demonstrated in various models of reactive astrogliosis, but so far without a clear understanding of the
mechanisms involved (Shigetomi et al. 2019). Widespread reactive astrogliosis is nonetheless highly
likely to perturb the physiological signaling of astrocytes, as prominent changes in the expression of
key receptors and molecules of intracellular pathways are known to occur (Habib et al. 2020; Escartin
et al. 2021). In this study we employed 15 months old tg-ArcSwe mice and WT littermates. These
mice exhibit a relatively high plaque burden, and represent a relatively advanced stage of AD. It is
possible that the attenuated astrocytic Ca\(^{2+}\) responses we observe is something found in advanced
disease, and not earlier in the disease development. Similarly, the sparsity of pathological
‘hyperactive’ astrocytic Ca^{2+} signaling events in our data compared to previous studies (Delekate et al. 2014; Kuchibhotla et al. 2009; Reichenbach et al. 2018), could stem from the other studies being performed at ages that represents earlier stages in the disease progression. Further studies across AD models and stages of disease progression mice are warranted to conclusively establish these conclusions.

Previous studies have shown mixed results regarding an effect of plaque-astrocyte distance, with reports of both hyperactivity close to plaques (Delekate et al. 2014) and no such correlation except for plaques serving as initiation sites for pathological intercellular Ca^{2+} waves (Kuchibhotla et al. 2009). In our awake-behaving mice, we found no clear correlation between overall Ca^{2+} activity and 3D distance reconstruction of plaque positions relative to the imaging plane (Fig. 2E). The apparent discrepancies in the literature and the present study may be due to different AD mouse models, different age groups investigated, different Ca^{2+} indicators employed or lastly, the effects of removing anesthesia allowing for a much richer repertoire of behaviorally induced astrocytic Ca^{2+} signaling to emerge, effectively masking a potential weak correlation.

Astrocytes have a highly complex and specialized morphology, and this morphology is known to change in reactive astrogliosis (Escartin et al. 2021). The majority of astrocytic processes are much smaller than what can be clearly delineated by non-super resolution optical microscopy, but to what extent these small processes are altered in reactive astrogliosis is unknown, although the astrocytic territories are known to be preserved (Wilhelmsson et al. 2006). Moreover, GFAP, which was used as the promoter for the GCaMP6f expression was highly upregulated in tg-ArcSwe mice. This difference in GFAP expression did not translate into obvious differences in sensor expression levels as judged by GFP labeling of brain slices (Fig. 2 – figure supplement 3). We cannot rule out that gliosis-induced morphological changes in combination with potential subtle differences in GCaMP6f expression and potentially an elevated baseline intracellular Ca^{2+} concentration (Kuchibhotla et al. 2009) may influence our results. However, the ability for the whole FOV to be activated during spontaneous running and startle responses, and the lack of correlation (if anything a negative correlation) between distance to the nearest Aβ plaque and gross level of Ca^{2+} signaling suggests that our findings are not due to degree or pattern of sensor expression.

The present study underscores the importance of studying astrocytic Ca^{2+} signals in unanesthetized mice, and to carefully consider animal behavior when interpreting astrocytic Ca^{2+} dynamics. By lifting the confounding effects of anesthesia, we found that the astrocytic hyperactivity previously reported in Alzheimer’s disease mouse models was only a part of the total picture. At first glance, the physiological Ca^{2+} responses were remarkably well preserved, and not characterized by a general increase in astrocytic Ca^{2+} signaling. However, behavior like quiet wakefulness and locomotion are
not static entities, and the degree of activation of all relevant parameters needs to be taken into account with statistical modeling to be able to conclude if there are relevant differences between the genotypes. We were able to demonstrate attenuated Ca\(^{2+}\) dynamics and norepinephrine signaling in the tg-ArcSwe mice, and given the growing spectrum of roles ascribed to the norepinephrine-astrocyte Ca\(^{2+}\) signaling in higher brain functions, the present findings may highlight one potential cause for the cognitive decline of AD patients.

MATERIALS AND METHODS

Animals

Tg-ArcSwe mice carry a human \(APP\) cDNA with the Arctic (p. E693G) and Swedish (p. KM670/671NL) mutations where the human \(\alpha\)PP gene is inherited only from male mice to ensure a more uniform onset of \(\alpha\) deposition (Lillehaug et al. 2014). The transgenic animals develop parenchymal \(\alpha\) plaques from 6 months of age, and cerebral amyloid angiopathy (CAA) from 8 months of age (Yang et al. 2011; Lord et al. 2006). 6 tg-ArcSwe mice and 5 WT littermates (both males and females) were used in this study. Genotyping was done as previously described with primers annealing to the Thy1-promoter and the human \(APP\)-transgene (Lord et al 2006). The animals were housed under standard conditions with 12-hour dark-light cycles and unrestricted access to food and water. All animal procedures were in accordance with the National Institutes of Health Guide for the care and use of laboratory animals and approved by the Norwegian Food Safety Authority (project number: FOTS #11983).

Viral transduction and delivery of fluorophores

Serotype 2/1 recombinant adeno-associated virus (rAAV) from plasmid construct pAAV-GFAP-GCaMP6f (Chen et al. 2013) was generated (rAAV titers about 1.0–6.0 \(\times\) 10\(^{12}\) viral genomes/mL) and used for visualizing astrocytic Ca\(^{2+}\) signaling. GCaMP6f was amplified by PCR from pGP-CMV-GCaMP6f with 5′ BamHI and 3′ HindIII, and sub-cloned into the recombinant rAAV vector pAAV-6P-SEWB (Shevtsova et al. 2005) for generating pAAV-SYN-GCaMP6f. The human glial fibrillary acidic protein (GFAP) promoter (Hirrlinger et al. 2009) was inserted with MluI and BamHI into pAAV-SYN-GCaMP6f construct for obtaining pAAV-GFAP-GCaMP6f. Serotype 2/1 rAAVs from pAAV-GFAP-GCaMP6f was produced (Tang et al. 2015), and purified by AVB Sepharose affinity chromatography (Smith, Levy, and Kotin 2009), following titration with real-time PCR (rAAV titers
about $1.0-6.0 \times 10^{12}$ viral genomes/mL, TaqMan Assay, Applied Biosystems). For norepinephrine imaging experiments, mice were injected with ssAAV-9/2-\textit{hSyn1-GRAB-NE1m-WPRE-hGHp(A)} (University of Zürich Viral Vector Facility). To visualize Aβ plaques, 7 mg/g methoxy-X04 (Syverstad Skaaraas et al. 2021) dissolved in 0.1 M phosphate buffer saline (PBS), was injected intraperitoneally 24 hours prior to imaging. Also WT littermates were injected with methoxy-X04 to rule out any confounding effects in one of the groups. We found that one injection provided enough signal to outline Aβ plaques for up to three days.

Surgical preparation

Mice were anesthetized with isoflurane (3% for initiation, then 1–1.5% for maintenance) in room air enriched with 20% pure oxygen, and given buprenorphine 0.1 mg/kg s.c. preemptively for analgesia. Bupivacain was administered subcutaneously over the skull, and left for 10 minutes before a boat shaped skin flap was removed. After removing the skin, a 2.5 mm diameter craniotomy was drilled over the somatosensory cortex with center coordinates 3.5 mm lateral and -1.5 mm posterior to Bregma. Virus was injected (70 nL at 35 nL/min at 200 μm depth from the brain’s surface) at three evenly spaced locations positioned to stay clear of large blood vessels, and a glass plug consisting of two coverslips glued together were placed in the craniotomy, slightly pressing the dura to prevent dural overgrowth (Bojarskaite et al. 2020). The surrounding area of the skull was sealed with cyanoacrylate glue and a layer of dental cement. Post-operatively the mice were given meloxicam 2 mg/kg for two days. Only animals with normal post-operative recovery were included in the study. Mice were left to recover for a minimum of two weeks before habituation to head-fixation and imaging.

Two-photon microscopy

After recovery, the animals were imaged in layers 1–3 of the barrel cortex using a two-photon microscope (Ultima IV from Bruker/Prairie Technologies, Nikon 16 × 0.8 NA water-immersion objective model CFI75 LWD 16XW, Spectra-Physics InSight DeepSee laser, Peltier cooled photomultiplier tubes model 7422PA-40 by Hamamatsu Photonics K.K.). An excitation wavelength of 990 nm was used for GCaMP6f imaging, and separate recordings were made using 790 nm excitation light for imaging methoxy-X04, and 890 nm for Texas-Red labeled dextran for visualization of the vasculature. GRABNE fluorescence was recorded at 920 nm excitation wavelength. Image time series were recorded at 30Hz. 3D volume recordings of the morphology and plaque locations were performed in a subset of experiments. All images were recorded at 512x512 pixels with a resolution of approximately either 0.42 or 0.67 μm per pixel (FOV of 215x215 μm, or 343x343 μm, respectively). For imaging, the mice were head-fixed to a custom build stage that allowed free
locomotion on a wheel attached to a rotary encoder. During experiments the mice were illuminated by an IR LED diode, and monitored with an IR sensitive camera. A second camera was used to capture the pupil dynamics (see below). Air puffs to elicit startle responses were delivered by a Picospritzer III (Parker). Instrument synchronization and data acquisition were performed with a custom-made LabVIEW 2015 (National Instruments) virtual instrument.

Behavioral analysis

Mouse locomotion was recorded with a rotary encoder connected to the running wheel. Locomotion signal was captured using a National Instruments data acquisition card using a counter task in the NI Max software, activated through a custom LabView (2015) VI. Data was processed with custom MATLAB scripts to classify running and quiet wakefulness. Criteria for run and quiet wakefulness episodes were validated by manual observation, and defined as follows: Running was defined as continuous segment of at least 4 s forward wheel motion at over 30 degrees/s, no movement faster than 20 degrees/s the last 10 seconds prior to start of running. Spontaneous running was not defined within 30 s following air puff. Quiet wakefulness was defined as continuous segment of at least 10 s duration with less than 2 degrees/s locomotion, as well as no locomotor activity faster than 2 degrees/s for 15 s before segment start. Quiet wakefulness episodes were not defined within 30 seconds following air puff. Quiet wakefulness periods were reviewed manually through the IR-surveillance video recordings to ensure animals were awake when sitting still.

Pupillometry

Pupil size was recorded with a Basler Dart USB camera (daA1600-60um) with a 25mm fixed focal length lens and 2X fixed focal length lens extender (Edmund optics, items #59-871 and #54-356). As two-photon imaging must be conducted in the dark where the pupil would be fully dilated, we illuminated the eye using 470 nm blue light fiber, with a shielding to avoid light contamination, to slightly constrict the pupil. Pupil size was delineated for time periods of isolated runs and startle using a custom build tool developed in MATLAB 2020a. The ratio of pupil to eye-size ratio was calculated and used in the analysis.

Image processing and analysis

Recordings were exclusively processed using MATLAB 2018a to 2020b. Imaging data were corrected for movement artifacts using the NoRMCOrre movement correction software (Pnevmatikakis and Giovannucci 2017). We used our recently published imaging analysis toolbox ‘Begonia’ to remove motion artifacts, mark ROIs and perform event-based Ca²⁺ signal detection (regions-of-activity method; ROA). Astrocytic Ca²⁺ signals were detected using the ROA method, and the ROA density
(i.e. fraction (in %) of the compartment analyzed being active) and ROA frequency in the whole FOV or in anatomical subcompartments were calculated. Methods and algorithms related to this processing pipeline are described in detail in Bjørnstad et al. 2021 (Bjørnstad et al. 2021). In brief, movement corrected image time series data of astrocytic Ca\(^{2+}\) activity were pre-processed by variance stabilization (square root transform) and smoothing in the temporal and spatial domain to achieve a signal-to-noise ratio of at least 9. All image time series were for this reason smoothed with 14 frames in the temporal domain (\(\sim 0.45\) s), and 2 pixels in the \(x\)-\(y\) domain. Four times the median standard deviation of the variance stabilized data plus the baseline value for a pixel was then used to calculate a threshold for signal detection for each pixel. Once the time series had been binarized into signal and non-signal, adjoining \(x\)-\(y\)-\(t\) voxels (in time and space) were connected to ROAs. ROAs with a spatial extent of less than 3 pixels per frame, and duration of less than 3 frames in time were not included in the analyses. For GRAB\(_{NE}\) fluorescence, data were first down sampled from \(\sim 30\)Hz to \(\sim 5\) Hz and corrected for vascular artifacts (dips in fluorescence) due to increases in blood flow by software kindly shared by Prof. Kira Poskanzer at University of California San Francisco with colleagues and Prof. Guoqiang Yu with colleagues at Virginia Tech.

3D A\(\beta\) plaque mapping

Before any recording, z-stacks of images at 5 \(\mu\)m intervals taken at 512 x 512 pixels were recorded while illuminating the tissue with 790 nm laser light. The stacks started approximately at or below dura mater and extended 100–200 \(\mu\)m down. All other recordings were undertaken inside this mapped volume to ensure plaques outside the imaging plane were accounted for. For each imaged time series inside the volume we recorded an additional single image at 790 nm to ensure the precise location of plaques were known. Plaque locations were detected by binarizing the stack at a manually set threshold at which the morphology of plaques were visible. The binarized single images were used to manually align the 2D time series data with the 3D plaque volume. The resulting 3D binary image had small points removed using the Matlab function bwareaopen. Distances from plaque to nearest ROI (RP) were calculated using the 3D distance formula,

\[
= \sqrt{(x - x_0)^2 + (y - y_0)^2 + (z - z_0)^2}.
\]

Tissue processing

All animals were sacrificed after final imaging procedures, at the age of 18 months. The animals were anesthetized with Isofluran Baxter (IsoFlo, Abbot Laboratories) and intraperitoneally injected with
ZRF mixture (Zolazepam 3.8 mg/ml, Tiletamin 3.8 mg/ml Xylazine 0.45 mg/ml and Fentanyl 2.6 µg/ml) before transcardial perfusion with 4°C 2% dextran in 0.1 M phosphate buffer (PB) for approximately 30 s, immediately followed by 4% formaldehyde (FA) in PB for 10 minutes at the speed of 6 ml/min. Following perfusion, the brains were extracted and post fixed by immersion in the fixative at 4°C overnight protected from light. The tissue was stored in 0.1 % FA in PB at 4°C protected from light until further processing. Cryoprotective steps in graded sucrose solution (10%, 20%, and 30% sucrose in PB) were performed before the brains were cut on a freeze microtome (Thermo ScientificTM Microm KS 34) in 40 µm free floating coronary sections and stored in 0.1 % FA in PB at 4°C protected from light until usage. In addition, 8 tg-ArcSwe animals and 8 wild type littermates were sacrificed at 12 months of age and used for qPCR analysis. These animals were anesthetized as described above and decapitated. The brains were extracted, and the left hemisphere was dissected into the frontal cortex, hippocampus, cerebellum and the rest of the brain. This tissue was frozen and stored in -80°C pending analysis.

RNA isolation and real time PCR

48 hours prior to RNA extraction, the samples were suspended in RNAlater™-ICE (Ambion; Cat#: AM7030). To isolate total RNA from the frontal cortex tissue samples, the RNeasy Mini Kit (QIAGEN, Hilden, Germany), including the on column DNase digestion, was used. The RNA concentration and integrity were determined using a NanoDrop 2000c spectrophotometer (ThermoFisher Scientific) and ethidium bromide visualization after agarose gel electrophoresis, respectively. Following the manufacturer's protocol, 1 µg of total RNA was reverse-transcribed into cDNA with Oligo (dT)$_{15}$ using the GoScript Reverse Transcription System (Promega, Madison, USA, Cat#: A5001). All the cDNA samples were diluted in Tris-EDTA buffer (pH 8.0) to a final concentration of 2.5 ng/µl. Real-Time PCR was carried out in a total volume of 20 µl, containing 2x AB Power SYBR® Green PCR Master Mix (ThermoFisher Scientific) with gene specific primers (at a final concentration of 200 nM) and 2 µl cDNA samples. Amplification was performed on the StepOnePlus system (Applied Biosystems) with the following conditions: 95°C for 10 minutes, followed by 40 cycles of 95°C for 15 seconds and 60°C for 1 minute, followed by melting curve analysis to check for unspecific products. Each sample was run in duplicates. Using the NormFinder software (Andersen et. al. 2004), HPRT1 was determined as an internal control for normalization of the gene expression. The primers were designed online using Primer BLAST and setting the amplicon size to a maximum of 200 bp. The primers designed span exon-exon junctions, and standards prepared as previously described (Rao et al. 2021). Details of the Gfap forward and reverse primer are presented in Table 7.
Table 7. Primer used for mRNA quantification.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Protein names</th>
<th>Accession</th>
<th>Forward (5')</th>
<th>Reverse (3')</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gfap</td>
<td>Glial fibrillary acidic protein</td>
<td>NM_010277.3</td>
<td>GC ACTCAATACGAGGCAGTTG</td>
<td>GCTCTAGGGACTCGTTCGT</td>
</tr>
</tbody>
</table>

Immunohistochemistry

One section from each animal was chosen for quantification of astrogliosis and washed in PBS 0.01 M for 10 minutes, followed by two times in 0.1% TritonX100 in PBS (PBST) for 5 minutes. The PBST was removed and blocking (10% normal donkey serum (NDS), 1% bovine serum albumin (BSA), 0.5% Triton X100 in PBS) performed for one hour at room temperature. This was directly followed by incubation overnight at room temperature with primary antibodies (GFAP; host: mouse; diluted 1:1000; Sigma-Aldrich; Cat# MAB360. GFP; host: chicken; diluted 1:2000; Abcam; cat# ab13970) diluted in antibody solution (ABS; 3% NDS, 1% BSA, 0.1% Triton X100 in PBS). The following day the sections were rinsed in 0.1% PBST two times for 1 minute, followed by three times for 10 minutes. Secondary antibodies (CY5 Donkey Anti-Mouse; Jackson ImmunoResearch Labs; Cat# 715-175-151. CY3 Donkey Anti-Chicken; Jackson ImmunoResearch Labs; Cat#703-165-155) were spun in a centrifuge for 10 minutes at 13000 rpm, diluted 1:500 in ABS, and the sections incubated for 1 hour at room temperature. After the second incubation, the sections were washed in PBS for 10 minutes, three times. Propidium iodide (diluted 1:5000 in 0.01 M PBS; Sigma Aldrich; Cat # 04511 (Cellstain double staining kit)) for nuclear staining was added for 10 minutes, before rinsing the sections twice for 5 minutes in PBS. All sections were transferred to distilled water and mounted with ProLong Gold antifade reagent (ThermoFisher Scientific; Cat# P36934). They were stored in -20 °C protected from light until confocal imaging. For electron microscopy: EM standard procedure was followed for embedding and preparing of the tissue (Yang et al. 2011). For enhancing the contrast, uranyl acetate (Fluochem) in double distilled water and lead citrate was used. The sections were examined in a transmission electron microscope (TECNAI 12).

Confocal imaging

GFP-positive astrocytes (indicating GCaMP6f expression) were used to locate the appropriate cortical area used for in vivo imaging. Only sections where we could locate positive GFP staining within the cortex corresponding to the image area were chosen for confocal imaging and further analysis. All
single plain and z-stack images were acquired using a Zeiss LSM 710 confocal microscope. To provide overview and verify that the correct area was identified, one tile scan of 3072 x 3072 pixels was achieved with a 40x objective (1.20; water korr M27) using 3 channels (CY2, CY3 and CY5) for wild types and 4 channels (Dapi, CY2, CY3 and CY5) for tg-ArcSwe. Next, one z-stack of 2048 x 2048 pixels (40x objective; 1.20; water korr M27) from within the GFP positive area was acquired from cortical layer 2. All single plane and z-stack images were obtained with identical settings. In addition, a z-stack of 1024 x 1024 pixels was obtained (40x objective; 1.20; water korr M27) outside the located GFP positive stained area. No post-processing on the analyzed images was performed.

3D reconstruction analysis of astrocytes

This procedure was adapted from the details outlined in Tavares et al. (Tavares et al. 2017) that utilize the free software plugin Simple Neurite Tracer (SNT) of FIJI-ImageJ (Longair, Baker, and Armstrong 2011). Only astrocytes with a single nucleus where at least ⅔ of the circumference was covered by GFAP staining were selected for 3D reconstruction. Astrocytes with processes touching the borders of field of view were omitted. 8 randomly selected astrocytes from each z-stack (2 from each image quadrant) obtained within the GFP positive imaging area were used for analysis. To quantify and visualize the morphological complexity of the astrocytes, we analyzed the number of processes, total length of processes in µm, process thickness (µm³) and number of intersections (provided from the Sholl analysis).

GFAP and GFP area fraction analyses

Z-projections based on the CY5-channel to visualize GFAP/GFP positive astrocytes of all stack images were rendered using the FIJI-ImageJ software (Schindelin et al. 2012). The images were blinded to the analyst, converted to 8 bit images and the scale removed. The threshold was manually adjusted so that only what was considered to be GFAP positive staining was red, and a percentage value of positive staining in the image was obtained.

Statistical analyses

Astrocytic Ca²⁺ signals were studied by means of the ROA density – the fraction of the compartment with activity per time unit. Here, the area may be the entire FOV, or we may limit ourselves to the area identified as belonging to cellular subcompartments; the astrocytic processes, somata or endfeet. In each trial we had time series of ROA density lasting around 300 s (approximately 9000 frames). Within each trial, the startle period was defined as starting from the air puff at 150 s and lasting 600 frames (~20 seconds). In addition, one or more time periods within the trial could be identified as spontaneous runs, or as periods of quiet wakefulness. A first important question concerns the choice
of summary statistics adequately describing the Ca\textsuperscript{2+} response in such periods of interest (runs and startle), which are dynamic behavioral states that entail both acceleration, steady locomotion and deceleration. We have studied the mean and max ROA density and the ROA density rise rate which is defined as the maximal increase in ROA density over a maximum of 50 frames. Initial explorations indicated that the main results are fairly robust to the choice of window length, and 50 frames appeared to be a sensible choice compared to the kinetics of astrocytic Ca\textsuperscript{2+} signals. The ROA density rise rate is meant to capture some of the dynamics in astrocytic Ca\textsuperscript{2+} signaling, and can be understood as the maximum acceleration inside the time period of interest. The rise rate has a high correlation with the maximum, and if the length of the window is increased sufficiently these two statistics tend to become almost identical (since most traces are close to zero at some point in the trial). See Fig. 4 – figure supplement 3, panel 6A, which displays both the max ROA density and the rise rate in an example.

Pupil size measurements had a coarser time resolution than for the astrocytic Ca\textsuperscript{2+} signals; see example in Fig. 4 – figure supplement 3, panel 6B, showing the pupil sizes around a startle response. The pattern in the figure – a sharp increase in pupil size after air puff, before a gradual decline – was quite typical. Therefore, we chose to only consider the measurements in a time-window of 6.67 seconds on each side of the air puff (or start of running). We defined the pupil dilation as the relative increase in the ratio of pupil diameter to eye diameter, inside this window. In other words, we calculate the average ratio before the air puff (or start of running) and after the air puff, compute the difference and divide by the average before the air puff.

For GRAB\textsubscript{NE} analyses, time series were segmented similar to the Ca\textsuperscript{2+} imaging trials. Due to the slower response, change in GRAB\textsubscript{NE} fluorescence was calculated slightly different than the increases in Ca\textsuperscript{2+} signaling, as the difference between the median GRAB\textsubscript{NE} \(\Delta F/F_0\) in the baseline period (10 s immediately prior to locomotion/startle), and during locomotion/startle. Pupil responses and locomotor responses were defined within the same time intervals for these trials.

Interpreting hierarchical plots

We have chosen to present some of our data in the form of hierarchical plots (see Figure 3B and C, Figure 4B and C, Figure 5B and figure supplements). These plots allow the reader to assess the degree of separation between the genotypes and at the same time get an impression of the variation at different levels of the analysis – in our case, the variation between different mice of the same genotype and the variation between repeated measurements on the same mouse. At the lowest level – the trial level – we have points representing the observations themselves. For the spontaneous runs,
there are sometimes more than one run per trial and in that case the points are the median ROA density rise rate in these runs. The maximal number of spontaneous runs in a single trial was 5 (average: 1.6 runs per trial). At the middle level we have the median ROA density rise rate for each mouse, and at the top level the median ROA density rise rate for each genotype. The lines between the levels indicate which observations belong to each mouse, and to each genotype respectively. We have made similar plots for the max ROA density also. The hierarchical plots are a useful tool for exploratory analysis. They are also meant to promote transparency in scientific reporting, and to highlight the importance of intra-group variation. Still, it is important to realize that the impression conveyed by the plots might not be identical to the results from statistical modeling. In the plots, we do not include the influence of various technical and biological covariates which one typically would include in a statistical model. Some of the variation between observations belonging to the same mouse and between different mice of the same genotype may be explained by such covariates, as we will see in the next section.

Modeling

Statistical analyses were conducted in R (version 4.1.1). The ROA density rise rate was modeled by linear mixed effect regression models which were fitted using the glmmTMB package (Brooks, Kristensen, and Van Benthem 2017). We conducted two sets of analyses: (i) to investigate potential differences in ROA density rise rate between the two genotypes; (ii) to investigate the relationship between ROA density rise rate and the pupil dilation, including potential differences between the two genotypes with respect to this relationship. For (i) the coefficient of primary interest is the one belonging to the genotype variable, while for (ii) we are interested in the effect of pupil dilation on the ROA density rise rate, as well as the interaction between this pupil effect and the genotype. In both of these sets of analyses we adjusted for the following fixed effect covariates: the level of magnification (μm per pixel) (2 levels), the depth of the measurements (in μm) and the maximal speed in the relevant time window. We included random intercepts for each mouse (5 WT and 6 tg-ArcSwe). We analyzed the ROA density max and mean values with similar models as the ones described here for the rise rate. For (i), the number of observations ranged between 77 to 117 trials with startle data (depending on the subcompartment) and between 72 and 109 episodes of spontaneous running, while for (ii) the number of observations ranged between 60 and 86 for the startle data and between 35 and 44 for the spontaneous runs. There were less observations for the (ii) analyses because some episodes/trials had missing or incomplete pupil measurements. Similar modeling was performed for GRABNE analyses.

The sensitivity of our result to these modeling choices were assessed by various robustness checks, see next section. The adequacy of model assumptions was investigated by residual plots (Hartig,
n.d.). In the cases where the residual plots indicated deviations from the assumption of constant residual variance, we extended the model by allowing the residual variance to vary as a function of genotype. The reported p-values are based on the t-distribution, with degrees of freedom as provided from the glmmTMB package. No corrections for multiple comparisons were applied.

When analyzing the relationship between distance from the nearest plaque and astrocytic Ca\(^{2+}\) signaling (Figure 2E) we considered the mean ROA density in each ROI (n = 10988) in the quiet wakefulness episodes. If present, any effect of plaque proximity on Ca\(^{2+}\) signaling should be discernible among ROIs observed in the same episode. The dashed lines in Figure 2E show the effect of distance on Ca\(^{2+}\) signaling within each episode, and they form an uncertain picture: in some episodes there is a weak positive relationship, with seemingly higher mean ROA density further away from plaques, while in many episodes there is a negative relationship, with somewhat higher mean ROA density close to plaques. The overall line is found by fitting a linear mixed effect model with mean ROA density as the response, with a fixed effect of distance and with each episode having its own random intercept and slope for the distance effect.

For the results presented in Figure 1: Unless otherwise stated, the data are presented as mean ± standard error of the mean (SEM). A p-value equal to or below 0.05 was considered statistically significant. Mann-Whitney U-test was used to analyze the number of processes, total length of processes in µm, process thickness in µm\(^3\) and area fraction of positive GFAP staining. Two-way ANOVA followed by Sidak post hoc comparison was used to analyze the number of intersections. Statistical analysis was performed in GraphPad Prism version 8.0.1 for Windows (GraphPad Software). For qPCR analysis, mean copy number per ng of total RNA was compared between genotypes by Mann-Whitney U test in SPSS Statistics 26 (SPSS).

Robustness checks

In order to check the robustness of the various statistical analyses, the stability of estimates and p-values was examined with respect to the length of window, for the rise rate response variable, and also sensitivity to individual mice and trials.

The type of sensitivity analysis performed is illustrated (see Fig. 2 – figure supplement 2) for the analysis of the uncoupling between pupil dilation and astrocytic Ca\(^{2+}\) responses (see Fig. 3 and 4 for details about the data). Again, for the purpose of illustration, we focus on the interaction between pupil dilation and genotype for the ROA density rise rate response variable; see Fig. 2 – figure
supplement 2 for details. Similar analyses were performed for the main statistical models, none of these sensitivity analyses provided clear or strong evidence for any change in the main conclusions.

Data and source code availability
A complete dataset of raw and processed data including two-photon microscopy traces, behavioral monitoring and pupil tracking data are provided at: https://doi.org/10.11582/2021.00100. Data analyses were performed with the Begonia toolkit (Bjørnstad et al. 2021), which is available at https://github.com/GliaLab/Begonia.

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References


https://doi.org/10.1093/brain/awab111.


Stobart, Jillian L., Kim David Ferrari, Matthew J. P. Barrett, Chaim Glück, Michael J. Stobart, Marc Zuend, and Bruno Weber. 2018. “Cortical Circuit Activity Evokes Rapid Astrocyte Calcium...


Figure 1. Widespread reactive astrogliosis in tg-ArcSwe mice. (A) Representative micrographs of 15 month old WT and tg-ArcSwe mice somatosensory cortex labeled with anti-GFAP antibodies (green) and anti-NeuN antibodies (blue). Scale bar: 40 μm. (B) Gfap mRNA expression was considerably higher in tg-ArcSwe mice (p < 0.0001, n = 8 animals in both groups, Mann-Whitney U test). (C) The area fraction of GFAP labeling was significantly higher in tg-ArcSwe mice compared to controls (n = 6 tg-ArcSwe and 4 WT littermates). (D) Astrocytes were isolated (inset in A) and analyzed with Simple Neurite Tracer (SNT) plugin in FIJI ImageJ. Tg-ArcSwe mice displayed an increase in the total number of branches (p = 0.02), the length and volume of processes (p = 0.0095 and p = 0.095, respectively, two-way ANOVA) of the GFAP labeled astrocytes (n = 8 astrocytes from each animal. 6 tg-ArcSwe and 4 WT littermates), as well as an increased number of branching points and intersections at 16–32 μm distance from the nucleus (p < 0.0001, two-way ANOVA).

Figure 2. Experimental setup and astrocytic Ca^{2+} signaling in quiet wakefulness. (A) A craniotomy was performed and virus encoding GCaMP6f was injected into the somatosensory cortex of tg-ArcSwe mice and WT littermates. After 3 weeks of recovery, mice were habituated to head-fixation on a disc-shaped treadmill, allowing the mice to move freely at will. Methoxy-X04 was injected 24 hours prior to imaging to visualize Aβ plaques. During imaging, both locomotor activity and pupil responses were recorded. At 150 s in 300 s recordings, the mice were subjected to an air puff to induce a startle response. (B) A representative imaging trial. Simultaneous recording of astrocytic Ca^{2+} signaling, pupil diameter and mouse locomotion during quiet wakefulness, spontaneous locomotion and air puff-induced startle (C) The mice were ~15 months of age during experiments, at a time when they exhibited dense-core Aβ plaques. Left image: confocal micrograph of an Aβ plaque (methoxy-X04, blue) and astrocytes (anti-GFAP, green). Scale bar: 40 μm. Right image: Electron micrograph showing a dense Aβ plaque (purple overlay), autophagic vacuoles (yellow overlay) and relatively normal neuropil morphology (green overlay). Scale bar: 2 μm. (D) Astrocytic Ca^{2+} signals during quiet wakefulness (absence of locomotion) in the form of regions-of-activity (ROAs) displayed in an x-y-t 3D rendering where red regions denote signal. Box-and-whisker plots representing overall Ca^{2+} signals in quiet wakefulness in tg-ArcSwe mice and littermates. (E) 3D visualization of the imaging plane relative to Aβ plaques, with lines representing shortest distance from plaque to ROI. We found a low correlation between distance to nearest plaque and the overall level of astrocytic Ca^{2+} signaling in quiet wakefulness. (F) Example of pathological astrocytic Ca^{2+} wave emanating from an Aβ plaque. Methoxy-X04 visualized by 2P LSM (upper left) and time series from the same FOV of astrocytic Ca^{2+} imaging with a pathological signaling event outlined in red. Scale bar: 50 μm.

Figure 3. Uncoupling of pupil dilation and astrocytic Ca^{2+} responses during spontaneous running. (A) (Left) Representative x-y-t rendering of ROAs during quiet wakefulness and spontaneous locomotion in WT littermates and tg-ArcSwe mice, where red regions denote signal. A spontaneous run occurred at about 60 s.. The slight increase in ROA density at 25 s is due to small locomotor activity that was not large enough to be defined as a run. (Right) Mean time-course of (top-to-bottom) locomotion expressed as wheel speed, relative increase in pupil diameter and astrocytic Ca^{2+} signals expressed as ROA density. The locomotion trace shows mean response across all trials with 95% confidence interval, pupil trace and ROA density traces show median
across all trials ± median absolute deviation. (B) Same as (A), but for tg-ArcSwe mice. A spontaneous run occurred at about 40 s. (C) Hierarchical plots showing median value per genotype in upper level, median value per mouse in middle level, and median level per trial in lower level, in the two genotypes. Upper row: Plots showing the median levels of ROA density rise rate in (left-to-right) the full FOV, astrocytic somata, astrocytic processes and astrocytic endfeet. Lower row: Same as upper row, but showing median levels of max ROA density. (D) Scatterplots of ROA density rise rate (left) and max ROA density (right) vs. relative increase in pupil size upon spontaneous running. See Table 2 for statistical analyses and p values.

Figure 4. Uncoupling of pupil dilation and astrocytic Ca²⁺ responses during startle. (A) In WT mice, in an imaging trial of 300 s duration, mice were subjected to an air puff to the face/vibrissa contralateral to the imaging window at 150 s. This caused an increase in locomotor activity, pupil dilation, and a pronounced increase in astrocytic Ca²⁺ signaling. ROAs are presented as an x-y-t 3D rendering where red regions denote signal. Locomotion trace shows mean response across trials with 95% confidence interval, pupil trace and ROA density trace show median activation across trials ± median absolute deviation. (B) Same as (A), but for tg-ArcSwe mice. (C) Hierarchical plots showing median value per genotype in upper level, median value per mouse in middle level, and median level per trial in lower level, in the two genotypes. Upper row: Plots showing the median level of the ROA density rise rate in (left-to-right) the full FOV, astrocytic somata, astrocytic processes and astrocytic endfeet. Lower row: Same as upper row but for max ROA density. (D) Scatterplots of ROA density rise rate (left) and max ROA density (right) vs. relative increase in pupil size. See tables 3 and 4 for statistical analyses and p values.

Figure 5. Attenuated norepinephrine responses in tg-ArcSwe mice. Mice expressing the fluorescent extracellular norepinephrine sensor GRAB<sub>NE</sub> were subjected to two-photon imaging trials similar to the astrocytic Ca²⁺ imaging experiments. (A) Mean traces ± standard error of locomotor responses, pupil responses and GRAB<sub>NE</sub> fluorescence during spontaneous runs (left column) and during startle (right column). (B) Hierarchical plots showing median ΔF/F₀ of GRAB<sub>NE</sub> fluorescence per (from bottom to top:) trial, mouse and genotype, in spontaneous runs and startle responses.
Figure 2 – figure supplement 1. Locomotor behavior in WT and tg-ArcSwe mice. ▲ indicates air-puff. (A) Behavioral classification for each individual trial, grouped by mouse. See methods for classification and filtering algorithm used. (B) Running wheel speed for each individual trial, grouped by mouse. 20 degrees of rotation of the running wheel per s was the threshold used to classify running. (C) Percentage of total time a mouse spent above a given speed across all trials. (D) Percentage of total time a mouse was running (> 20 degrees/s). Raw running data not filtered by the run classification algorithm (p = 0.35). (E, F, G) Post-classification run episode characteristics.

Figure 2 – figure supplement 2. Robustness and sensitivity analyses for the uncoupling between pupil dilation and the ROA density rise rate. All analyses are with respect to the interaction effect between pupil dilation and genotype. (A) Leave-one-trial-out analysis. The full model is repeatedly estimated removing one trial at the time from the dataset (as a type of jackknife resampling). The estimated interaction coefficients are shown as a histogram. Note that the histogram indicates less variation in the estimated interaction when comparing the variance or empirical quantiles of the histogram with the corresponding confidence interval obtained from the linear mixed effect model (see solid and dashed lines in Plot B). (B) Leave-one-mouse-out analysis. In each iteration, one mouse, M10-M24, is removed from the estimation of the model. The estimated interaction effects with corresponding confidence intervals are then compared to the estimate and confidence interval from the full model (solid and dashed horizontal lines). Some mice influence the results more than others, but there appears to be no systematic bias associated with a single mouse. (C) The p-value for the interaction effect for different window lengths. The window length is measured in number of frames. It is clear that the significance of the interaction effect does not depend strongly on a particular choice of window length. In the analysis we used a window of length 50 frames.

Figure 2 – figure supplement 3. (A) Median GFP-labeling fluorescence indicative of GCaMP6f expression was similar in the two groups. Dots represent median values per mouse. (B) Mean area fraction of the FOV with GFP-labeling was comparable between the two groups.

Figure 2 – figure supplement 4. Regions-of-Activity during quiet wakefulness exhibit the same characteristics in both genotypes. During quiet wakefulness episodes, active regions detected by the ROA algorithm. Compartment association is determined by the centre of the region-of-activity. (A, B, C) Mean duration of active regions during quiet wakefulness in different cellular compartments. Right: GLME statistical analysis estimates. (D, E, F) Mean maximum size of a detected region. Right: GLME statistical analysis estimates.

Figure 3 – figure supplement 1. (A) Hierarchical plot showing medians of mean ROA density per trial, mouse and genotype in the FOV and astrocytic subcellular compartments during spontaneous runs. (B) Same as (A) but during startle responses. (C) Scatterplots of mean ROA density vs. relative increase in pupil size in spontaneous running and startle responses.

Figure 4 – figure supplement 1. (A) Neither tg-ArcSwe nor WT mice displayed any noticeable
habituation to air puffs across all trials and experimental days. (B) Within a single experimental day, there was no habituation to air puffs across the different trials.

**Figure 4 – figure supplement 2.** The dynamic range of pupil dilation and constrictions in the two genotypes. Dots represent median values per mouse.

**Figure 4 – figure supplement 3.** (A) Computation of the ROA density rise rate, and max ROA density, in particular startle period. The vertical dashed lines indicate the time window which had the maximal increase in ROA density, which in this example took place over 49 frames. (B) Computation of the pupil dilation in a particular startle period, here we find (0.268-0.202)/0.202=0.33.

**SOURCE DATA LEGENDS**

**Figure 2 - Source Data 1.** A .csv file containing Ca^{2+} response data (ROA density and frequency), locomotion data and covariates used for statistical modeling for quiet wakefulness.

**Figure 3 - Source Data 1.** A .csv file containing Ca^{2+} response data (ROA density and frequency), locomotion data and covariates used for statistical modeling for spontaneous running.

**Figure 4 - Source Data 1.** A .csv file containing Ca^{2+} response data (ROA density and frequency), locomotion data and covariates used for statistical modeling for startle responses.

**Figure 5 - Source Data 1.** A .mat file containing GRAB\textsubscript{Ne} fluorescence traces during spontaneous running and startles as well as, locomotion data and covariates used for statistical modeling.
Simultaneous 2PLSM, pupil tracking, movement and startle response recording in awake mice.
A

Spontaneous running

Genotype

Field-of-view

Mean ROA density

Mouse

Somata

Processes

Endfeet

Trial

B

Startle

Genotype

Field-of-view

Mean ROA density

Mouse

Somata

Processes

Endfeet

Trial

C

Spontaneous running

WT
tg-ArcSwe

Mean ROA density

0.00 0.25 0.50 0.75 1.00

Relative increase in pupil size

Startle

WT
tg-ArcSwe

Mean ROA density

0.00 0.25 0.50 0.75

Relative increase in pupil size
The box plot compares the range of pupil diameter/eye width between WT and tg-ArcSwe conditions. The p-value for this comparison is 0.93, indicating no significant difference between the two groups.
A

ROA density in FOV

Time (s) since air puff

rise rate = 0.53
(Over 49 frames)

max = 0.69

B

Normalized pupil size

Time (s), air puff at 150 s

0.202

0.268