

Guide to image data on FlyLight websites

For: splitgal4.janelia.org & gen1mcfo.janelia.org

The screenshot shows a detailed view of a FlyLight sample page. At the top, the FlyLight logo and "FlyLight Split-GAL4 Driver Collection" are displayed, along with links to various labs: Aso Lab, Branson Lab, Card Lab, Descending Interneuron, Dickson Lab, and Rubin Lab. The HHMI Janelia Research Campus logo is also present. The main content area features a teal header with the identifier "SS00043" and the text "Upcoming top cell-type-specific line". Below this, a table provides experimental details: Primary publication (DOI) 10.1002/cne.24512, Robot ID 2502208, AD R65B12-p65ADZp in attP40; MKRS/TM6B, DBD R30E10-ZpGdbd in attP2, Fly Core Project Split_GAL4, Genotype w; R65B12-p65ADZp in attP40; R30E10-ZpGdbd in attP2, and Grade A. To the right, instructions for viewing projections and movies are provided, mentioning Fiji as a suitable viewer. At the bottom, three buttons offer links to NeuronBridge, Virtual Fly Brain, and the Bloomington Stock Center.

Primary publication (DOI) 10.1002/cne.24512

Robot ID 2502208

AD R65B12-p65ADZp in attP40; MKRS/TM6B

DBD R30E10-ZpGdbd in attP2

Fly Core Project Split_GAL4

Genotype w; R65B12-p65ADZp in attP40; R30E10-ZpGdbd in attP2

Grade A

View on NeuronBridge View on Virtual Fly Brain Order from Bloomington Stock Center

Adult 20x Objective Images [2 images]

Reporter: 20XUAS-CsChrimson-mVenus trafficked in attP18

Sex	Female
Age	Day 3-10
Slide code	20180411_33_C1
Release	Wolff 2018

Select an image to view

Brain

Ventral nerve cord

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Contents:

1. Image formats
2. FlyLight sample labeling methods
3. 63x and 40x image tile names

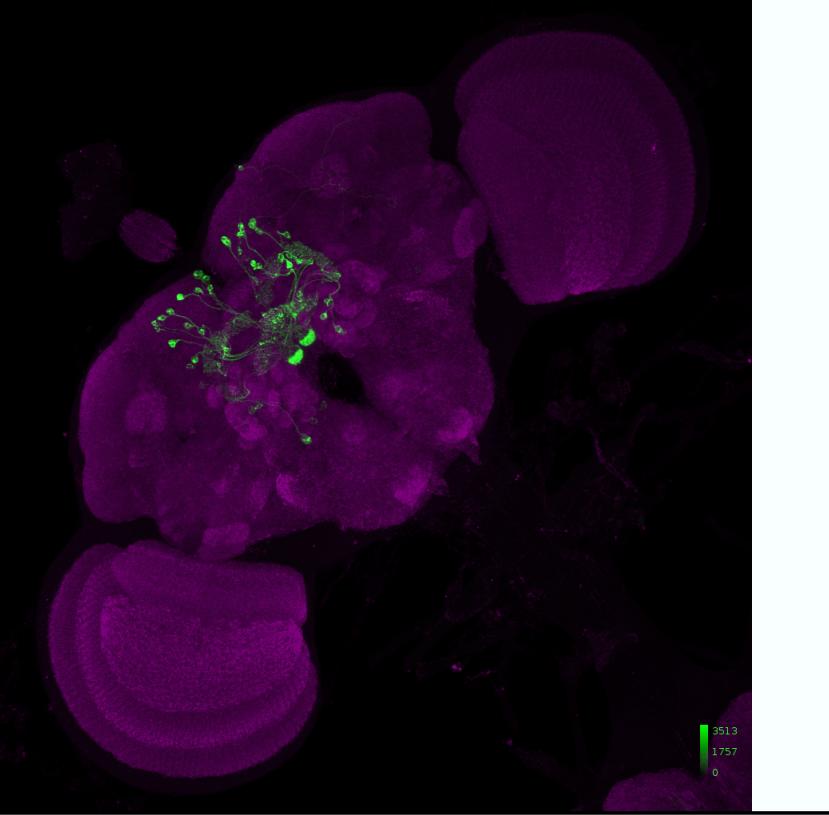
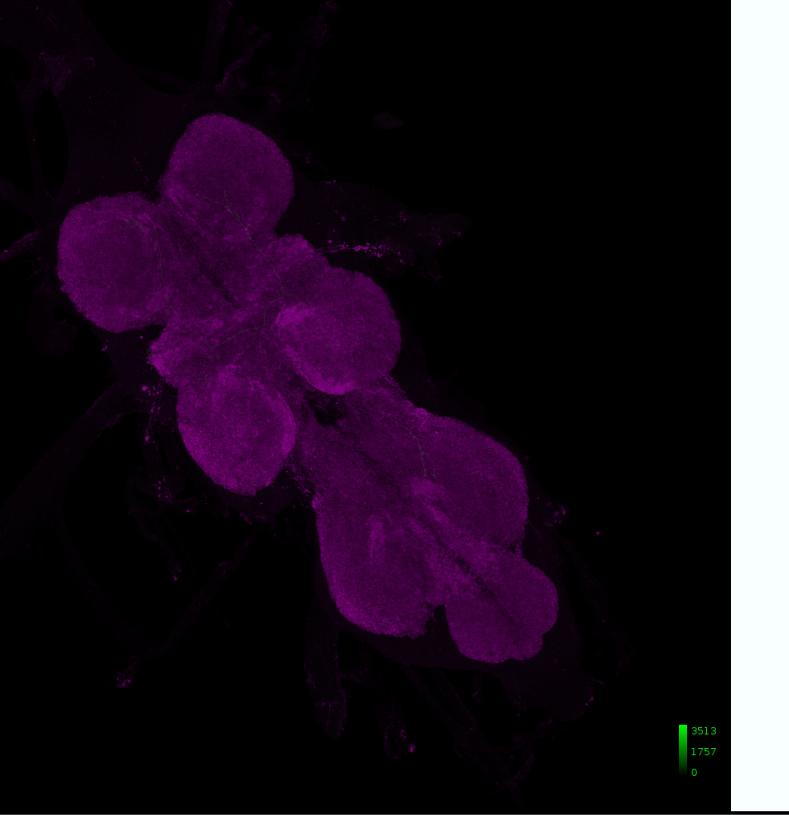
Organization

▽

Adult 20x Objective Images [2 images]

Reporter: 20XUAS-CsChrimson-mVenus trafficked in attP18

Sex	Female
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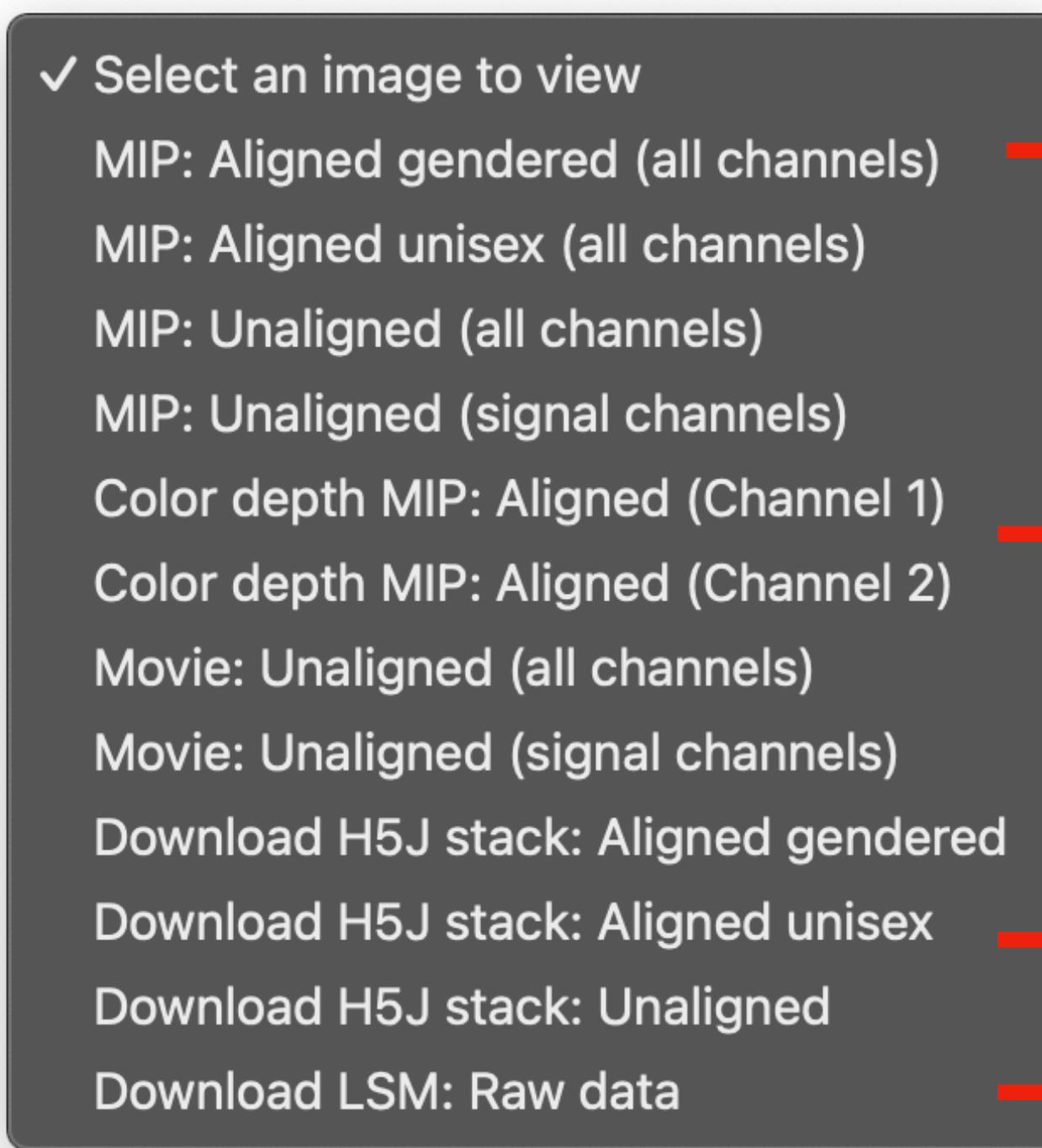
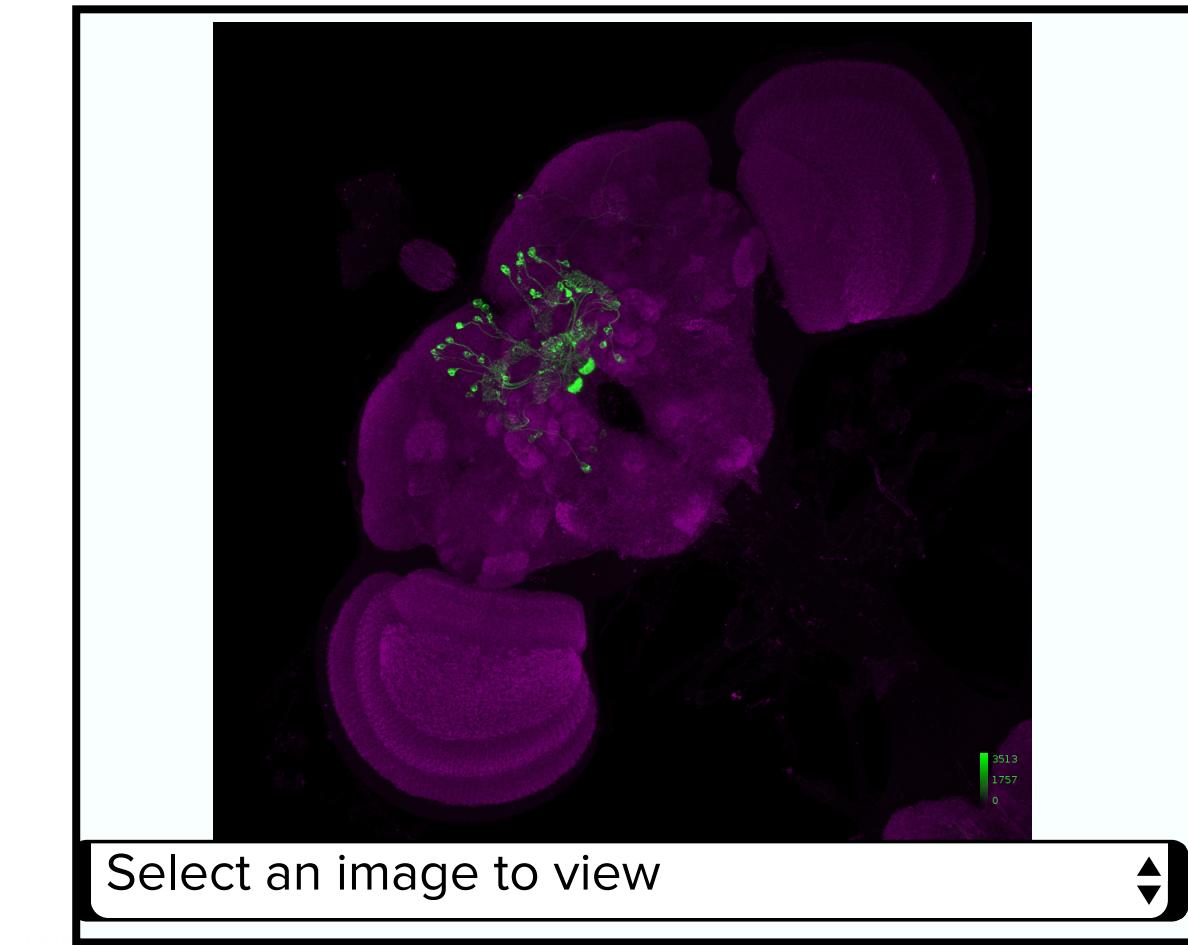
Brain

Select an image to view 

Ventral nerve cord

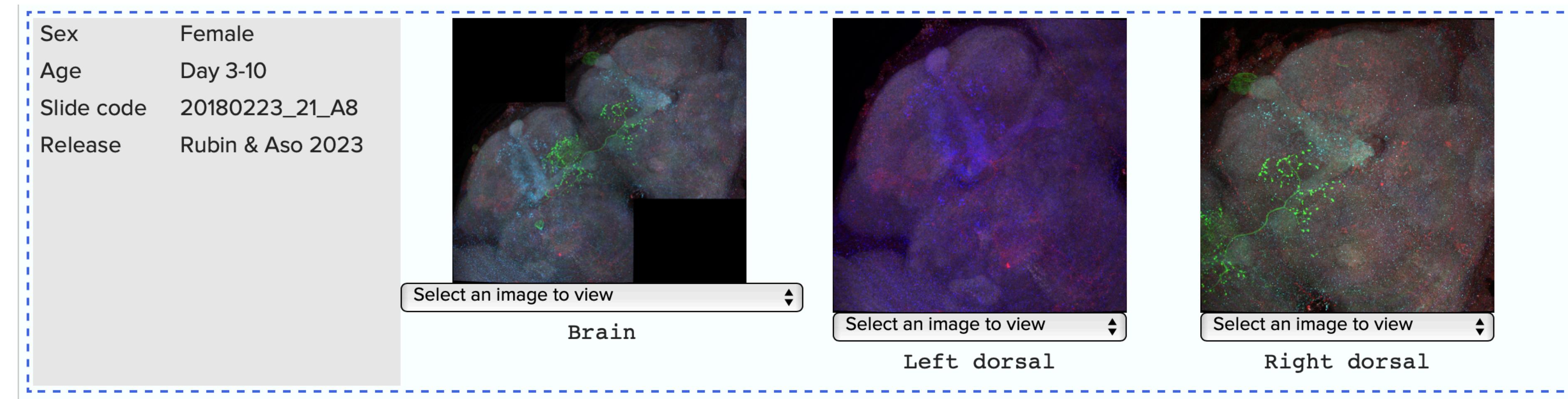
- Images for each line are grouped by objective, reporter, and sample (fly), with image tiles shown as thumbnails
 - Select the menu beneath a thumbnail to view or download a variety of 2D image, movie, or microscope format 3D stacks, detailed on next page

Image formats



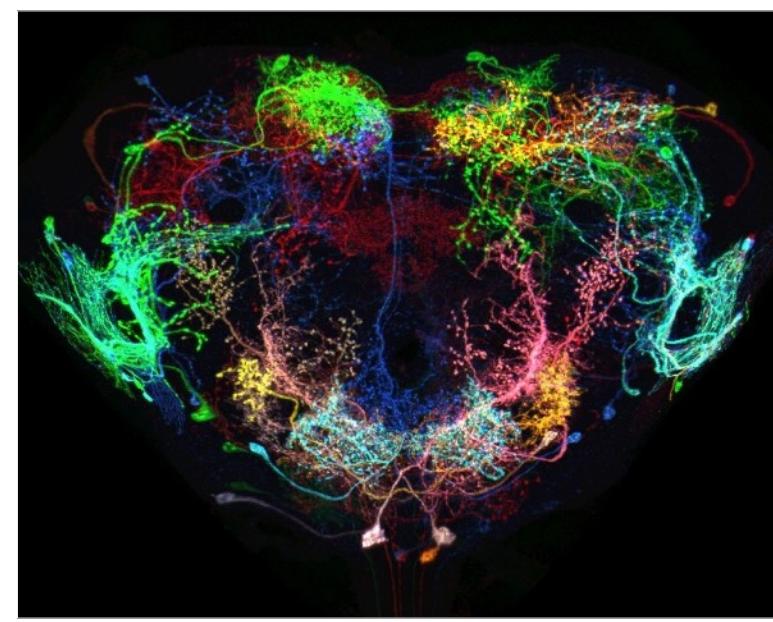
- Thumbnails show unaligned neuropil reference and neuron signal channels
- Maximum Intensity Projections (MIPs) give 2D views of data with original colors
- Images can be unaligned (as captured) or aligned to JRC2018 template (<https://doi.org/10.1371/journal.pone.0236495>)
- Color Depth MIPs (CDM) use color to indicate depth in z dimension; blue is close and red is far (<http://dx.doi.org/10.1101/318006>)
- Most stacks are H5J compressed (<http://data.janelia.org/h5j>)
- Original data capture format is a minimally-compressed LSM stack

Image processing details

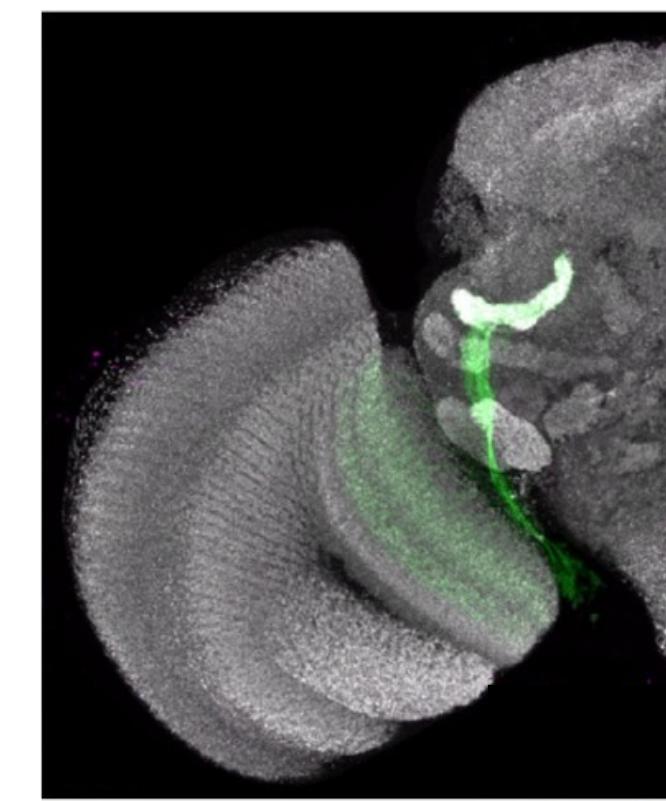


- The number of samples per line varies
- Slide code indicates date of slide creation, pipeline code, and location of individual sample on slide
- Images are individually optimized for brightness
- Some types of image outputs may show saturation; raw data (LSM stacks) have minimal saturation
- Brain and VNC images may be stitched from multiple tiles. Tiles for each sample/region are captured with consistent imaging parameters but are shown scaled independently.

FlyLight pipelines characterizing genetic lines

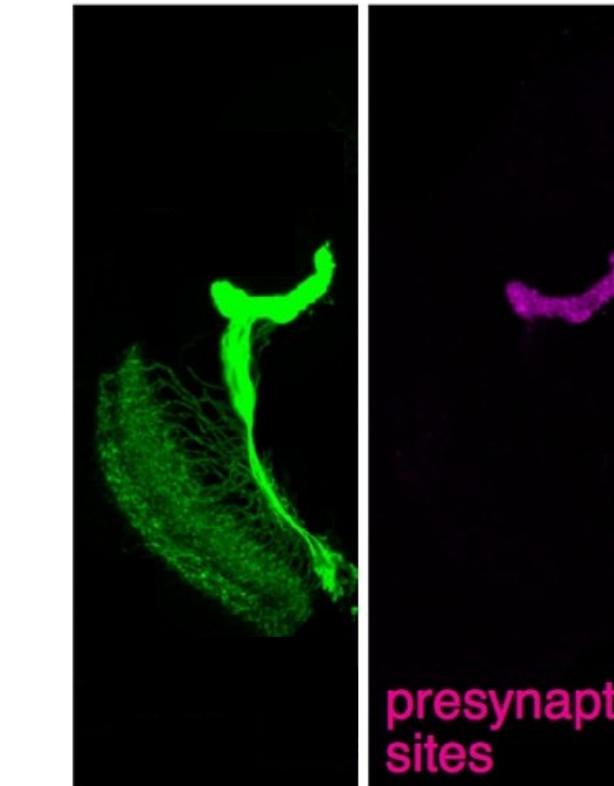


Generation 1
MCFO:
Identifying potential
Gen1 GAL4 lines

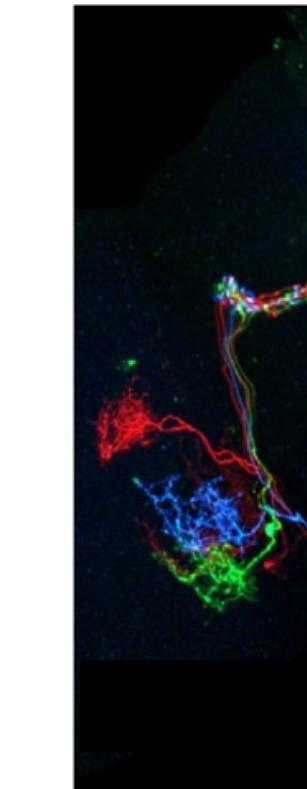


Wu, Nern, et al., 2016

Screen:
Identifying
the best split-
GAL4
combinations

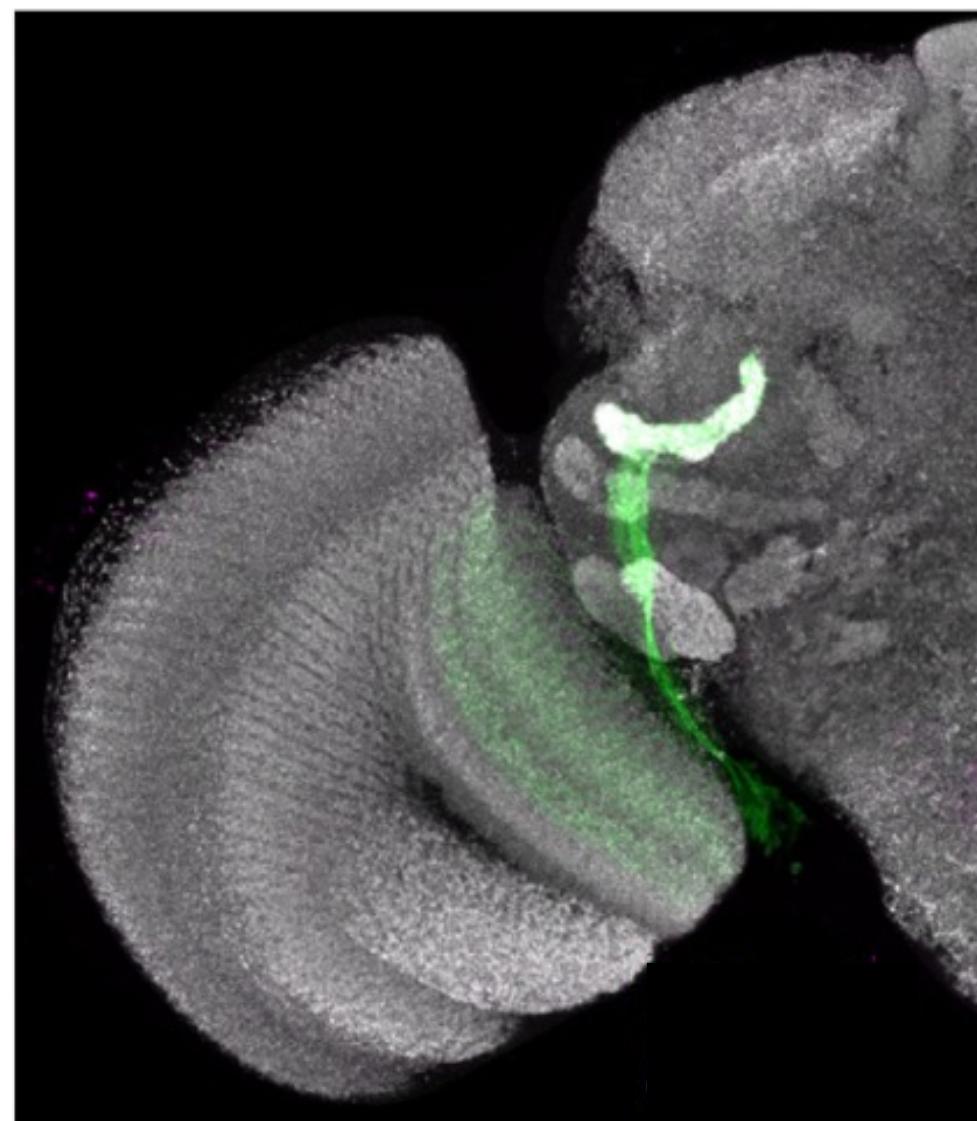


Polarity:
Determining
neuronal
input and output



**Multicolor Flp-
out (MCFO):**
Understanding
cells within a
line

Screen pipeline

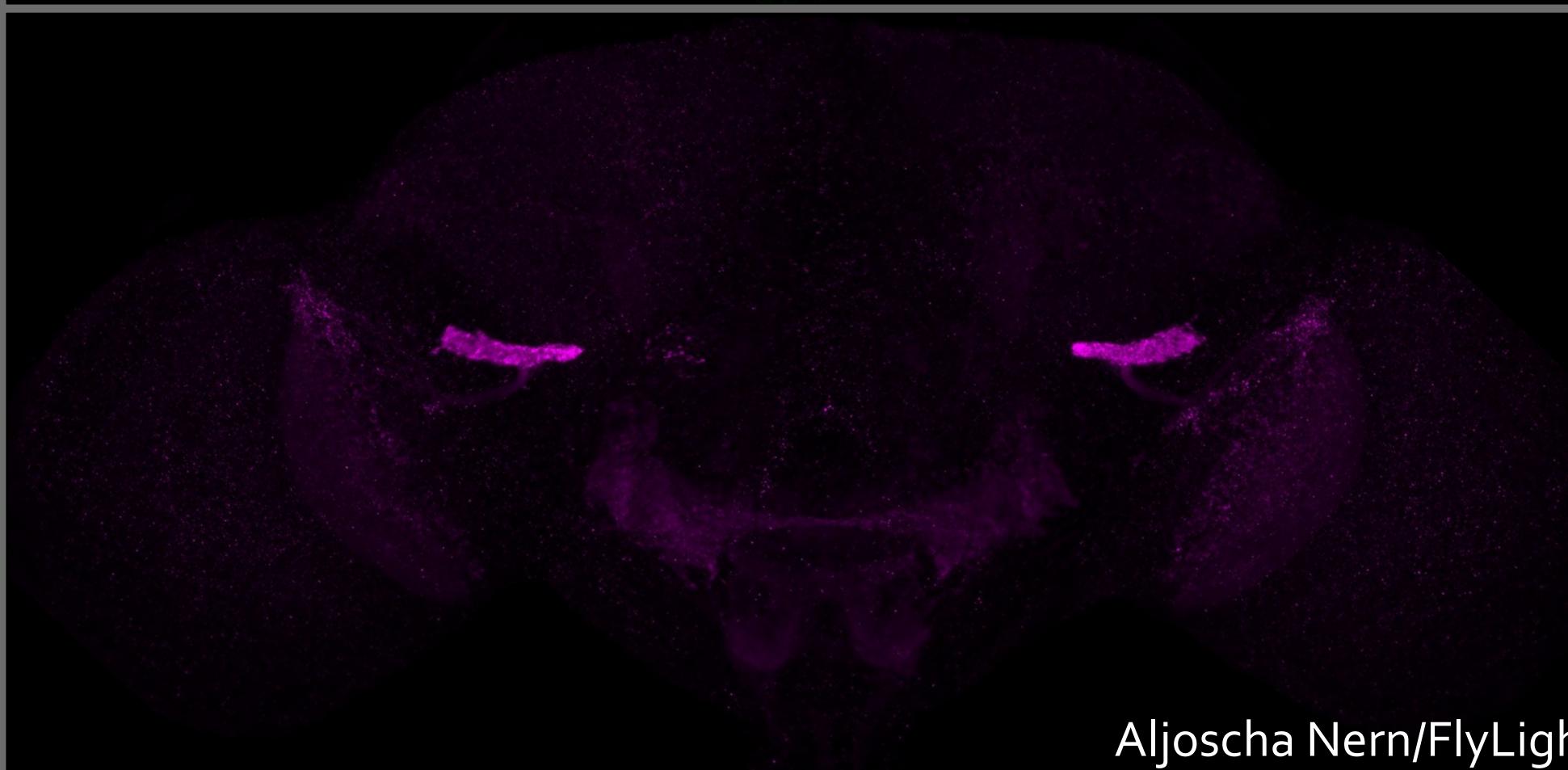
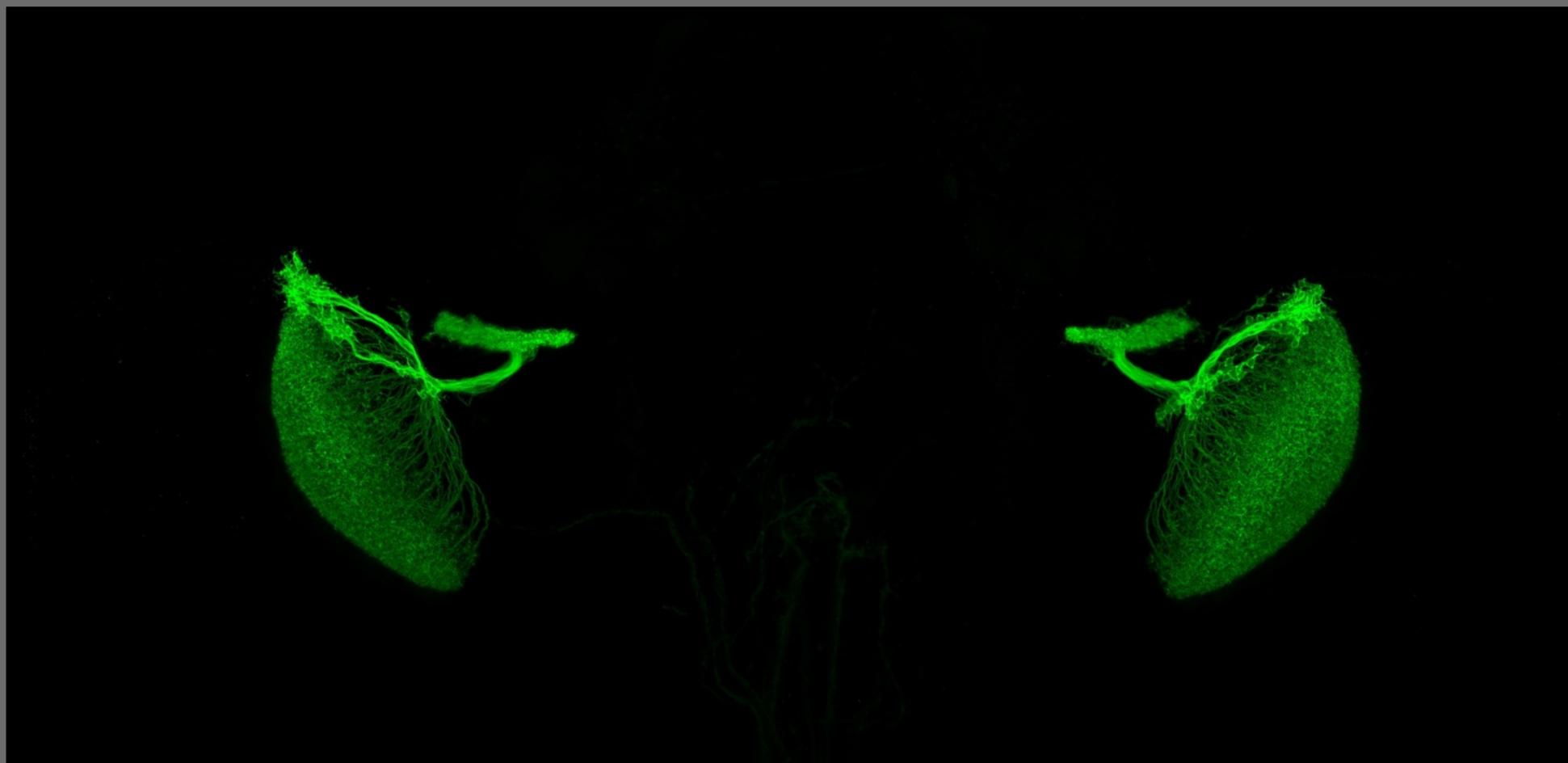
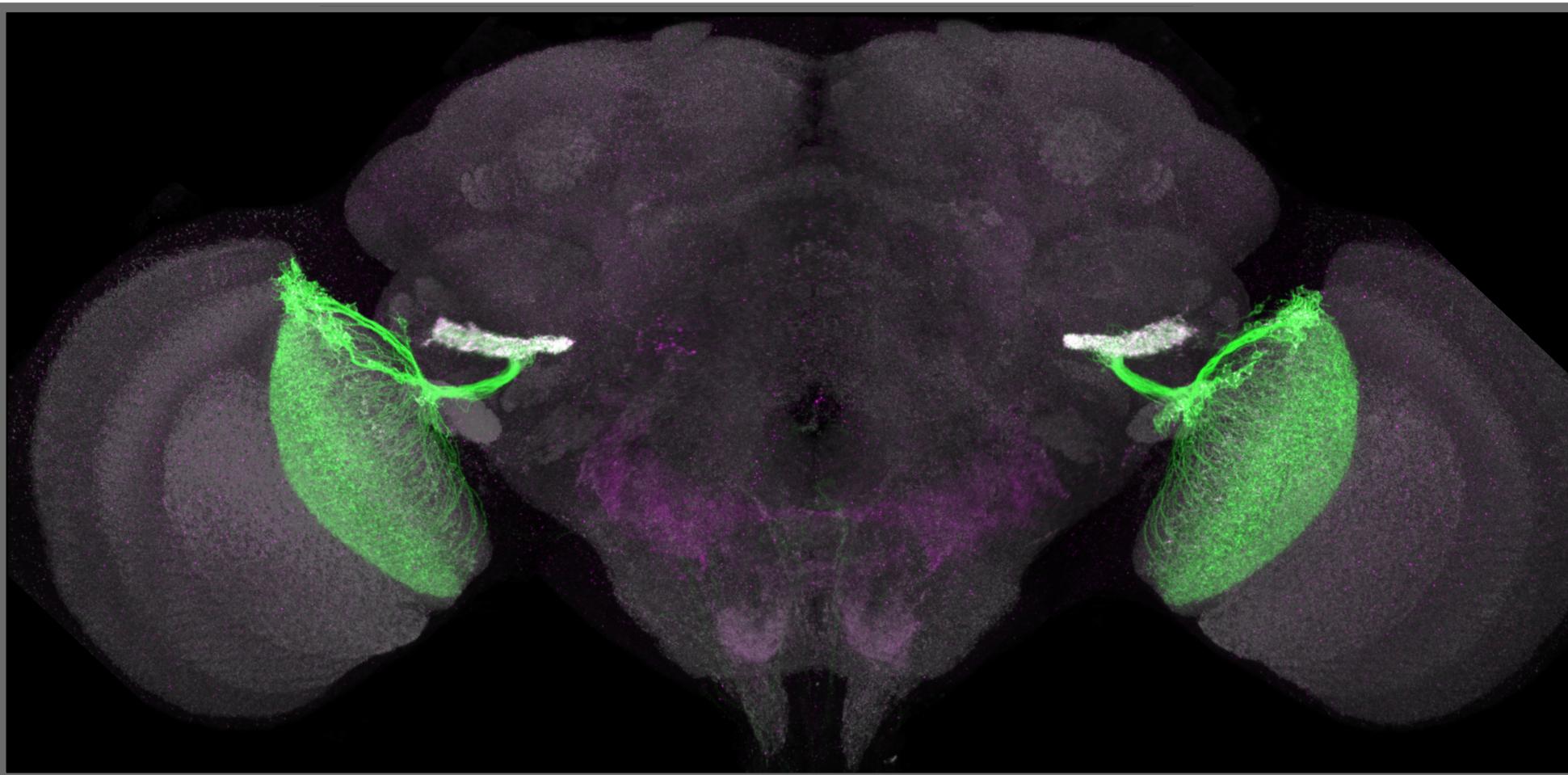


UAS reporter: 20XUAS-Cs-Chrimson-mVenus trafficked in attP18

Screen		
Target	Reference neuropil	neuron
Genetic marker	endogenous	UAS-GFP
Primary	mouse nc82	rabbit anti-GFP
Secondary	568 anti-mouse	488 anti-rabbit



Polarity pipeline



Standard approach is to label:

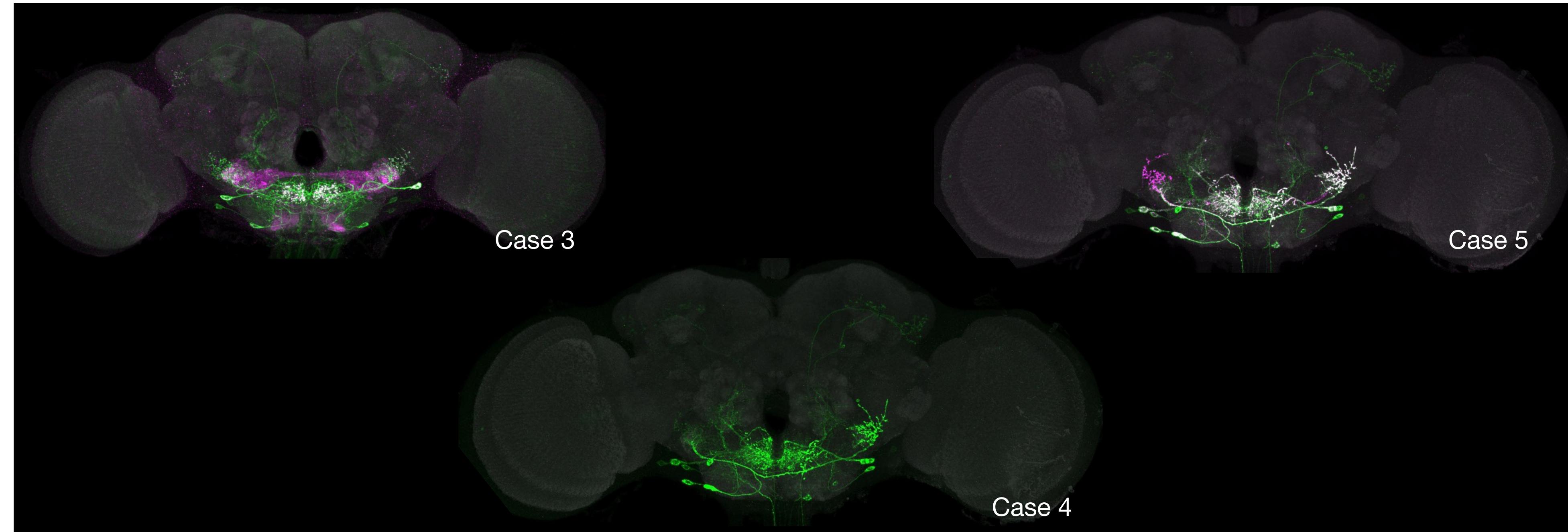
- GAL4-specific neuronal membrane
- GAL4-specific presynaptic terminal
- nc82 neuropil reference

Two main purposes:

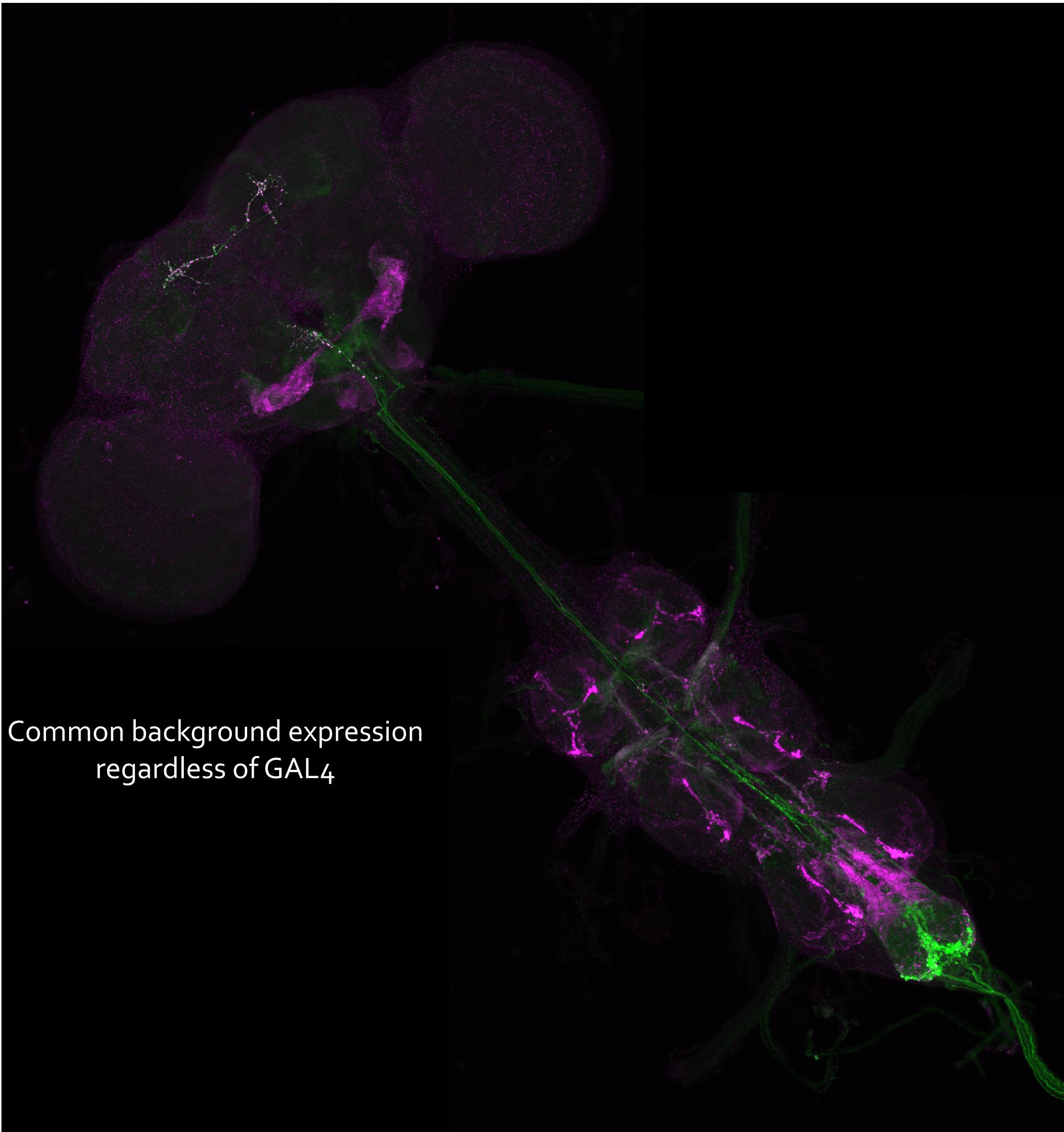
- Determine polarity (inputs and outputs) of neurons
- High-quality GAL4 membrane labeling

Polarity Cases (variations on labeling methods)

	Case 3		Case 4		Case 5	
	Primary Ab	Secondary Ab	Primary Ab	Secondary Ab	Primary Ab	Secondary Ab
nc82 (neuropil)	mouse nc82	Cy2 anti-mouse	mouse nc82	AF568 anti-mouse	mouse nc82	AF568 anti-mouse
UAS-Syt-HA (synapse)	Rabbit anti-HA	Cy3 anti-rabbit	-	-	Rat anti-HA	ATTO647N anti-rat
UAS-myr-FLAG (membrane)	Rat anti-FLAG	ATTO647N anti-rat	-	-	-	-
UAS-Chrimson (membrane)	-	-	Rabbit anti-GFP	AF488 anti-rabbit	Rabbit anti-GFP	AF488 anti-rabbit

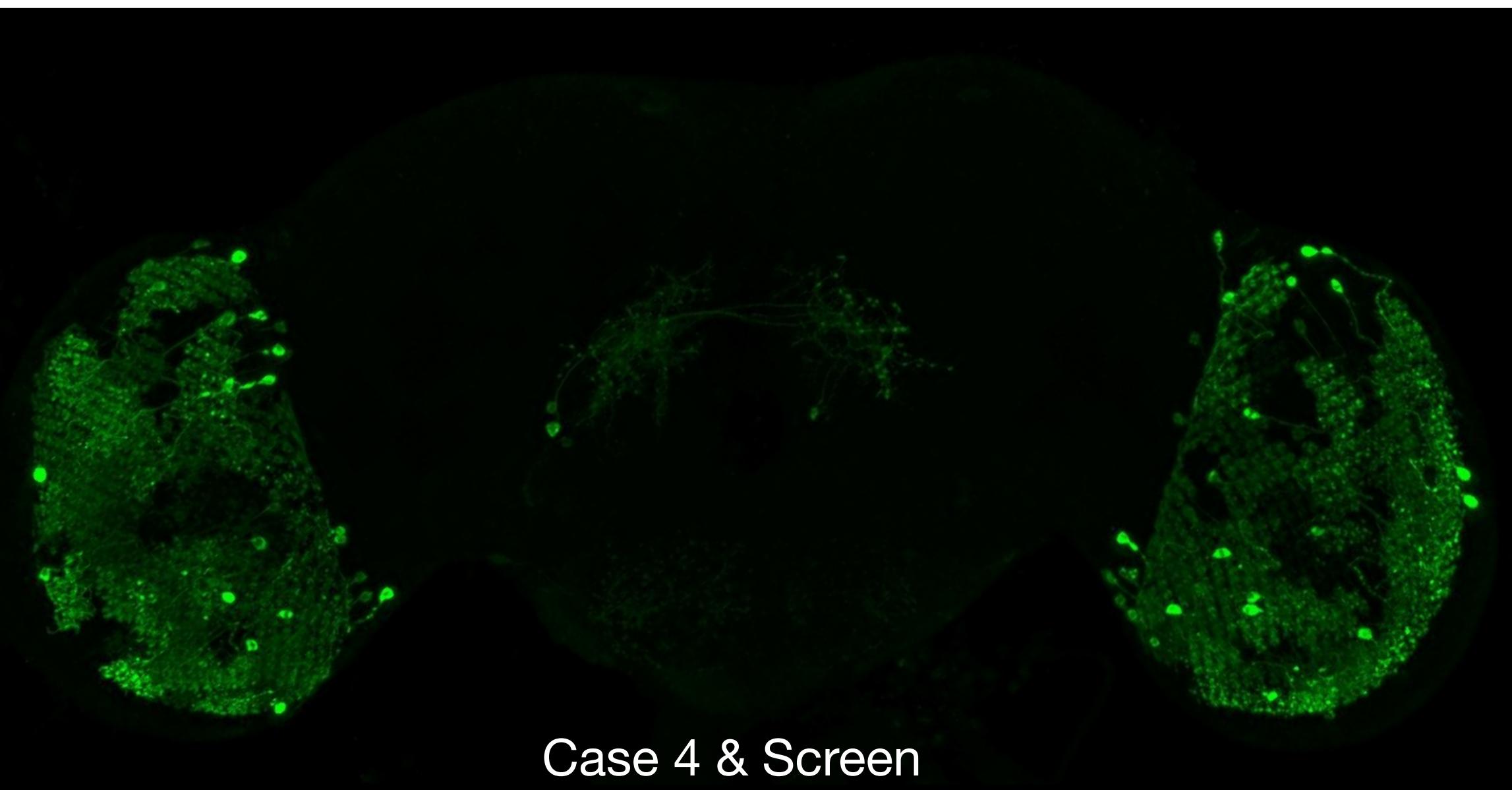


Polarity Case 3

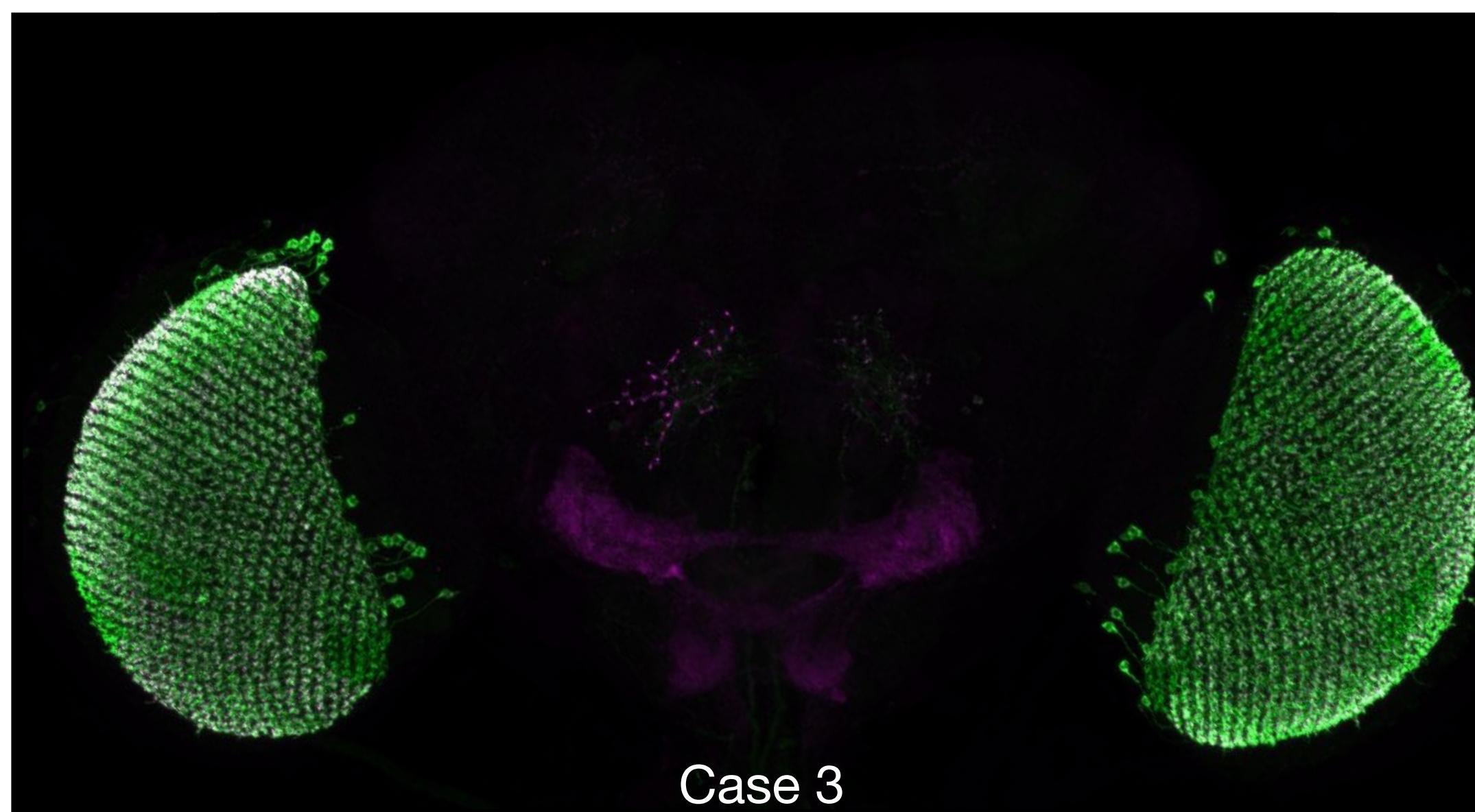


- UAS Reporters:
- Membrane: pJFRC225-5XUAS-IVS-myr::smGFP-FLAG in VK00005
- Synapse: pJFRC51-3XUAS-IVS-Syt::smGFP-HA in su(Hw)attP1
- Excellent intracellular synaptic labeling specificity
- Membrane label reliably fills GAL4 patterns
- But high background expression of membrane and synaptic reporters
- UAS-myr-FLAG reporter instead of Chrimson-mVenus
 - Different reporter from Screen & behavioral studies
 - Different attP insertion site
 - Harder to correlate Polarity pattern with other studies

Polarity Case 4

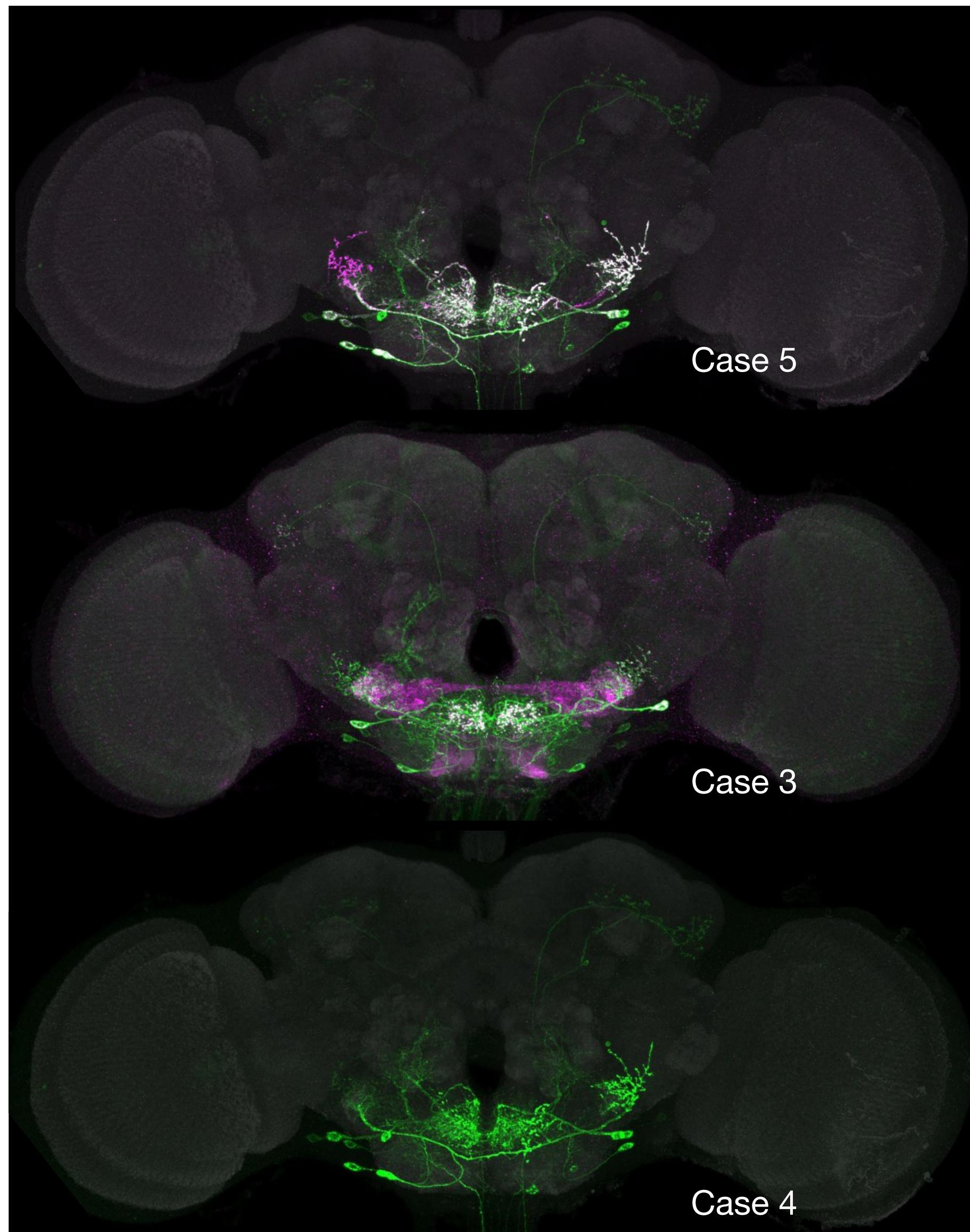


- UAS reporter: 20XUAS-Cs-Chrimson-mVenus trafficked in attP18



- Polarity Case 4 and Screen pipelines use the same UAS reporter and antibodies
- UAS-Chrimson-mVenus reporter allows direct comparison to physiology
- No synaptic label, despite Polarity name
- Very low background
- Less uniform labeling of GAL4 pattern than UAS-myr-FLAG
 - Greater variation between cell body and projection intensities
 - Doesn't always label all neurons in GAL4 pattern

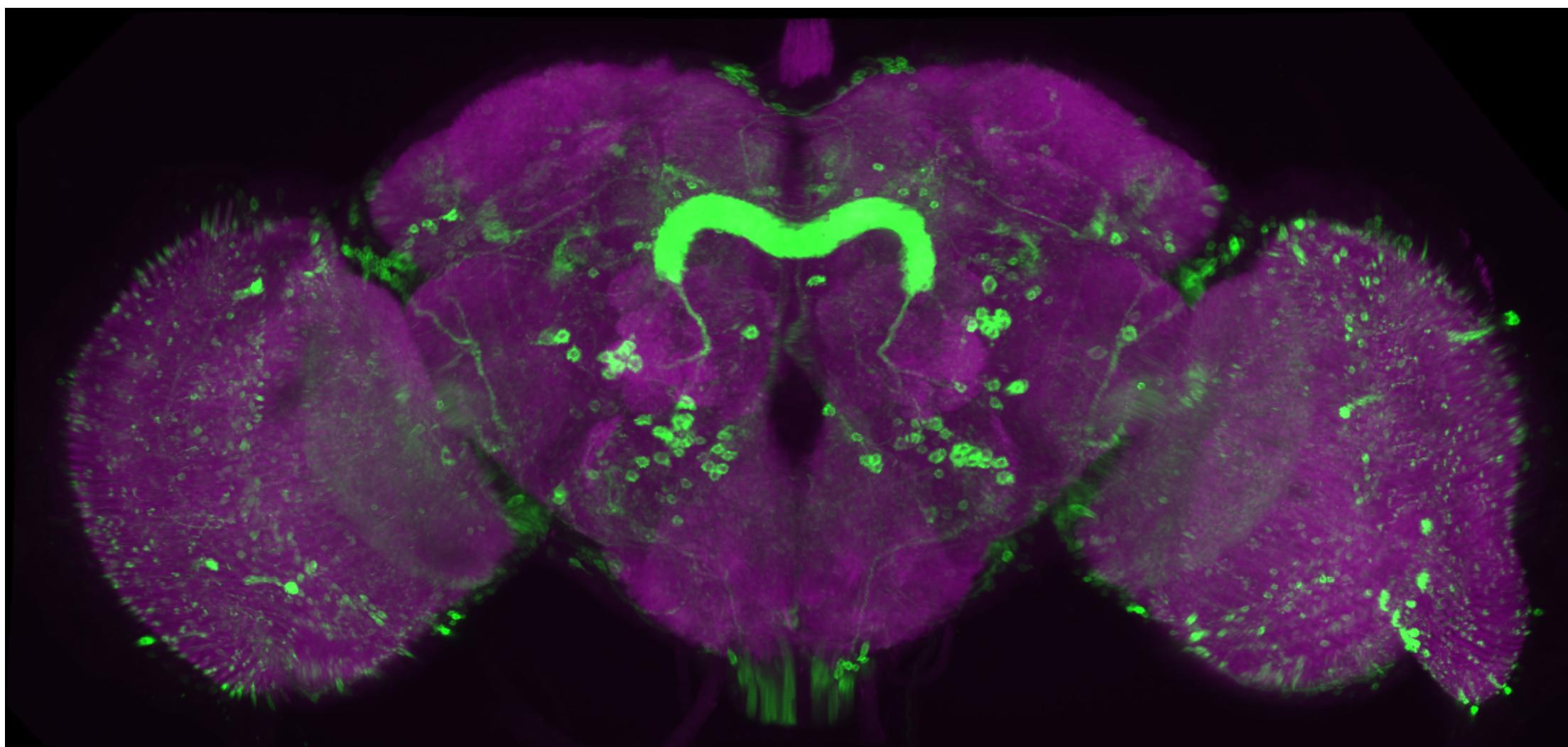
Polarity Case 5



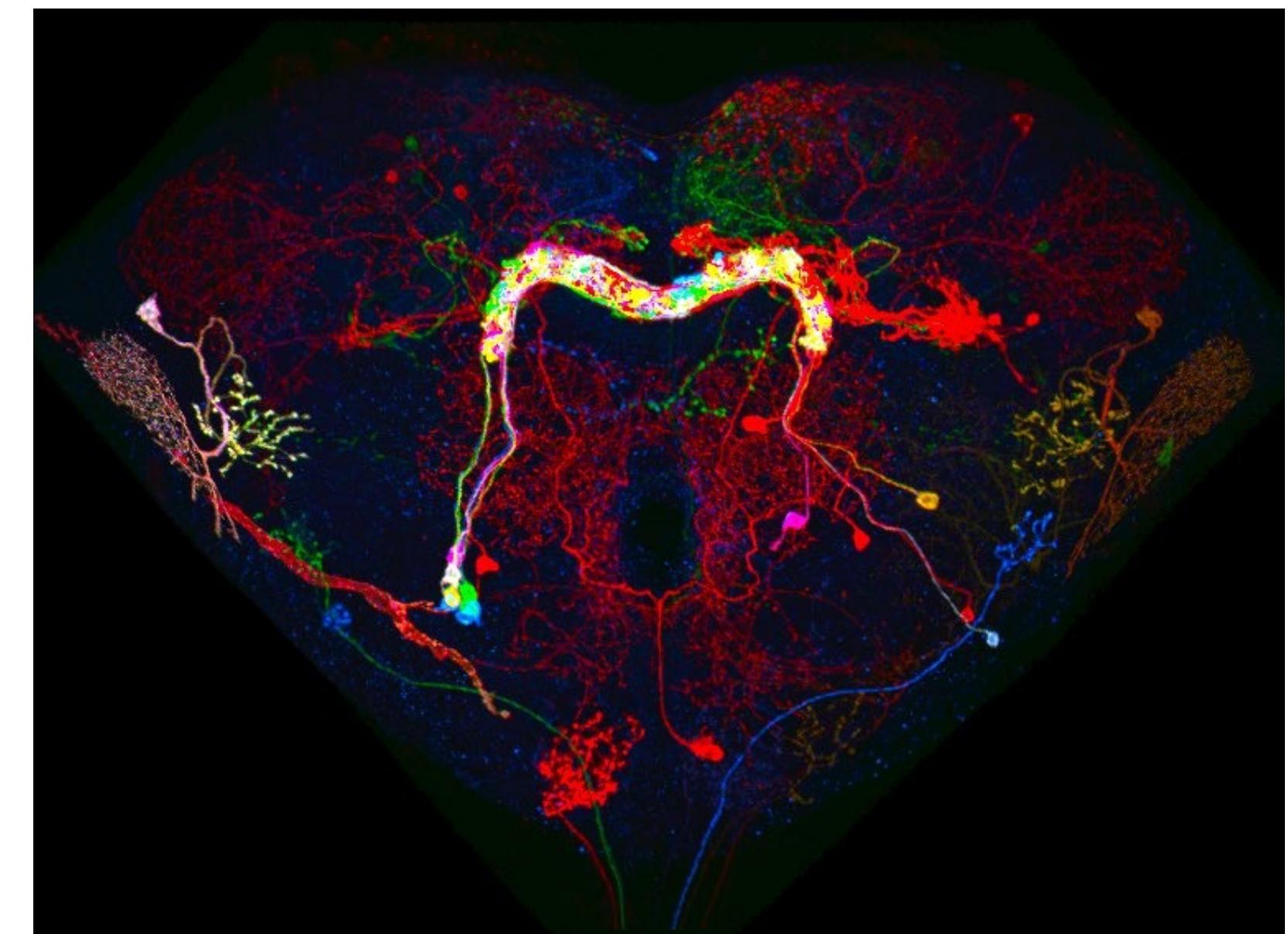
- UAS Reporters:
- Membrane: 20XUAS-Cs-Chrimson-mVenus trafficked in attP18
- Synapse: UAS-Syt-HA (different from Case 3)
- Chrimson-mVenus instead of myr-FLAG
 - Maximizes consistency with screening & physiology results
- Labels presynaptic terminals, but less accurate intracellular localization than Case 3
- Very low background in most cases
- Common contaminants can grow during IHC and be labeled by anti-GFP antibody, requiring sodium azide suppression

MultiColor FlpOut (MCFO) subdivides GAL4 & split-GAL4 lines

Full GAL4 pattern with UAS-Chrimson-mVenus reporter



Subsets of neurons labeled in different colors by MultiColor FlpOut (MCFO)



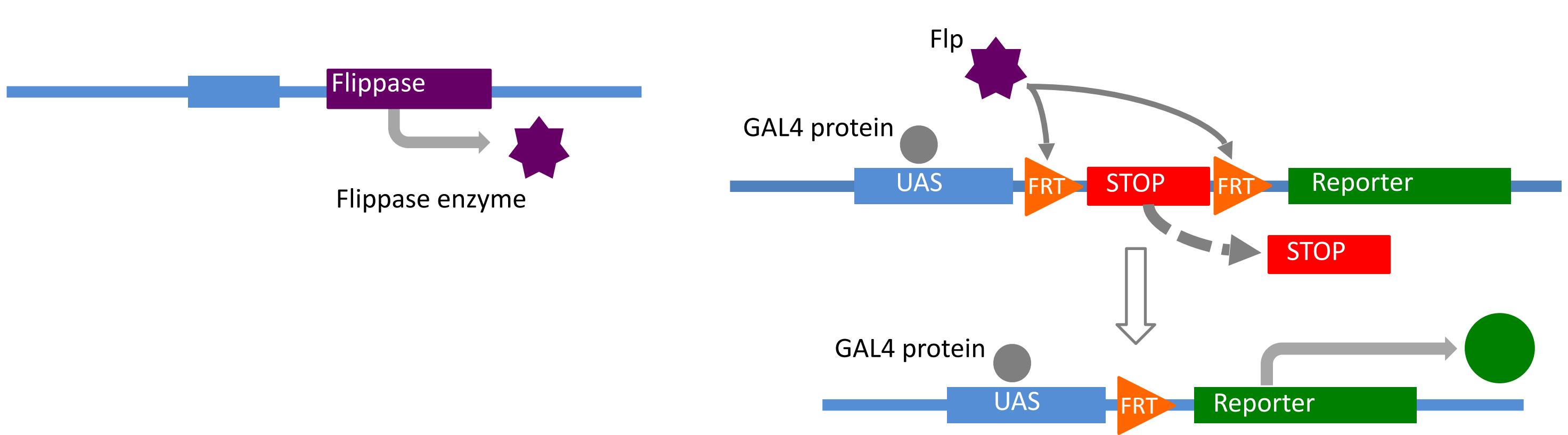
Multicolor FlpOut (MCFO)

MCFO-1 UAS Reporter:
pBPhsFlp2::PEST in attP3;
pJFRC201-10XUAS>STOP>myr::smGFP-HA in
VK0005,
pJFRC240-10XUAS>STOP>myr::smGFP-V5-
THS-10XUAS>STOP>myr::smGFP-FLAG in
su(Hw)attP1

Multicolor Flp-out				
Target	Reference neuropil	neuronal membrane 1	neuronal membrane 2	neuronal membrane 3
Genetic marker	endogenous	UAS>>HA	UAS>>V5	UAS>>FLAG
Primary/Direct Ab	mouse nc82	rabbit anti-HA	DL550 anti-V5	rat anti-FLAG
Secondary Ab	AF488 anti-mouse	AF594 anti-rabbit		ATTO647N anti-rat

See Nern et al., 2015 for
more details on MCFO
variations

<https://doi.org/10.1073/pnas.1506763112>



63X Adult Tiles - Brain

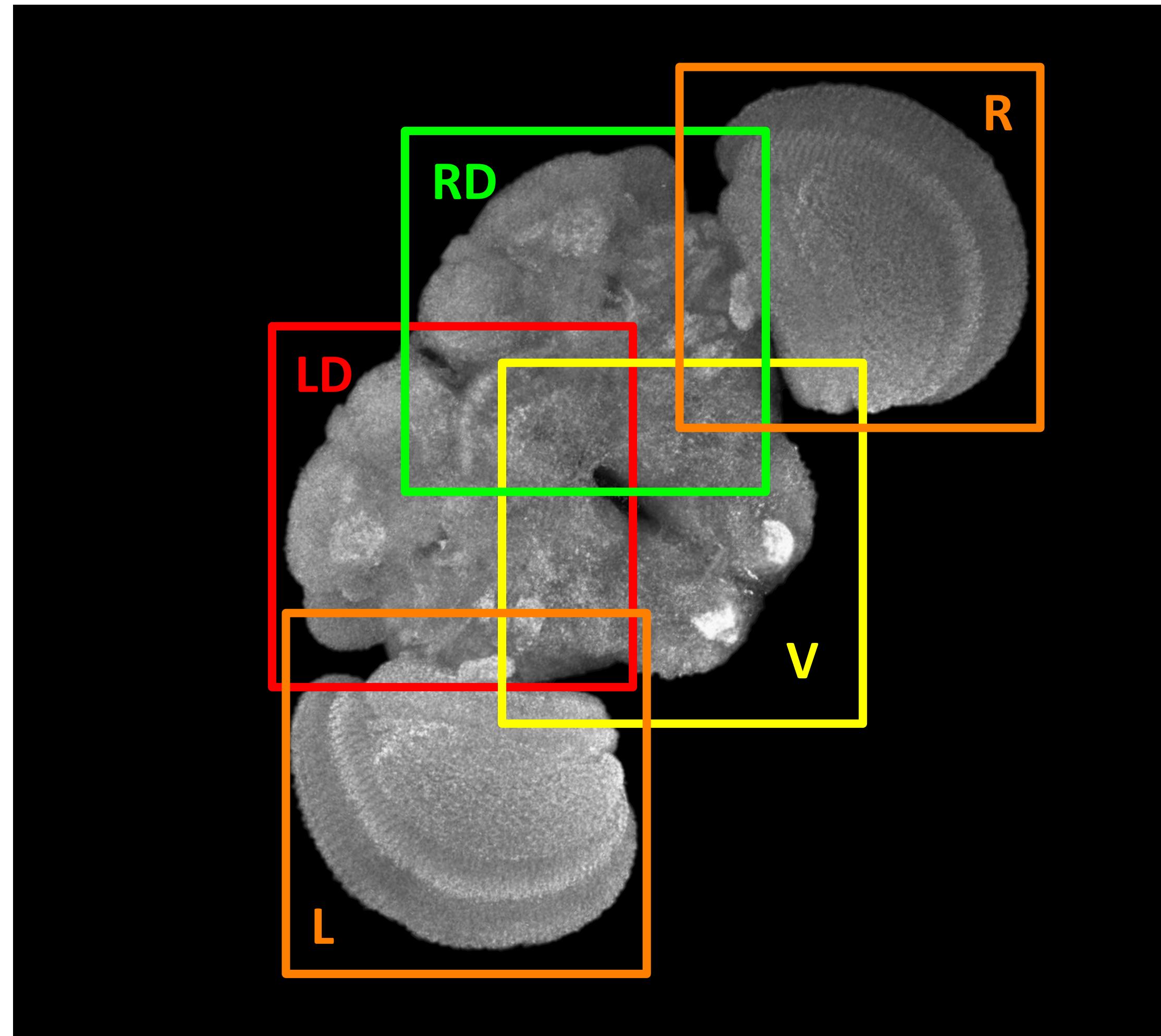
L – Left Optic Lobe

V – Ventral

LD – Left Dorsal

RD – Right Dorsal

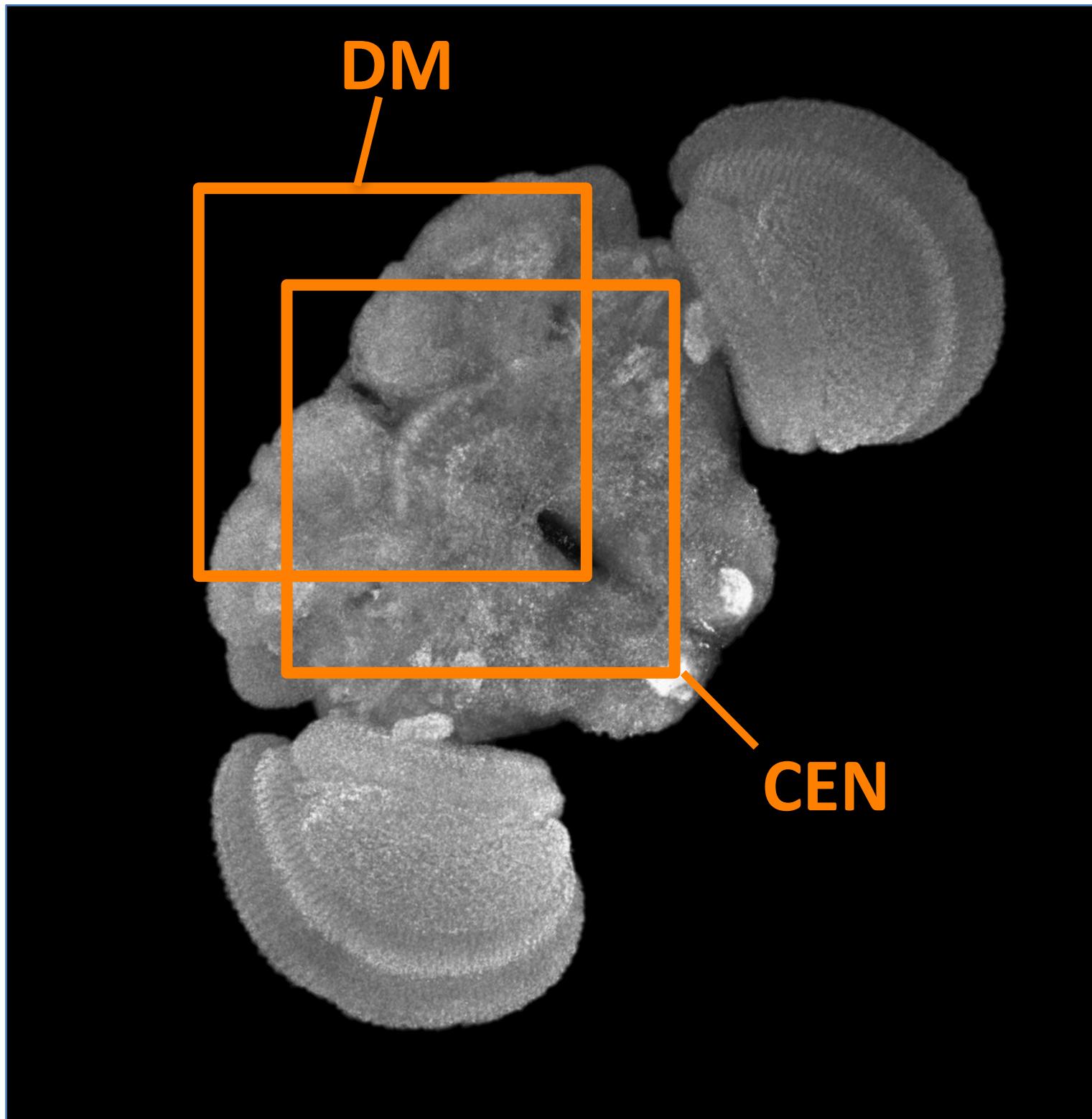
R – Right Optic Lobe



63X Adult Tiles – Additional Brain Tiles

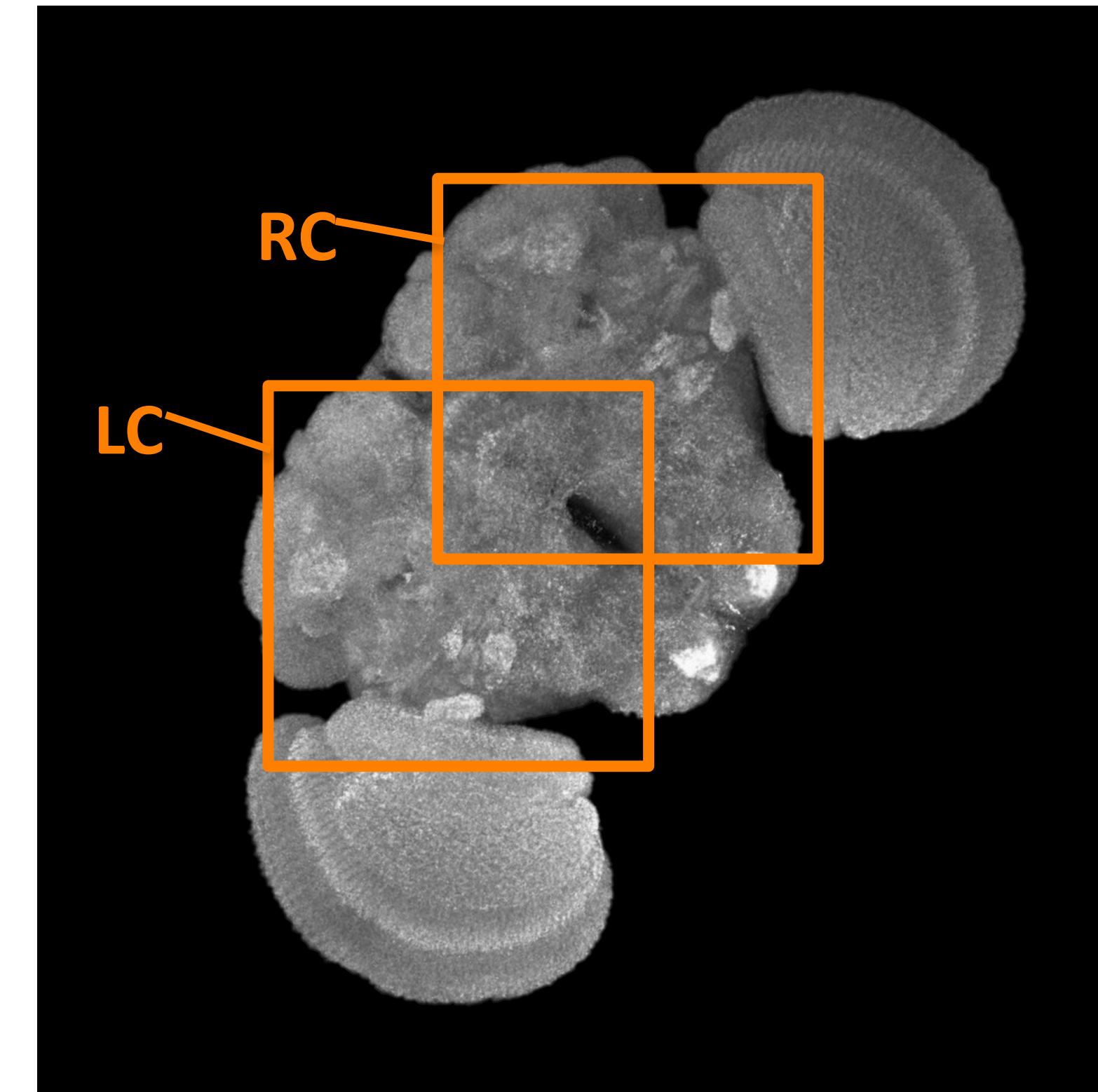
DM – Dorsal Medial

CEN – Central



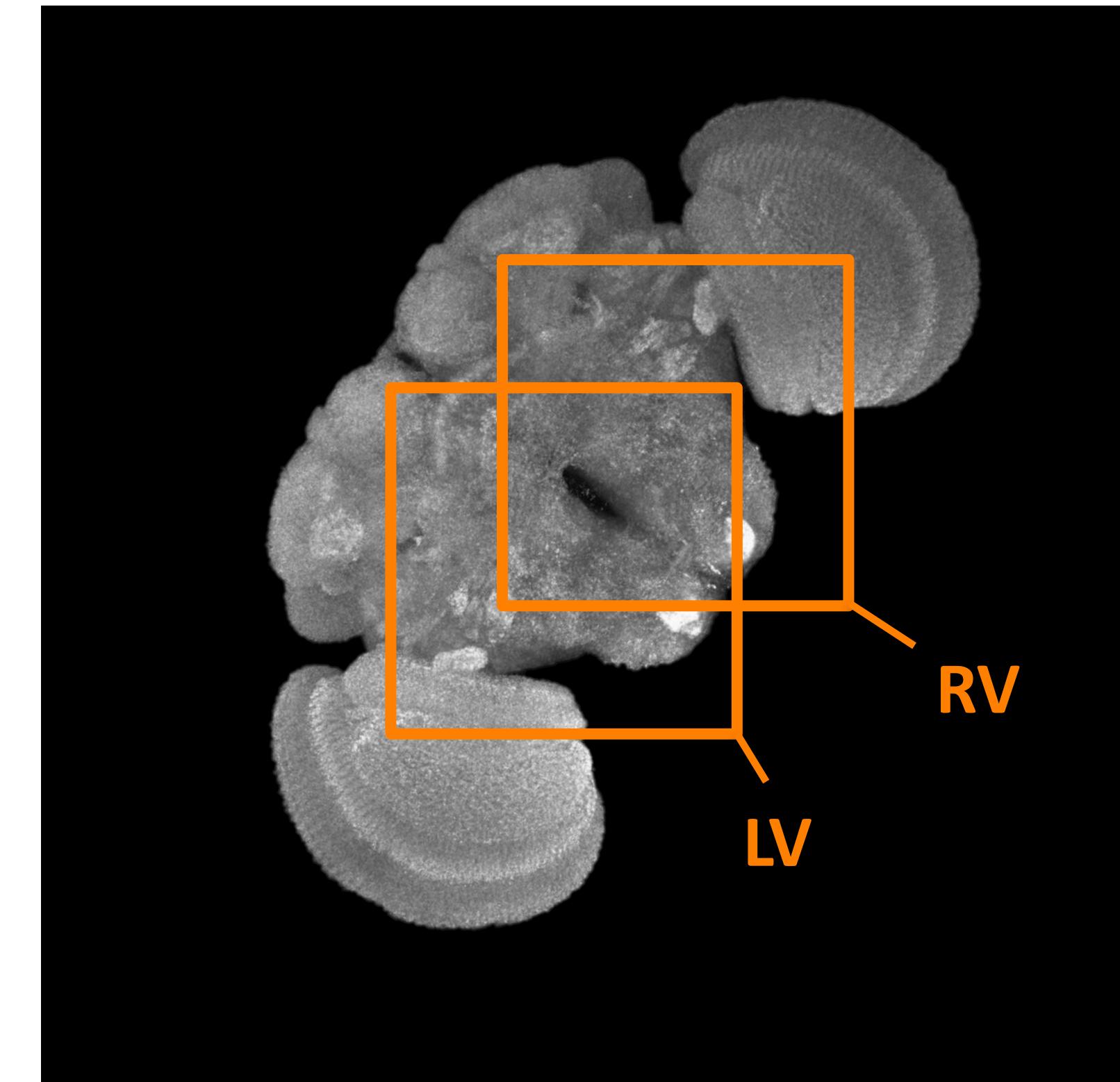
LC – Left Central

RC – Right Central



LV – Left Ventral

RV – Right Ventral



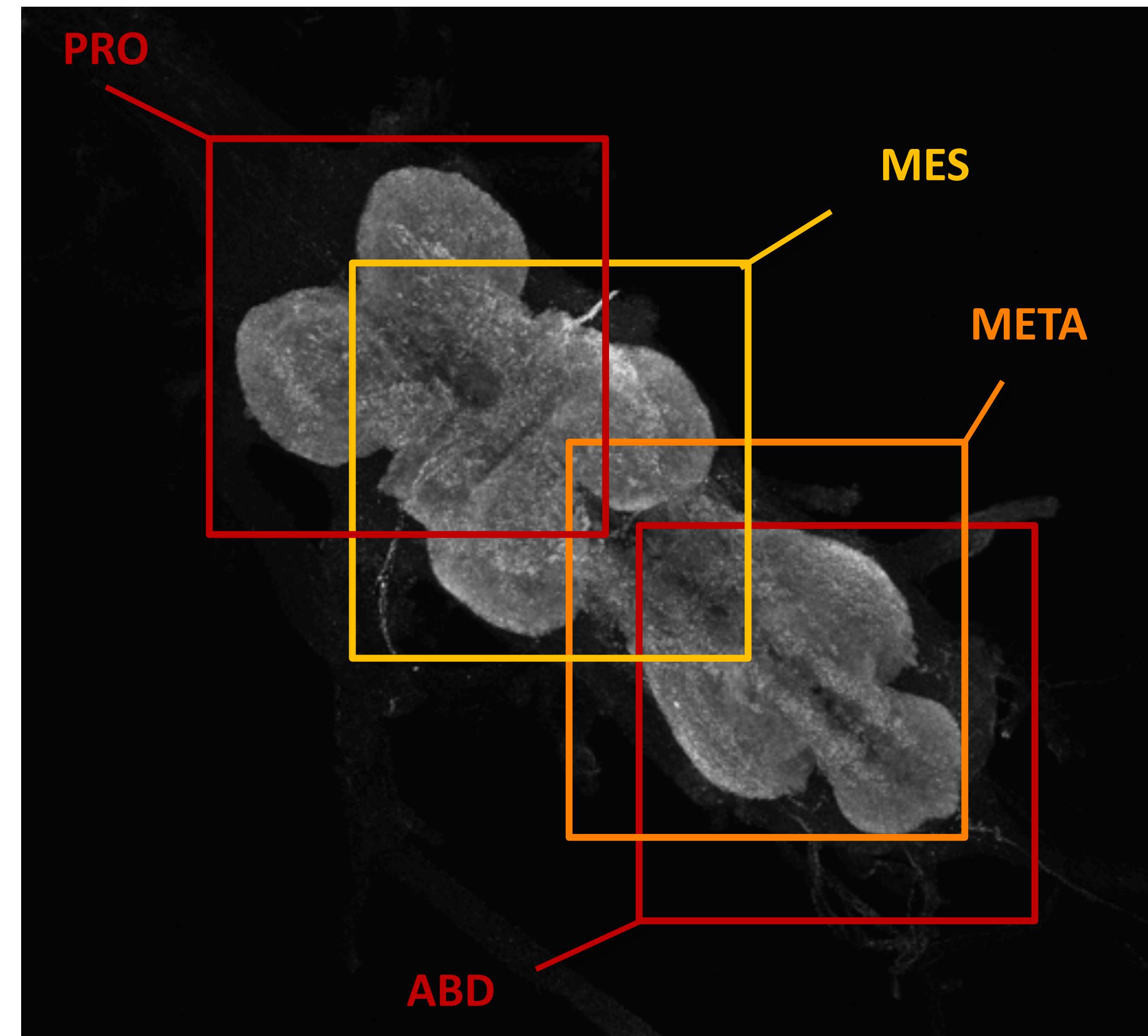
63X Adult Tiles - VNC

PRO – Prothoracic

MES – Mesothoracic

META – Metathoracic
(sometimes covers Abd)

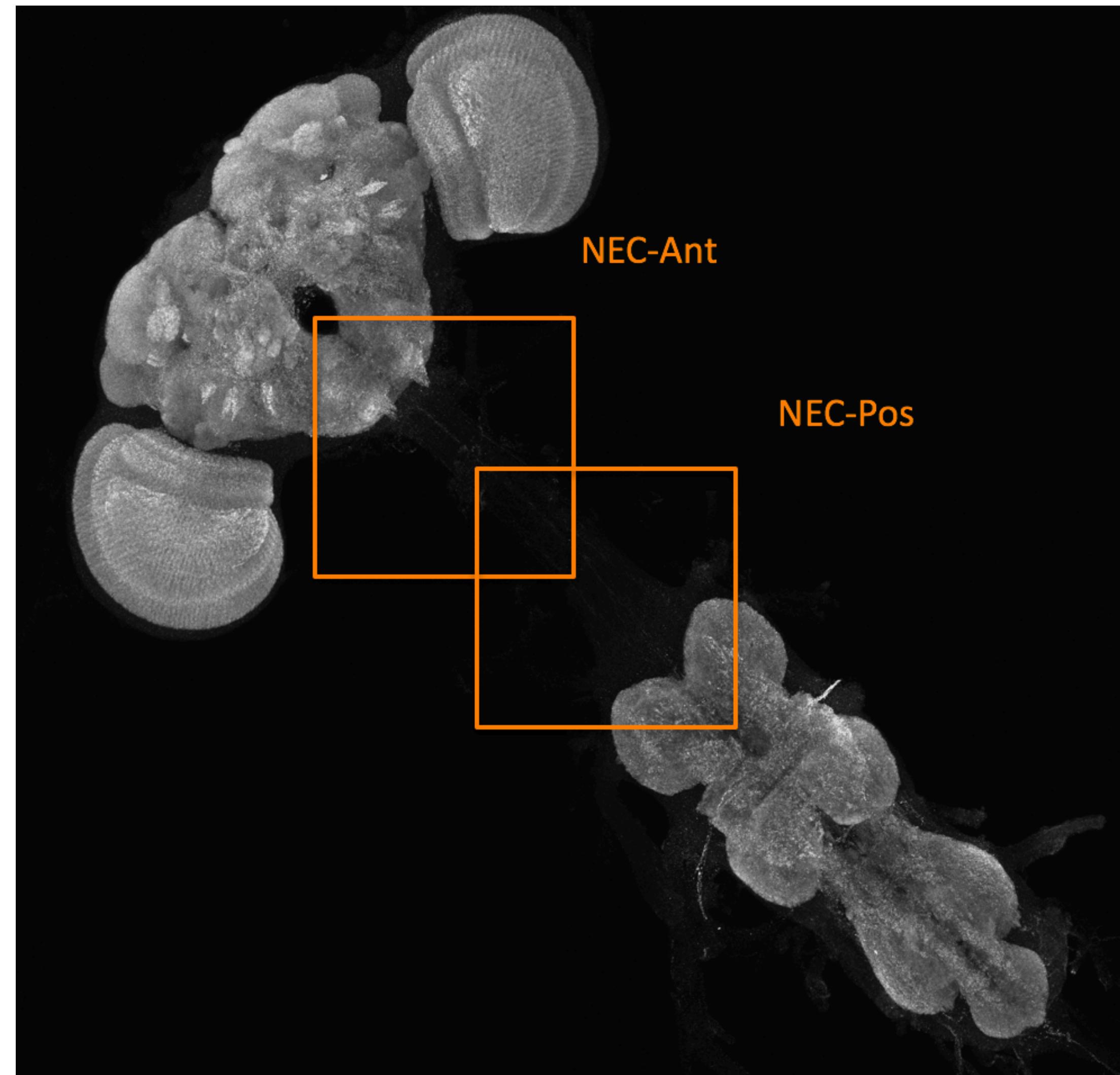
ABD – Abdominal



63X Adult Tiles - Neck

NEC-Ant – Anterior Neck

NEC-Pos – Posterior Neck



40X Adult Tiles - Brain & VNC

BRAIN – 1 tile:

CEN - Central brain

VNC – 2 tiles:

PRO - Prothoracic (anterior VNC)

META - Metathoracic (posterior VNC)

