Methylglyoxal-derived hydroimidazolone, MG-H1, increases food intake by altering tyramine signaling via the GATA transcription factor ELT-3 in *Caenorhabditis elegans*

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The Maillard reaction, a chemical reaction between amino acids and sugars, is exploited to produce flavorful food ubiquitously, from the baking industry to our everyday lives. However, the Maillard reaction also occurs in all cells, from prokaryotes to eukaryotes, forming Advanced Glycation End-products (AGEs). AGEs are a heterogeneous group of compounds resulting from the irreversible reaction between biomolecules and α-dicarbonyls (α-DCs), including methylglyoxal (MGO), an unavoidable byproduct of anaerobic glycolysis and lipid peroxidation. We previously demonstrated that Caenorhabditis elegans mutants lacking the glod-4 glyoxalase enzyme displayed enhanced accumulation of α-DCs, reduced lifespan, increased neuronal damage, and touch hypersensitivity. Here, we demonstrate that glod-4 mutation increased food intake and identify that MGO-derived hydroimidazolone, MG-H1, is a mediator of the observed increase in food intake. RNAseq analysis in glod-4 knockdown worms identified upregulation of several neurotransmitters and feeding genes. Suppressor screening of the overfeeding phenotype identified the tdc-1-tyramine-tyra-2/ser-2 signaling as an essential pathway mediating AGEs (MG-H1) induced feeding in glod-4 mutants. We also identified the elt-3 GATA transcription factor as an essential upstream regulator for increased feeding upon accumulation of AGEs by partially controlling the expression of tdc-1 gene. Further, the lack of either tdc-1 or tyra-2/ser-2 receptors suppresses the reduced lifespan and rescues neuronal damage observed in glod-4 mutants. Thus, in C. elegans, we identified an elt-3 regulated tyramine-dependent pathway mediating the toxic effects of MG-H1 AGE. Understanding this signaling pathway may help understand hedonistic overfeeding behavior observed due to modern AGEs-rich diets.

Keywords: Feeding, Advanced Glycation End-products (AGEs), glod-4, MG-H1, tyramine, GATA transcription factor, elt-3, tyra-2, ser-2, C. elegans, neuronal damage, pharyngeal pumping.
INTRODUCTION

Processed modern diets enriched with Advanced Glycation End-products (AGEs), formed by the Maillard reaction, are tempting to eat but at the same time deleterious for health [1] [2] [3]. In 1912, a French Chemist, L.C. Maillard, reported a reaction between glucose and glycine upon heating, resulting in the formation of brown pigments [4]. Later, the covalent bonds formed between carbohydrates and proteins during heating in a non-enzymatic browning reaction was named the Maillard reaction [5] [6]. Glycation is a part of the Maillard reaction, or browning of food, during cooking which enhances the taste, color, and aroma of the food to make it more palatable [4] [7]. The Maillard reaction has revolutionized the food industry by playing an important role in food chemistry [8]; however, this reaction also results in the formation of adverse AGEs as well as toxic byproducts including acrylamide [9] [10] [11].

Figure 1: Graphical representation for the formation of α-Dicarbonyls and AGEs. Dicarbonyls are highly reactive by-products from metabolic pathways such as lipid peroxidation, glycolysis, etc. In
the above example, methylglyoxal (MGO) spontaneously forms from dihydroxyacetone phosphate which interacts with biomolecules resulting in the formation of Advanced Glycation End-products (AGEs). Toxic MGO is detoxified by glyoxalase enzymes to non-toxic lactate. One of the examples of glyoxalase enzyme in *C. elegans* is *glod-4*. Lack of glyoxalase enzyme leads to increased levels of MGO resulting in increased accumulation of AGEs.

In addition to food sources, AGEs are also endogenously produced in cells when α-dicarbonyl compounds (α-DCs) (such as glyoxal (GO), methylglyoxal (MGO), 3-deoxyglucosone (3DG), etc.) non-enzymatically react with biomolecules. α-DCs are unavoidable byproducts of cellular metabolisms, such as glycolysis and lipid peroxidation (Figure 1). AGEs include GO derivatives such as carboxymethyl lysine (CML) and glyoxal lysine dimer (GOLD). AGEs derived from MGO include hydroimidazolone (MG-H1), carboxyethyl lysine (CEL), and methylglyoxal lysine dimer (MOLD), and 3DG derivatives include 3-deoxyglucosone-derived imidazolium cross-link (DOGDIC), Pyrraline, etc. [1][12][13][14]. The glyoxalase system utilizes enzymes Glo1 and Glo2 and reduced glutathione (GSH) to detoxify α-DCs stress, especially MGO to lactate (Figure 1), in cytosol and nucleus. Differential expression levels of glyoxalases are reported in various disease conditions such as diabetes, hypertension, neurodegenerative disorders, anxiety disorders, infertility, cancer, etc., suggesting their role in exacerbating their pathogenesis [15]. Glo1 has been linked with several behavioral phenotypes, such as anxiety, depression, autism, and pain, among other mental illnesses [16]. Also, we have previously demonstrated increased neuronal damage in the *C. elegans glod-4* glyoxalase mutant model, which is shown to accumulate high levels of α-DCs and AGEs (Figure 1 and Figure 2-figure supplement 1H+1I) [17]. AGEs accumulate in long-lived proteins, such as collagen [18]; further, quantifying the glycated form of hemoglobin (HbA1c) is utilized as a biomarker in diabetes [19]. Increased AGEs are associated with aging, obesity, diabetes, neurodegeneration, inflammation, cardiomyopathy, nephropathy, and other age-related diseases [1][20][21]. Furthermore, neurodegenerative
diseases have also demonstrated a strong correlation between increased levels of AGEs and pathogenesis.

Overconsumption of food and excessive availability of cheap, highly processed foods have contributed to the obesity pandemic. Obesity is a key risk factor for other diseases including diabetes, hypertension, cancers, cardiovascular, inflammatory, and neurodegenerative disorders, among other non-communicable chronic diseases [22] [23] [24] [25] [26] [27] [28] [29]. Thus, identifying signaling pathways that modulate increased feeding behavior is important to understand the underlying causes of obesity and identify novel therapeutics to overcome it. Here, we report that loss of the glyoxalase system or exogenously feeding methylglyoxal (MGO) derived AGEs increased feeding behavior in *C. elegans*. We also identified the mechanism for the observed phenotype and found that the MGO-derived AGE, MG-H1, acts via the *elt-3* GATA transcription factor to partially regulate the expression of *tdc-1* gene, tyramine decarboxylase - an enzyme that biosynthesis neurotransmitter tyramine, and its receptor, *ser-2*, to mediate adverse effects of AGEs such as increased feeding, reduced lifespan, and neuronal damages. This study is the first to identify the signaling pathway mediated by specific AGEs molecules downstream of MGO (such as MG-H1) to enhance feeding and neurodegeneration. Our study emphasizes that AGEs accumulation is deleterious and enhances disease pathology in different conditions, including obesity and neurodegeneration. Hence, limiting AGEs accumulation is relevant to the global increase in obesity and other age-associated diseases.

**RESULTS**

**AGEs increase food intake and food-seeking behavior in *C. elegans***

Our initial observations revealed that *glod-4* glyoxalases enzyme mutants exhibit a significantly enhanced pharyngeal pumping than wildtype N2 animals (Figure 2A). This increase
in pharyngeal pumping was consistent from day 1 (young adult, post-65 hours of timed egg-laying) till day 3 of adulthood (Figure 2A). We performed a food clearance assay to validate whether increased pharyngeal pumping was accompanied by enhanced food intake (Figure 2B and Figure 2-figure supplement 1A). We found increased bacterial clearance after 72 hours in glod-4 mutants.
Figure 2: The glyoxalase mutant, *glod-4*, and methylglyoxal-derived AGE, MG-H1, increases pharyngeal pumping and feeding in *C. elegans*. (A) Quantification of pharyngeal pumping (#/30 sec) in N2 (wt) and *glod-4 (gk189)* mutant at different stages of adulthood. (B) Food clearance assay in N2 (wt) and *glod-4 (gk189)* mutant after 72 hours of feeding. (C) Quantification of pharyngeal pumping (#/30 sec) in N2 (wt) after treatment, with either 150 µM of Arginine (control) or MG-H1. (D) Food clearance assay in N2 (wt) and *glod-4 (gk189)* mutant worms after treatment for 72 hours with either 150 µM of Arginine (control) or MG-H1. (E) Quantification of pharyngeal pumping with different concentrations of MG-H1. (F) Food racing assay in N2 (wt) and *glod-4 (gk189)* at different stages of adulthood towards OP50-1. (G) Food racing assay of N2 (wt) towards OP50-1 when combined with either MGO or MG-H1 (100 µM). (H) Food racing assay of *glod-4 (gk189)* mutants towards OP50-1 when combined with either MGO or MG-H1 (100 µM).

Student t-test for A, B, C, E & F. One-way ANOVA with Fisher’s LSD multiple comparison test for D, G & H. Comparison between two specific groups are indicated by lines above the bars; otherwise, the groups are compared with control group. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bar ± SD.

Serotonin treatment increased bacterial clearance in both wildtype N2 worms and *glod-4* mutants (Figure 2-figure supplement 1B+1C). Further, worms lacking *tph-1* (tryptophan hydroxylase, an enzyme that catalyzes the formation of 5-hydroxy-tryptophan, precursor for serotonin) enzyme as well as *tph-1;glod-4* double mutants which lacks serotonin [30] [31] show decreased pumping compared to wildtype N2 worms (Figure 2-figure supplement 1D). However, *tph-1;glod-4* double mutants show significantly increased pumping compared to *tph-1* single mutants (Figure 2-figure supplement 1D). These data suggest that *glod-4* null mutation mediated increase in pharyngeal pumping is independent of the serotonin signaling [30]. These preliminary observations lead to the hypothesis that enhanced feeding in *glod-4* mutant worms is mediated by endogenous accumulation of AGEs characterized previously [17] [32] [33]. To this end, we
explored AGEs such as MG-H1, CEL, CML, and F-ly as possible mediators of feeding and identified MG-H1 and CEL as potential MGO-derived AGEs to increase feeding in *C. elegans* (Figure 2-figure supplement 1E). Just feeding MGO was not sufficient to increase the pharyngeal pumping rate (Figure 2-figure supplement 1E). Time course analysis in wildtype N2 worms treated with MG-H1 showed that 24 hrs of MG-H1 (150 µM) treatment was enough to increase pharyngeal pumping significantly (Figure 2C). A significant increase in bacterial clearance was observed after 72 hours (Figure 2D). Also, note that treatment of *glod-4* null mutants with MG-H1 did not further increase either the bacterial clearance or the pharyngeal pumping (Figure 2D and Figure 2-figure supplement 1F), suggesting that MG-H1 and *glod-4 null* mutation increases feeding by overlapping mechanism. In addition, we also demonstrated that MG-H1 regulates pharyngeal pumping rate in a dose-dependent manner (Figure 2E). Since MG-H1 is the product of arginine modification by MGO (see ‘Materials and Methods’), we used arginine as a negative control for our MG-H1 treatment. We did not observe a significant difference between worms treated with arginine versus water versus PBS (Figure 2-figure supplement 1G). Since MG-H1 induces increased pharyngeal pumping (Figure 2C+2D), we validated an increase in MG-H1 in the *glod-4* null mutants compared to N2 wildtype using Liquid Chromatography-Multiple Reaction Monitoring (LC-MRM) mass spectrometry (Figure 2-figure supplement 1H+1I). In addition to food consumption, the *glod-4* mutant exhibited a significantly increased preference towards food source OP50-1 at day 1 and day 3 of adulthood compared to wildtype N2 worms (Figure 2F and Figure 2-figure supplement 1J). Furthermore, we noticed that wildtype N2 worms preferred exogenous MG-H1 compared to MGO when provided with bacterial food source *E. coli* OP50-1 (Figure 2G). We did not observe this phenotype in the *glod-4* mutant background (Figure 2H), suggesting MG-H1 in food makes it more appealing for control worms.

**Tyramine regulates MG-H1 mediated feeding behavior via G-protein-coupled receptors (GPCRs) TYRA-2 and SER-2**
Figure 3: Role of tdc-1 and tyramine receptors in mediating the MG-H1 induced feeding behavior.

(A) Differential expression of 66 neurotransmitters and feeding genes in glod-4 RNAi background. 

(B) The flowchart shows the pathway of biogenic amine synthesis, which functions as a neurotransmitter. 

(C) Quantification of pharyngeal pumping in N2 (wt) and tdc-1 (n3419) mutant worms after 24 hrs of treatment of MG-H1. 

(D) Quantification of pharyngeal pumping in N2 (wt), tdc-1 (n3419), glod-4 (gk189), and tdc-1;glod-4 double mutants. 

(E & F) Quantification of pharyngeal pumping in N2 (wt), tyra-2 (tm1846), ser-2 (ok2103), tyra-2;glod-4 and ser-2;glod-4 mutants. One-way ANOVA with Fisher’s LSD multiple comparison test for C-F. Comparison between two specific groups are indicated by lines above the bars; otherwise, the groups are compared with control group. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bar ± SD.

Next, we sought to elucidate how MG-H1 increases the feeding behavior in worms. We performed an unbiased RNA sequencing approach to analyze the global transcriptome profile between control and glod-4 knockdown worms (Supplementary File 1). Our RNAseq analysis identified a total of 20,277 genes, of which 5035 genes were significantly changed (2237 upregulated genes and 2798 downregulated genes) in glod-4 RNAi knockdown worms compared to N2 wildtype. Gene set enrichment analysis showed that the functional category of genes regulating feeding behavior was significantly upregulated in glod-4 knockdown worms (> 2-fold enrichment score) (Figure 3-figure supplement 1, red * marked GO category). This analysis supports our above observation that glod-4 mutants have an altered feeding rate. Previous studies in C. elegans have documented the role of neurotransmitters in C. elegans feeding behavior [34] [35] [36] and we observed differential expression of 66 neurotransmitters and feeding genes (which comprises ~19% of the total feeding and neurotransmission-related genes in C. elegans) (Figure 3A).

We next tested the involvement of these neurotransmitter genes in regulating MG-H1 mediated feeding behavior and systematically analyzed (suppressor screen) MG-H1-induced
feeding in the background of genetic mutants limited in producing different biogenic amines and neurotransmitters in C. elegans (Figure 3B and Figure 3-figure supplement 2A). We found that mutation in tdc-1, the gene involved in synthesizing neurotransmitter tyramine, suppressed the enhanced feeding phenotype in MG-H1 treated animals (Figure 3-figure supplement 2A, indicated by a black arrow and Figure 3C). We also confirmed suppression of increased feeding rate in tdc-1;glod-4 double mutant animals (utilizing two different tdc-1 allelic mutants n3419 and n3420) compared to glod-4 single mutants (Figure 3D and Figure 3-figure supplement 2B ). Next, we checked putative receptors for tyramine that could potentially mediate downstream signaling.

Receptors for tyramine and octopamine are well-studied G-protein-coupled receptors (GPCRs) [37] [38]. We screened seven GPCRs to identify the potential link in regulating tyramine-mediated increased feeding rate exhibited by glod-4 mutant or MG-H1-treated worms. Observed results showed a mutation in ser-2 and tyra-2 suppresses enhanced feeding in MG-H1 treated animals (Figure 3-figure supplement 2C, indicated by black arrows). A similar reversal of feeding phenotype was observed in tyra-2;glod-4, and ser-2;glod-4 double mutant strains (Figure 3E+3F). Our findings support the idea that MG-H1-induced overfeeding is mediated by tyramine signaling.

**GATA transcription factor elt-3 acts upstream of tdc-1 to regulate MG-H1-mediated feeding behavior**

To check for putative transcription factors (TFs) that could regulate the differentially expressed genes in glod-4 knockdown worms (Figure 3), we performed a motif-enrichment analysis (based on available ChIP-Seq data) (Figure 4A+4B). We chose the top five TFs (with a threshold of >18.75% target sequence match for TF- binding motif) for further screening. We knocked down each of the five TFs individually and checked for the suppression of MG-H1-induced feeding behavior (Figure 4-figure supplement 1A). Knocking down pha-4 and elt-3 suppressed the increase in pharyngeal pumping induced by MG-H1 treatment (Figure 4-figure supplement 1A, indicated by black arrows). The pha-4 gene is crucial for pharynx development,
and loss of *pha-4* results in a morphological defect of the pharynx [39] [40], therefore, we followed the results from *elt-3* knockdown.
List of transcription factors identified by motif analysis. (B) Flowchart demonstrating the method of identification of transcription factors. (C) Quantification of pharyngeal pumping after treatment with either Arginine or MG-H1 in elt-3 (gk121) mutants. (D) Quantification of pharyngeal pumping in N2 (wt), glod-4 (gk189), elt-3 (gk121), and double mutant worms. (E) Quantification of tyramine pathway genes in elt-3 (gk121) mutant worms. (F) Quantification of elt-3 and tyramine pathway gene expressions in wildtype N2 (wt) worms after MG-H1 treatment. The horizontal dotted line indicates the normalized expression levels of genes in N2 (wt) and untreated control in E and F, respectively. One-way ANOVA with Fisher’s LSD multiple comparison test for C & D. Student’s t-test for E & F. Comparison between two specific groups are indicated by lines above the bars; otherwise, the groups are compared with control group. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bars ±SD.

Analysis of pharyngeal pumping in elt-3;glod-4 double mutant showed that elt-3 is essential to increase pharyngeal pumping observed in glod-4 mutant worms (Figure 4D). Also, note that the knockdown of elt-3 using RNAi feeding in glod-4 single mutants suppressed the pumping (Figure 4-figure supplement 1B). To determine the role of elt-3 in the tyramine signaling pathway, we performed a HOMER (Hypergeometric Optimization of Motif EnRichment) analysis and identified the binding site of elt-3 on the tdc-1 promoter, which suggested elt-3 may potentially regulate tdc-1 expression levels (Figure 4-figure supplement 1C). This was further validated by reduced expression of tdc-1 mRNA levels in the elt-3 mutant worms (Figure 4E). Next, to check if the elt-3 expression is changed on exposure to MG-H1, we treated wildtype N2 worms with MG-H1 and quantified mRNA levels of elt-3. We observed a moderate but significant increase in the elt-3 expression (Figure 4F). Although tdc-1 and tyra-2 did not change significantly, expression levels of other receptors, tyra-3 and ser-2, increased significantly after MG-H1 exposure (Figure 4F). Note that ser-2 is necessary to mediate the increased pharyngeal pumping (Figure 3F).
Together, these experiments identified a key role for \textit{elt-3} in tyramine-induced feeding increase in response to MG-H1.

\textbf{Tyramine signaling is necessary to increase feeding in \textit{glod-4} mutants}
Figure 5: Exogenous tyramine rescues the suppressed pumping in double mutants. (A) Quantification of pharyngeal pumping in N2 wildtype worms treated with tyramine at various concentrations. (B) Quantification of pharyngeal pumping in elt-3, glod-4, elt-3;glod-4 untreated and elt-3;glod-4 double mutant treated with tyramine. (C-E) Quantification of pharyngeal pumping after treatment of double mutant worms (tdc-1;glod-4, ser-2;glod-4, tyra-2;glod-4) with exogenous tyramine. One-way ANOVA with Fisher’s LSD multiple comparison test for A & B. Student’s t-test for C-E. Comparison between two specific groups are indicated by lines above the bars; otherwise, the groups are compared with control group. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bars ±SD.

To strengthen the role of tyramine in mediating increased feeding in glod-4 KO worms, we treated mutants lacking tyramine signaling with exogenous tyramine. Tyramine has been demonstrated to decrease pharyngeal pumping in wildtype N2 worms [30]. Consistent with this finding, we found that exogenous treatment of tyramine significantly decreased pharyngeal pumping (Figure 5A). However, exogenous treatment of tyramine to rescue tyramine signaling in both elt-3;glod-4, and tdc-1;glod-4 double mutants shows a significant increase in pharyngeal pumping to the levels similar to that of glod-4 single mutants (Figure 5B+5C). Exogenous tyramine did not increase pumping in the double mutants lacking tyramine receptors such as tyra-2;glod-4, and ser-2;glod-4 (Figure 5D+5E). These results strongly demonstrate that tyramine suppresses pharyngeal pumping in N2 wildtype worms; however, it increased the pumping in the glod-4 mutant background. This data supports the notion that tyramine signaling is necessary to mediate glod-4 mutant-dependent increase in feeding behavior.

Absence of tyramine rescues α-DC and AGEs mediated pathogenic phenotypes

Accumulation of α-DC in glod-4 mutants results in pathogenic phenotypes, including neurodegeneration and shortening of lifespan [17]. Here, chronic accumulation of MGO leads to
the build-up of AGEs (Figure 2-figure supplement 1I), thereby increasing feeding in glod-4 worms (Figure 2).

Figure 6: Suppression of glod-4 phenotypes in tdc-1;glod-4 double mutant. (A) Survival assay with N2 (wt), tdc-1, glod-4, and tdc-1;glod-4 double mutants. (B) Survival assay with N2 (wt), tyra-2, glod-4, and tyra-2;glod-4 double mutants. (C) Survival assay with N2 (wt), ser-2, glod-4, and ser-2;glod-4 double mutants. (D) Image of worm neurons showing neuronal damage at day 8 of adulthood. Red arrows indicates damages. (E) Quantification of neuronal damage with pan-neuronal GFP marker in glod-4 vs. tdc-1;glod-4 double mutants. Scale Bar – 10 µm. Log-rank
(Mantel-Cox) test for survival assays. One-way ANOVA with Fisher’s LSD multiple comparison test for E. Comparison between two specific groups are indicated by lines above the bars; otherwise, the groups are compared with control group. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bar ± SD.

Further, accumulation of MG-H1 significantly reduced the lifespan of N2 wildtype worms; however, it did not further exacerbate the damage in glod-4 mutant worms (Figure 6-figure supplement 1). To test whether tyramine signaling is essential for mediating the pathogenic phenotypes such as neuronal damage and reduced lifespan in glod-4 mutants, we compared the lifespan between wildtype N2 and glod-4 worms in the genetic mutants that lack tyramine. The lifespan of glod-4 was significantly increased upon inhibition of tyramine signaling in the tdc-1;glod-4 double mutation (Figure 6A). Next, we tested if the absence of tyra-2 and ser-2 could also rescue the shortened lifespan of glod-4 mutants. Lifespan increased significantly in the absence of either tyra-2 or ser-2 in double-mutant animals (Figure 6B+6C). In addition to rescuing lifespan and feeding rate, the lack of tyramine also resulted in the partial but significant rescue of neuronal damage in glod-4 animals (Figure 6D+6E).

DISCUSSION

Our observation that glod-4 mutants run out of bacterial lawn faster than wildtype N2 animals during routine maintenance led to the elucidation of a novel signaling pathway that mediates AGEs-induced feeding behavior in C. elegans. Glyoxalases are enzymes involved in the detoxification of α-dicarbonyls (Figure 1), and we have previously characterized glod-4 mutant, which lacks one of the glyoxalase enzymes, to accumulate increased levels of α-dicarbonyls [17] and thereby AGEs, especially MG-H1 (Figure 2-figure supplement 1H+1I). In this study, using genetic mutants, RNAi knockdown, synthesized AGEs, and functional genomics, we
elucidate that AGEs (especially MG-H1) induce increased feeding through tyramine signaling regulated by GATA transcription factor ELT-3. The *glod-4* KO worms, which enhance AGEs accumulation, showed increased feeding, which led to the hypothesis that increased accumulation of AGEs is a potential stimulator of binge feeding. Thus, we studied changes in pumping rate by exogenous administration of MGO and AGEs. As previously reported by Ravichandran *et al.* 2018, MGO treatment did not change the pumping rate; however, MG-H1 and CEL increased the pumping rate in wildtype N2 worms (Figure 2-figure supplement 1E) [41]. Further, a recent study demonstrated that treatment with sugar-derived AGE-modified Bovine Serum Albumin (BSA) accelerated the pharyngeal pumping rate [42]. Our study demonstrates that either treatment with purified MG-H1 or endogenous production and accumulation of MG-H1 via genetic mutation increases feeding and adversely affects lifespan. We also found that MG-H1-induced hyper-feeding is independent of serotonin-mediated hyper-feeding [30] in *C. elegans* (Figure 2-figure supplement 1B-1D).

Our detailed investigation of the time-dependent increase in pumping rate after MG-H1 treatment indicates a more robust and highly significant increase after 24 hours of treatment (Figure 2C). We also observed a MG-H1 dose-dependent increase in feeding rate (Figure 2E), which is based on the stronger significance (lower p-value) caused by reduced dispersion of the data at higher concentrations of MG-H1. Our analysis indicates that higher concentrations of MG-H1 can increase the pharyngeal pumping in almost all the treatment worms, thus predisposing the worms with lower pumping rates in the Gaussian distribution to higher pumping rates. It is well established that AGEs are formed during cooking, browning the food during dry heating, making the food more appetizing [14]. Further, feeding is a multisensorial process regulated by several signaling pathways subjected to evolutionary adaptations [9]. Thus, we wanted to analyze the changes in sensory behavior of *C. elegans* induced by either endogenous accumulation of AGEs or by exogenous administration of MG-H1 with the food. Since the *glod-4* mutant lacks a
glyoxalase system to detoxify methylglyoxal and leads to the accumulation of AGEs (Figure 1),
the MG-H1 mediated signaling pathway can be responsible for the increased chemoattraction of
*glod-4* mutant worms to food source OP50-1 (Figure 2F). It can be explained that including MG-
H1 in bacterial lawn increased chemoattraction of wildtype N2 worms towards food, resulting in
increased attraction to palatable MG-H1-mixed bacterial food OP50-1 (Figure 2G). However,
unlike wildtype N2 worms, exogenous MG-H1 treatment had no further increase in the feeding
rate or chemoattraction of *glod-4* mutant worms (Figure 2D+2H and Figure 2-figure supplement
1F), indicating the maximum sensory modulation attained by the endogenous accumulation of
MG-H1 in the *glod-4* mutant. Although our screening identified CEL, a lysine-derived adduct of
MGO, as another AGEs increasing the food intake, a detailed analysis is necessary to conclude
the effect of CEL on feeding behavior (Figure 2-figure supplement 1E).

We utilized RNAseq data from *glod-4* knockdown worms to identify the novel signaling
pathway that mediates AGEs-induced feeding in *C. elegans*. Since *glod-4* knockdown data are
enriched with several genes regulating the synthesis of neurotransmitters and feeding (Figure 3A
and Figure 3-figure supplement 1), we performed suppression screening in mutant worms for
genes involved in synthesizing biogenic amine neurotransmitters after MG-H1 treatment (Figure
3B and Figure 3-figure supplement 2A+2C). Thus, our screen identified *tdc-1*, involved in tyramine
biosynthesis, and tyramine receptors (*tyra-2* and *ser-2*) to mediate AGEs-induced increased
pharyngeal pumping (Figure 3C-3F and Figure 3-figure supplement 2). Tryptophan and tyrosine
are the substrates for synthesizing biogenic amines implicated in modulating various behaviors in
*C. elegans* [35, 43]. Tyrosine to tyramine conversion in the presence of the enzyme tyrosine
decarboxylase (*TDC-1*) followed by tyramine β-hydroxylase (*TBH-1*), is crucial for the synthesis
of neurotransmitters tyramine and octopamine, respectively [43, 44]. Previous studies have
shown the role of tyramine and its receptor (*ser-2*) in regulating feeding and foraging behavior in
*C. elegans* [30] [44] [45] [46]. Further, the *tyra-2* receptor is expressed in MC and NSM pharyngeal
neurons and is discussed to regulate pharyngeal pumping potentially [30] [47]. Especially, tyramine has been shown to reduce pharyngeal pumping when applied exogenously to the worms (Figure 5A) [30]. Supporting previous findings [30], our observation shows increased pharyngeal pumping in tyra-2 and ser-2 single mutant worms (Figure 3E+3F); at the same time, tdc-1 single mutants did not increase pumping (Figure 3D). Converse to our observation of tyra-2 and ser-2 single mutants, Greer et al. 2008 did not find any difference in the pumping rate of tyra-2 and ser-2 single mutants compared to wildtype N2 worms. However, the same study reported no changes in the pumping rate of the tdc-1 single mutant, similar to our results [44], which is also demonstrated by Li et al. 2012 [46]. Interestingly, double mutants of either tdc-1 or its receptors (tyra-2-partial suppression and ser-2) with glod-4 mutant significantly suppress the increased pharyngeal pumping observed in either glod-4 or tyra-2 or ser-2 single mutants (Figure 3D-3F). It is to be noted that only two interneurons, namely RIM and RIC, uv1 cells near vulva and gonadal sheath cells [43] express the tdc-1 gene, which is involved in the biosynthesis of tyramine; however, receptors of tyramine are expressed in distant tissues explaining an endocrine activity for tyramine neurotransmitter [30] leading to the multi-pathway mode of action to exert differential response which should be elucidated in the future. Since ser-3 mutant worms did not suppress the pumping (Figure 3-figure supplement 2C) and ser-3 has been demonstrated to be a receptor for octopamine [30], we conclude that octopamine is not responsible for mediating MG-H1-induced feeding in C. elegans.

Our suppressor screen for the upstream effector of the tdc-1-tyramine-tyra-2/ser-2 pathway that mediates MG-H1-induced increased feeding identified the elt-3 transcription factor (Figure 4C+4D). Thus, we examined whether elt-3 TF regulates the tdc-1, tyra-2, or ser-2. Our analysis revealed that in elt-3 mutant worms, the tdc-1 gene is significantly reduced (Figure 4E), concluding that elt-3 TF regulates tyramine biosynthesis. In favor of the data, HOMER analysis identified that the tdc-1 gene is potentially regulated by elt-3 TF (Figure 4-figure supplement 1C).
Although *elt-3* TF is predominantly expressed in hypodermal cells, its expression is also reported in the pharyngeal-intestinal valve, intestine, a few neurons (head neurons and mechanosensory PVD neuron), etc. (Wormbase.org). In accordance with *elt-3* expression in PVD neurons and head neurons, *tyra-2* is also expressed in PVD neurons [47] and *tdc-1* in RIM and RIC head interneurons, respectively, suggesting a possible direct/partial regulation of *tdc-1* expression by *elt-3*. Also, *tyra-2* expression has been reported in pharyngeal MC neurons, which directly regulate pharyngeal pumping, [47] suggesting direct endocrine action of tyramine. Similarly, *ser-2* is expressed in pharyngeal muscle segment cells [45, 46, 48]. Increased expression of *ser-2* in *elt-3* mutant worms can be inferred as a compensatory mechanism for reduced tyramine signaling by increasing the expression of the tyramine receptor. Also, *ser-2* expression is significantly increased in MG-H1 treated wildtype N2 worms (Figure 4F). Although the mechanism of MG-H1-induced expression of *ser-2* is unclear, it is evident that the *ser-2* genetic mutant can suppress the increased feeding in the *glod-4* mutant (double mutants) (Figure 3F), demonstrating an important role of the SER-2 receptor in mediating the MG-H1 induced feeding via tyramine. Further, *elt-3* expression levels significantly increased after MG-H1 treatment. Altogether, our data strongly suggest the role of the *elt-3-tdc-1-tyramine-tyra-2/ser-2* pathway in mediating enhanced feeding. Finally, it is essential to note that the *ser-2* gene is upregulated in the *glod-4* knockdown RNAseq dataset, similar to significant upregulation after MG-H1 treatment, validating that MG-H1 is a critical player in mediating adverse phenotypes observed in *glod-4* mutant worms.

Exogenous tyramine suppressed pharyngeal pumping in N2 wildtype worms; however, tyramine rescued the pumping in double mutants (*elt-3;glod-4* and *tdc-1;glod-4*) to that of *glod-4* single mutants (Figure 5). Although our data strongly demonstrates the role of tyramine signaling in increasing feeding rate in *glod-4* mutant background, our study also identified a paradox in tyramine signaling to regulated pharyngeal pumping. From the literature [30] as well as from our data, it is evident that tyramine suppresses pumping in N2 wildtype genetic background (Figure
5A). The mechanism behind this behavioral switch, i.e., from suppressor of pharyngeal pumping in wildtype to a stimulator in glod-4 mutant background, remains elusive. It can be speculated that MG-H1 can modify the tissue-specific expression of tyramine receptors (Figure 4F), resulting in an observed behavioral switch in response to tyramine signaling. This hypothesis can only be addressed by methodologies such as single-cell RNA sequencing and exploration of cellular signaling circuitry in further studies. Thus, our current understanding is that MG-H1 (either exogenous or endogenous accumulation in glod-4 KO) modifies tyramine signaling by modulating genes in the tyramine pathway, leading to an increased feeding rate in C. elegans.

Previously, we have demonstrated reduced lifespan, hyperesthesia, and accelerated neurodegeneration-like phenotypes observed in diabetic conditions caused by excessive accumulation of α-dicarbonyls in glod-4 mutant worms [17]. Lack of dicarbonyl detoxification by glyoxalases enzyme in glod-4 mutant worms (Figure 1) should result in the accumulation of AGEs (Figure 2-figure supplement 1H+1I), which at sufficient concentration act as signaling molecules to modulate the feeding behavior (Figure 2) by causing differential gene expression (Figure 3). The increased amount of dicarbonyl stress, thereby AGEs, is observed in several systemic diseases such as obesity, diabetes, cardiovascular and neurodegenerative diseases, among other age-associated diseases [1]. In diabetic patients, three times higher plasma levels of MGO have been reported and is a leading cause of neuropathic pain [49] [50] [51]. Earlier reports in the literature show that the dicarbonyl levels correlate with diabetic complications. One of the major risk factors for diabetes is obesity [52], which is caused by overfeeding. Thus, exploring the regulatory pathways of feeding is essential to understand and identify ways to modulate feeding behavior.

Here, we show that AGEs can modulate feeding behavior in evolutionary primitive model organisms, and it will be worth exploring this pathway in mammals. The transcription factor elt-3 belongs to the GATA transcription factor family [53]. Shobatake et al. 2018 report that GATA 2
and 3 transcription factors induce the expression of appetite regulator genes such as POMC and CART [54]. With the easy availability and unlimited access to modern-day processed food enriched in sugars and AGEs resulting in overeating, a significant cause of the obesity pandemic, it is necessary to explore signals regulating feeding. Importantly, our study shows exogenous treatment with MG-H1 increases feeding in worms (Figure 2C +2D), indicating that a high AGES diet in our day-to-day life can modulate feeding behavior in humans. It is well known that food cooked by grilling, broiling, roasting, searing, and frying accelerates the formation of AGES in food; thus, methods are explored to cook food with fewer AGES accumulation [55]. Further, increased caloric intake and changes in eating habits have been reported in a behavioral variant of Frontotemporal Dementia (FTD) [56] and medication of antipsychotic drugs [57].

Finally, we show that a lack of tdc-1-tyramine signaling rescues glod-4 mutant phenotypes (lifespan and neuronal damage) (Figure 6). A strong association between worsening PD phenotypes with increased aggregation of α-synuclein and specific sites of increased glycation has been demonstrated in different genetic models with increased AGES [58]. Thus, it will be interesting to investigate the role of the tdc-1-tyramine pathway in modulating pathways enhancing neurodegeneration and feeding. Recent research identified neurodegenerative diseases to be influenced by metabolism [59, 60] and glod-4 mutants demonstrate increased neuronal damage, decreased lifespan, and increased feeding. Thus, it is essential to investigate the balance in energy metabolism to identify critical pathways to modulate the outcome of neurodegenerative diseases.

MATERIALS AND METHODS

Strains
Strains were either obtained from *Caenorhabditis* Genetic Center (CGC), Minneapolis, USA or National Bioresource Project, Tokyo, Japan and the following strains were used: N2 (wt), VC343 *glod-4(gk189)*, VC143 *elt-3(gk121)*, MT13113 *tdc-1(n3419)*, MT10661 *tdc-1(n3420)*, FX1846 *tyra-2(tm1846)*, RB1690 *ser-2( ok2103)*, MT9455 *tbh-1(n3247)*, CB1112 *cat-2(e1112)*, MT15434 *tph-1(mg280)*, DA1814 *ser-1( ok345)*, RB1631 *ser-3( ok2007)*, RB745 *ser-4( ok512)*, VC125 *tyra-3( ok325)*, and OH438 *otls117[unc-33p::gfp + unc-4( +)]*. All the mutant strains were outcrossed at least 3 times or more with N2 wildtype. Mutant strains are crossed to get the double mutants *elt-3;glod-4, tdc-1(n3419);glod-4, tdc-1(n3420);glod-4, tph-1;glod-4, tyra-2;glod-4, ser-2;glod-4, glod-4;unc-33p::gfp and tdc-1;glod-4;unc-33p::gfp*. RNAi clones were obtained from Ahringer’s RNAi feeding library and the following were used: *pha-4, ces-1, elt-3, lin-39, egl-5*, and *tdc-1*.

**Growth and maintenance**

Worms were cultured at 20°C for at least two generations on standard NGM agar plates seeded with 5X *Escherichia coli* OP50-1 bacterial strain (Broth culture of OP50-1 was cultured overnight at 37 °C at 220 rpm), which was propagated at room temperature (RT) for two days. For feeding RNAi bacteria, synchronized L1 larvae were transferred to NGM plates containing 3 mM of isopropyl β-D-1-thiogalactopyranoside/IPTG (referred to as RNAi plates) seeded with 20X concentrated HT115 bacteria (cultured overnight at 37 °C at 220 rpm), carrying the desired plasmid for RNAi of a specific gene or bacteria carrying empty vector pL4440 as control and allowed to grow on plates for 48 hrs at 37 °C. For drug assays, synchronized young adult worms (60 to 65 hrs from egg-laying) were transferred to NGM plates (with or without IPTG) with 20X HT115 RNAi bacteria or 5X OP50-1 bacteria (respectively), which are freshly overlayed by the desired drug (or vehicle control) that was air-dried and diffused. Final drug concentrations were calculated considering the total media volume on the NGM plates.
Note: For *glod-4* mutant animals, we found that the pathogenic phenotypes discussed in this paper are contingent on strictly maintaining an *ad-libitum* feeding regimen. Hence, care was taken not to allow the animals to starve by maintaining a low worm-to-bacteria ratio and transferring to fresh plates frequently (at least once every two days).

**Pharyngeal pumping assay**

*C. elegans* pharyngeal pumping was measured using a Leica M165 FC stereomicroscope utilizing a modified previously established method [61] on day 2 young adult worms (unless otherwise specified). Grinder movement in the terminal bulb was used as a read-out for the pumping rate phenotype. Pharyngeal pumping was recorded using a Leica M165 FC microscope; thus, obtained movies were played at X0.25 times the original speed and a manual counter was used to count the number of pumps for 30 sec. For quick pumping screening (pumping data in the figure supplements), the pumping rate was counted in real-time for 30 seconds using a stopwatch and a manual counter focusing the grinder using an Olympus SZ61 stereomicroscope. 10 – 30 animals were counted per biological repeat and 2 to 3 repeats were obtained for each experiment. The pumping data from all the repeats were combined for the presentation of data in the figures. At least one biological replicate was counted blind. Since pharyngeal pumping is very dynamic and changes with worms' development, we decided to record the minimum required treatment groups as possible to reduced variations caused by delayed time and worms' development. In case, when more treatment/genetic groups need to be compared the video recordings between different groups were staggered (recording of 10 worms per group followed by recording worms from other groups and repeating this cycle until 30 worms per group were recorded) to minimize the variations. To reduce the variations induced by fluctuations in the room temperature, 3 plates of worms were prepared for a single treatment group and only 10 worms were recorded per plate while other treatment plates were incubated at 20°C. Animals that did not pump during the recording time, worms that were stationary for prolonged amount of time and...
worms that are potentially injured with visible damages were eliminated from the analysis as well. A few outliers were identified using the Gaussian distribution curve. Under exogenous drug treatment, animals were incubated in the drug at least 18-24 or until 48 hours before measurement of the pump rate. The drugs were overlaid on the NGM plate containing bacterial lawn and air dried before the addition of worms.

**Food clearance assay**

Food clearance assay was performed following minor modifications to the established protocol by Wu et al. [62]. In brief, 20–25 age synchronized (L3-L4 stage) worms were washed twice in S basal then once with S complete medium and transferred to a 96 well plate containing 160 µl assay medium (S-complete medium, growth-arrested OP50–1 at final OD 0.8 (at 600 nm), antibiotics, FuDR and either 150 µM Arginine or MG-H1 or 5 mM serotonin). Initial bacterial density was measured by obtaining OD at 600 nm. Following the indicated number of hours, bacterial density was measured at OD600 after a brief and gentle mixing using a multichannel pipet. For each experimental data point, at least six wells were measured (at least 120-150 worms in total), with the results shown being representative of at least two to three independent assays. The relative food intake was determined by the change in OD for each well, normalized to the number of worms. Under these conditions, ample OP50-1 was available for feeding throughout the analysis, and worms were maintained in the same wells for the entire duration of the experiment.

**Food race assay**

The food race assay to evaluate *C. elegans* choice or attraction for a specific diet, a chemosensory behavior, was performed utilizing a previously established protocol [63]. For this assay, synchronized adult worms (50 per race) were spotted on a 60 mm NGM agar plate, freshly seeded with *E. coli* OP50-1 (with or without drug) approximately 2 cm from the edge of the Petri dish.
plate. Adult animals were aliquoted on the plate diametrically opposite to the food source to estimate the percentage of worms that reached the food source within 30 minutes. An illustration of the food-race assay has been provided (Figure 2-figure supplement 1J).

**Organic synthesis of Advance Glycation End-products (AGEs)**

**Nδ-(5-hydro-5-methyl-4-imidazolon-2-yl)-ornithine (MG-H1) (3)**

![Chemical structure of MG-H1](image)

MG-H1 (3) was synthesized according to the literature procedure with a slight modification as follows; (L)-Arginine (1) (6.07 g, 34.8 mmol, 1 equiv) was dissolved in 12 M HCl (50 mL). To this was added methylglycol dimethyl acetal (2) (4.53 g, 38.3 mmol, 1.1 equiv). It was then stirred at room temperature for 11 hrs. At this time, the reaction mixture was diluted with water (200 mL) and concentrated *in vacuo*. The resulting dark-red solution was purified by SiO_{2}-gel column chromatography (4:2:1 ethyl acetate:methanol:acetic acid) to give MG-H1 (3) as a yellow solid (5.23 g, 22.9 mmol, 66%). The spectroscopic data obtained are consistent with those previously reported in the literature [64].

**Nε-carboxymethyl-lysine (CML) (7a) and Nε-(1-carboxyethyl)-lysine (CEL) (7b)**

![Chemical structures of CML and CEL](image)
CML and CEL were synthesized according to the reported procedure [64] with a slight modification; To a 25 mL flask was added Naα-(tert-butoxycarbonyl)-L-lysine (4) (1.0 mmol, 1 equiv), palladium on carbon (10 wt.% loading, 100 mg, 0.94 mmol), and distilled H2O (7 mL). To this was added glyoxylic acid (120 mg, 1.3 mmol, 1.3 equiv) for CML synthesis or pyruvic acid (115 mg, 1.3 mmol, 1.3 equiv) for CEL synthesis. 1N NaOH (aq) was added dropwise to make pH of this solution 9. A balloon filled with hydrogen gas was attached, and the resulting solution was stirred at room temperature for 14 hrs. At this time, the reaction mixture was filtered through a celite pad and the filtrate was concentrated in vacuo. Purification by SiO2-gel column chromatography (1:2 ethyl acetate:methanol) yielded (6a) (260 mg, 0.85 mmol) or (6b) (263 mg, 0.83 mmol), respectively. To this was added 1 N HCl (aq) (3 mL), and it was then stirred at room temperature for 3 hrs. The resulting solution was concentrated in vacuo to give CML (7a) (164 mg, 0.80 mmol, 80%) or CEL (7b) (172 mg, 0.79 mmol, 79%). The spectroscopic data obtained are consistent with those previously reported in the literature [64].

**Synthesis of F-ly (12)**

F-ly (12) was synthesized according to the literature procedure with a slight modification as follows; To a 200 mL round-bottomed flask was added Naα-(tert-butoxycarbonyl)-L-lysine (4) (510 mg, 2.8 mmol, 1 equiv), D-(+)-glucose (6.15 g, 30.0 mmol, 10 equiv), and MeOH (90 mL). The condenser was attached, and it was refluxed for 7 hrs. After that, it was cooled to room temperature and concentrated in vacuo. The generated solid residue was purified by reversed-
phase SiO$_2$-gel chromatography (H$_2$O only) to provide the desired compound (11) in 53% yield (599 mg, 1.47 mmol). This compound was reacted with 1 N HCl(aq) (3.5 mL) at room temperature and stirred overnight. After the concentration in vacuo, F-ly (12) was obtained in 97% yield (438 mg, 1.42 mmol). The spectroscopic data obtained are consistent with those previously reported in the literature [65].

**Preparation of samples and methodology for RNAseq**

RNA preparation for RNAseq was performed using the Qiagen RNeasy Mini kit (Cat. No. 73404). Total RNA extraction was performed from day 1 adult animals (n=30) picked and collected in 20 µl M9 buffer per condition. 5 biological replicates were used for wildtype N2 and mutant animals. RNAseq on the extracted total RNA was executed at the University of Minnesota Genomics Core (UMGC) using their sequencing protocol for HiSeq 2500 High Output (HO) mode and 50-bp Paired-end sequencing following Illumina Library Preparation. RNAseq coverage was ~22 million reads per sample to perform downstream bioinformatics analyses.

**Bioinformatic analysis**

RNAseq global transcriptome data were subjected to Gene Ontology (GO) based functional classification using the Database for Annotation, Visualization and Integrated Discovery (DAVID) v.6.8. We employed the heatmap2 (Galaxy Version 3.0.1) function from R ggplot2 package to visualize the bioinformatics data.

HOMER (Hypergeometric Optimization of Motif EnRichment) analysis was used to identify the transcription factors for the 66 differentially expressed gene from Figure 3A. A threshold of 18% was used to select potential transcription factors for further screening. Please refer to the flow chart in Figure 4B.

**Reverse transcription polymerase chain reaction (RT-PCR)**
Total RNA was extracted from nearly 100 µl of tightly packed age-synchronized adult worm pellet collected in 1 ml TRIzol reagent provided by Qiagen RNeasy Mini kit (Cat. No. 73404) following manufacturer’s protocol. Subsequently, 1 µg total RNA was used as a template for cDNA synthesis. cDNA was synthesized using the iScript™ cDNA synthesis kit (Bio-Rad, CA) following the manufacturer’s protocol. q-PCR was carried out using the PCR Biosystems Sygreen Blue Mix Separat-ROX (Cat. No. 17-507DB) in a LightCycler 480 Real-Time PCR system (Roche Diagnostics Corp., IN). Quantification was performed using the comparative ΔΔCt method and normalization for internal reference was done using either act-5 or pmp-2. All assays were performed with 3 technical replicates followed by 5-6 biological replicates. Following are qPCR primers used: 1) act-5 gene primers are “TTCCAATCTATGAGATATGCCCTCCC and AAAGCTTCTTTGATGTCCCGGAC”. pmp-2 primer pair is “ATCTTTCAAGCCAATCTCGAC and GAGATAAGTCAGCCCAACTCC”. glod-4: TGTCTCTGTAATGCTTACG and GATGACATTGCTCTATAATCATTACGACTC. 

Lifespan assay

Lifespan assays were performed in Thermo Scientific Precision incubators at 20°C. Timed egg laying was performed to obtain a synchronized animal population, which were either placed onto NGM plates seeded with 5X concentrated E. coli OP50-1. Post-L4 stage or young adult worms (60 to 65 hrs from egg laying) were added to FuDR (5-fluoro-2 deoxyuridine) NGM plates to inhibit the development and growth of progeny. After three days, animals were transferred to a
new 60 mm NGM seeded with OP50-1 and scored every other day thereafter. 45-80 animals were considered for each lifespan experiment, and 2-3 biological replicates were performed. Animal viability was assessed visually and with gentle prodding on the head. Animals were censored in the event of internal hatching of the larvae, body rupture, or crawling of larvae from the plates [17].

**Assay for assessing neuronal damage**

Neuronal damage was assayed using a pan-neuronal GFP reporter strain under different conditions on day 8 of adulthood. Animals were paralyzed using freshly prepared 5 mM levamisole in M9 buffer and mounted on 2% agar pads under glass coverslips. Neuronal damage was visually inspected under an upright Olympus BX51 compound microscope coupled with a Hamatsu Ocra ER digital camera. Images were acquired under the 40X objective. Neuronal deterioration was examined and characterized by loss of fluorescent intensity of nerve ring, abnormal branching of axon/dendrite, and thinning and fragmentation of axons and neuronal commissures [66]. Quantification and imaging of animals harboring damage were performed using the Image J™ software (http://imagej.nih.gov/ij/). To reduce experimental bias, this assay was performed genotype blind with 2 biological repeats.

**Mass spectrometry quantification of MG-H1**

MG-H1 in worm homogenates were quantified using LC-MRM (liquid chromatography – multiple reaction monitoring) following 2,4,6-trinitrobenzene Sulfonate (TNBS) derivatization as described in Hashimoto et al. [67]. An Agilent 1260 HPLC connected to a Sciex 5500 QQQ mass spectrometer was used. The LC conditions were modified as the follows: an Acquity UPLC BEH C18 (130Å, 1.7 µm, 2.1 mm X 30 mm) column was used. The mobile phase (organic: methanol, aqueous: water containing 0.1% FA) was used with a linear gradient of 0-50% of the organic mobile phase over 1.5 min at 0.40 ml/min at 40 ºC. The gradient was increased to 100% organic
at 1.6 min, held for 0.4 min, and equilibrated for an additional 0.4 min. The injections volume was 3 µl. MRM parameters were used as described in Hashimoto et al. with instrument-specific optimization [67].

**Statistical analysis**

All data analyses for lifespan, pharyngeal pumping assays, and gene expression were performed using GraphPad Prism (GraphPad Software, Inc., La Jolla, CA). Survival curves were plotted using the Kaplan-Meier method, and a comparison between the survival curves to measure significance (P values) was performed using Log-rank (Mantel-Cox) test. Two groups were compared for significance using an unpaired Student’s t-test. Multiple group comparison was performed by one-way ANOVA with either Fisher’s LSD or Dunnett’s multiple comparisons test, or Sidak’s multiple comparisons test was used to compare between specific groups. P values from the significance testing were designated as follows: * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001.

**Data and material availability**

All the data generated in this study is presented in the article. Synthetic AGEs can be obtained upon request.

**SUPPLEMENTARY FILE 1**

Supplementary File 1 Legend: A list of genes identified in RNAseq between N2 wildtype and *glod-4* knockdown along with the data of fold change and significance.

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CONFLICT OF INTEREST

Authors declare no conflict of interest.

REFERENCE

33. Michael Morcos, et al., Glyoxalase-1 prevents mitochondrial protein modification and enhances lifespan in.


SUPPLEMENTAL FIGURES

Figure 2-figure supplement 1:
Figure 2-figure supplement 1 legend: (A) Food clearance assay demonstrating increased food intake with an increasing number of worms. (B & C) Food clearance assay of wildtype N2 and glod-4 (gk189) mutant with 5 mM treatment of serotonin, respectively. (D) Quantification of pharyngeal pumping in genetic mutants such as tph-1 (tryptophan hydroxylase), glod-4 and tph-1;glod-4 double mutant compared with N2 wildtype worms. (E) Quick visual quantification of pharyngeal pumping after treatment with different AGEs molecules at 100 µM for 12-18 hours. (F) Quantification of pharyngeal pumping with treatment of either N2 wildtype or glod-4 mutants with either arginine or MG-H1 at 150 µM. G) Quantification of pharyngeal pumping in N2 (wt) worms after treatment with phosphate-buffered saline or 150 µM of Arginine for 24 hours. (H) LC-MRM chromatograms for the exacted ion peaks for TNP-MG-H1 in N2 wildtype (left) and glod-4 mutant (right). (I) Relative quantification of MG-H1 in worm lysates of N2 wildtype and glod-4 mutant background from (H). (J) Pictorial representation of food racing assay. Student’s t-test for B+C+I. One-way ANOVA for A+D+E-G. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bar ±SD.
Figure 3-figure supplement 1:

Gene ontology analysis for upregulated genes in *gld-4* mutant worms.
Figure 3-figure supplement 2 legend: (A) Quantification of pharyngeal pumping (quick screening by visual counting) on mutants of enzymes involved in the biosynthesis of biogenic amines after MG-H1 treatment (suppressor screen). (B) Quantification of pharyngeal pumping in genetic mutants such as tdc-1 (tyrosine decarboxylase) n3420 allelic mutant, glod-4 mutant and tdc-1;glod-4 double mutant compared with N2 wildtype worms. (C) Quantification of pharyngeal pumping (quick screening by visual counting) on receptor mutants involved in feeding behavior after MG-H1 treatment. Student’s t-test for A+C. One-way ANOVA for B. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bar ±SD.
Figure 4-figure supplement 1 legend: (A) Quantification of pharyngeal pumping (Quick visual counting), suppressor screen for top 5 transcription factors listed in Figure 3A. (B) Quantification of pharyngeal pumping in glut-4 mutant worms along with elt-3 gene knockdown by RNAi feeding comparted with N2 (L4440 represent the control feeding plasmid without dsRNA). (C) List of genes obtained by HOMER analysis that are potentially regulated by the elt-3 transcription factor. Student t-test in A. One-way ANOVA for B. * p<0.05, ** p<0.01, *** p<0.001 and **** p<0.0001. Error bar ±SD.
Figure 6-figure supplement 1 legend: Survival analysis of N2 and *glod-4* mutant worms after MG-H1 treatment at 150 μM. Log-rant (Mantel-Cox) test for statistical analysis. ** p<0.01, *** p<0.001 and **** p<0.0001.