

Atomistic Simulation of Voltage Activation of a Truncated BK Channel

Zhiguang Jia, Jianhan Chen 

Department of Chemistry, University of Massachusetts, Amherst, United States

 https://en.wikipedia.org/wiki/Open_access

 Copyright information

Reviewed Preprint

v3 • July 9, 2025

Revised by authors

Reviewed Preprint

v2 • June 20, 2025

Reviewed Preprint

v1 • April 23, 2025

eLife Assessment

This **valuable** study addresses the structural basis of voltage-activation of BK channels using atomistic simulations of several microseconds, to assess conformational changes that underlie both voltage-sensing and gating of the pore. The findings, including movement of specific charged residues, combined with the degree to which these movements are coupled to pore movements, provide a **solid** basis for understanding voltage-gating mechanisms in this class of channels. This paper will likely be of interest to ion channel biologists and biophysicists focused on voltage-dependent channel gating mechanisms.

<https://doi.org/10.7554/eLife.105895.3.sa3>

Abstract

Voltage-dependence gating of ion channels underlies numerous physiological and pathophysiological processes, and disruption of normal voltage gating is the cause of many channelopathies. Here, long timescale atomistic simulations were performed to directly probe voltage-induced gating transitions of the big potassium (BK) channels, where the voltage sensor domain (VSD) movement has been suggested to be distinct from that of canonical Kv channels but remains poorly understood. Using a Core-MT construct without the gating ring, multiple voltage activation transitions were observed at 750 mV, allowing detailed analysis of the activated state of BK VSD and key mechanistic features. Even though the S4 helix remains the principal voltage sensor in BK, its vertical displacement is only ~ 3 Å and accompanied by significant lateral movements. The nature of the predicted VSD movement is in strong agreement with recent Cryo-EM structural studies of mutant BK channels with constitutively activated VSD. Free energy analysis based on the predicted activation transition yielded a total gating charge of $0.44 e$ per VSD, consistent with the experimental range of $0.48 - 0.65 e$. We further show that the ability of modest physical movements with a small total gating charge to drive effective voltage gating of BK can be attributed to large gradients in the local electric field as reshaped by the protein. Furthermore, the S4 movement is coupled to the pore opening through a non-canonical pathway that involves the tightly packed S4-S5-S6 interface. These distinct mechanistic features may be relevant to voltage gating of other ion channels where VSDs are not domain-swapped with respect to the pore-gate domain.

Significance Statement

The big potassium (BK) channel is the only potassium channel that integrates intracellular calcium signaling with membrane depolarization. It is considered one of the most important channels in cardiovascular and neurological disorders. It has been known that voltage gating of BK channels has distinct features compared to the canonical voltage gating mechanism established through studies of voltage-gated potassium (Kv) channels. Yet, little is known about the molecular nature of voltage sensing and pore activation of BK channels at present. Our work reports the first successful direct simulation of voltage-dependent activation of the big potassium (BK) channel, revealing novel voltage sensing and sensor-pore coupling mechanisms that have largely eluded the community until now.

Introduction

Voltage-dependent gating of ion channels mediates numerous physiological and pathophysiological processes (1–5), by regulating ion flows in response to membrane potential changes in excitable cells such as cardiac myocytes and smooth muscle cells in the lung and blood vessels (6–11). Disruption of normal voltage gating is involved in many cardiovascular and neurological diseases, including epilepsy, mental retardation, chronic pain, hypertension, arrhythmias, stroke, and ischemia (12–20). Among the family of potassium channels, the large-conductance potassium (BK) channels stand out in several ways (21–27). It has the largest single-channel conductance (up to ~300 pS) and is the only known K⁺ channel to be activated by both intracellular Ca²⁺ and membrane potential. Functional BK channels are homo-tetramers, with each subunit consisting of a seven-helix transmembrane domain (TMD) and a C-terminal Ca²⁺-sensing cytosolic domain (CTD) (Figure 1a). The deactivated state of BK channels contain a physically open central pore (28–30), lacking the classical bundle crossing constriction at the intracellular entrance observed in the voltage-dependent potassium (Kv) channels (31). Instead, BK channels likely follow the hydrophobic gating mechanism, where the pore undergoes hydrophobic dewetting transition to create a vapor barrier to block ion permeation in the deactivated state (32–36). Furthermore, the voltage sensor domains (VSDs) of BK channels are not domain-swapped with respect to the pore-gate domain (PGD), in contrast to the domain-swapped TMD organization of Kv channels (Figure 1b–c). The difference in TMD organization is likely a key factor underlying important but still poorly understood differences in the voltage sensing and gating mechanisms of BK channels (37–41) in comparison to the “canonical” framework established mainly through the study of Kv channels (42–50).

In Kv channels, the linker connecting transmembrane (TM) helices S4 and S5 forms a α -helix (~15 aa) that wraps around the S6, with the S4 of one subunit interacting with the S5 of a neighboring subunit and simultaneously connecting to the S5/S6 within the same subunit through the S4-S5 linker. Conversely, in BK channels, the S4-S5 linker is a short loop (~5 aa), and the S4 solely interacts with S5 within the same subunit (Figure 1). Within the canonical voltage activation of Kv channels, each of the first four Arginine residues in the VSD S4 accounts for about 1e gating charge (51–54), giving rise to a total gating charge of ~3–3.5 e per monomer. Membrane depolarization drives S4 to move upward by ~8 Å, which causes the S4-S5 linker helix to pivot upwards and change the interactions with S6 and releases the S6 helix bundle crossing to physically open the gate (55–57). For BK channels, despite sharing many conserved charged residues, the total number of gating charges is only 0.6 e per VSD (22, 58–62). In addition to Arginine residues on BK S4 (R207, R210 and R213), other residues also appear to contribute to voltage sensing, such as D153 and R167 on S2, D186 on S3 and E219 on S4 (39, 63, 64). Furthermore, fluorophore quenching suggested that the movement of BK VSDs during voltage

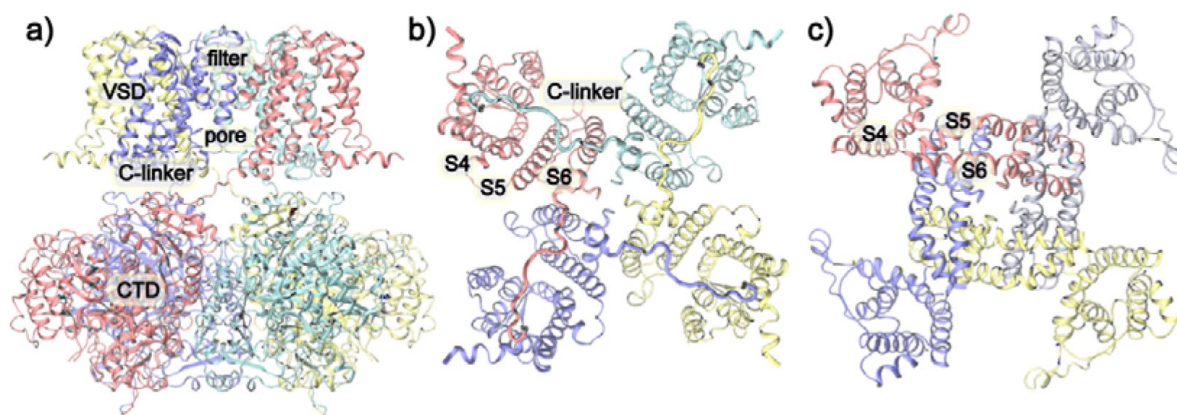


Figure 1.

Overall structures of BK and Kv channels.

a) The overall structure of BK channels in the Ca^{2+} -free state (PDB: 6v3g) with key domains and regions labelled. Each monomer is shown in the same color. **b)** Top view of the TMD of BK channels, showing the non-domain-swapped VSD/PGD configuration. **c)** Top view of the TMD of Kv 1.2 channel (PDB: 3lut), where the VSDs and PGDs are domain-swapped. Note the much tighter packing of S4 from VSD and S5/S6 from PGD in BK channels.

gating was smaller and involved both vertical and lateral motions (65–67). At present, there remains significant ambiguity in the identity of gating charges, details of VSD motions, and how they drive the pore opening in BK channels.

In this study, extensive atomistic molecular dynamic (MD) simulations up to 10 μ s in length were performed in explicit solvent and membrane to directly probe the voltage-driven activation of BK channels in the Ca^{2+} -free state, using the Core-MT construct that does not include the CTD but retains voltage gating (68, 69). We were able to directly observe spontaneous opening transitions and ion conductance of the channel under 750 mV membrane voltage within the 10 μ s simulation timescale. The observed VSD movements appear to be highly consistent with two recent Cryo-EM structural studies of mutant BK channels with constitutively activated VSDs (38, 41). Free energy analysis was performed to further quantify the total gating charge as well as the contributions of key charged residues. The results are in quantitative agreement with available experimental data (39, 63, 64), providing further support for the predicted voltage sensing and gating mechanism. The analysis reveals central roles of voltage-induced displacement of R210 and R213 on S4 in voltage sensing and the strong interactions at the S4-S5-S6 interface in VSD-pore coupling. This mechanism was further validated using steered MD simulations showing that pulling on the charged tip of R210 and R213 alone could readily drive pore opening in Core-MT BK channels. Taken together, the current work provides for the first time a reliable detailed molecular mechanism of BK voltage activation, which is distinct from the canonical voltage gating mechanism of Kv channels. These novel voltage gating principles are likely relevant in understanding the gating and regulation of other non-domain swapping ion channels.

Results and Discussion

Direct atomistic simulations of voltage activation of BK channels

Starting from a fully equilibrated closed state, multiple atomistic simulations were performed at 0 and 750 mV membrane voltages for up to 10 μ s to directly probe voltage-driven activation of Core-MT BK channels using the special purposed supercomputer Anton 2 (70, 71) (see Methods; Figure S1). The simulated voltage of 750 mV is higher than the experimental conditions, where $V_{1/2}$ of Core-MT BK channels is ~ 244 mV compared to ~ 180 mV for the full-length BK channel (69), to accelerate protein conformational transitions. Similar voltages have been used in atomistic simulations and found to generate realistic transitions (56, 72). We compare the membrane thickness at 300 and 750 mV and the results reveal no significant difference in the membrane thickness (Figure S2). At 0 mV, the Core-MT BK channel remained highly stable in an apparently closed state, with the pore fully dehydrated and not permeable to ions (Figure S1, top row). Similar to what has been observed previously in simulations and cryo-EM maps (30, 32–35, 38), lipid tails can enter the dewetted pore through the fenestration gap between the pore lining S6 helices. They are highly dynamic and likely contribute to the stability of the dewetted state of the pore. Note that the absence of CTDs increases the flexibility of pore-lining S6 helices, and the fully relaxed pore profile (red trace in Figure S1d, top row) shows substantial differences compared to that of the Ca^{2+} -free Cryo-EM structure of the full-length channel (black trace). Importantly, the VSDs were highly stable and showed minimal movements (~ 1 Å or less) for all charged groups as well as the TM helices themselves.

At 750 mV, significant movements were observed with multiple charged groups on S4 (Figure 2a and Figure S1). In particular, the guanidium groups of R210 and R213 moved upward along the membrane normal (z-axis) by up to ~ 8 and ~ 10 Å, respectively, plateauing after ~ 5 μ s (e.g., see Figure 2a). Other VSD charges showed much smaller z-displacements. The apparent modest movements of ~ 3 –4 Å of E219 and R207 seem to be mainly a result of the upshift of S4 itself, which reached a maximum of ~ 3 Å during the second half of the simulation (Figure 2b). In contrast,

other VSD helices S1-S3 exhibited much smaller z-axis movements (~ 1 Å). As illustrated in supplementary Movie S1, the large charge displacements of R210 and R213 charges with modest S4 movement were enabled by side chain snorkeling.

Voltage-driven VSD movements were apparently coupled with an opening transition of the pore, allowing one to directly observe Core-MT activation and K^+ conductance during both 10- μ s atomistic simulations on Anton 2 (Figure S1). As illustrated in **Figure 2** and Movie S2, the backbone root-mean-squared distance (RMSD) of the pore from the open state decreased sharply from ~ 4.5 Å around the 4 μ s mark to below ~ 2.7 Å around the 5 μ s mark, as the S4 helix shifted up along z-axis by ~ 2 Å during the same time span. The pore opening transition mainly involved the increase of the tilt of S6 segments below the glycine hinge (residues 313-324) (**Figure 2d**). The pore continued to move closer to the open state after the initial rapid response to VSD activation, eventually reaching a state that has similar S6 helix tilt and is only ~ 2.2 Å from the Ca^{2+} -bound Cryo-EM structure at the end of the 10 μ s run (**Figure 2e**, red vs. cyan cartoons). These pore structural changes roughly doubled the pore diameter, from ~ 10 Å to ~ 20 Å at the intracellular entrance ($z \sim -20$ Å; **Figure 2f**). It should be noted that the full-length BK channel in the Ca^{2+} bound state has an even larger intracellular opening (**Figure 2f**, green trace), suggesting that additional dilation of the pore may occur at longer timescales, or in response to Ca-binding to the full-length channel. On the other hand, the simulation construct does not include the 11-residue Kv mini-tails required for assembly and membrane insertion of Core-MT (68), which could impact the pore conformation. Furthermore, the single-channel conductance of Core-MT is $\sim 30\%$ lower (68), suggesting that its open pore is more constrictive than that of the full-length BK channels.

The opening transitions were accompanied with rehydration of the pore and the channel became conductive to K^+ (**Figure 3** and Movie S3). Consistent with the smaller pore diameter, the final states of the pore in the simulations only accommodate ~ 30 waters, compared to ~ 40 waters for the Ca^{2+} -bound state of full-length BK (32). Furthermore, the single-channel conductance estimated from the last 2 μ s of *sim 2b* (**Figure 3a**) is only ~ 1.5 pS, much lower than the experimental value of ~ 220 pS for Core-MT (69). We note that classical force fields, such as CHARMM36m used in the current simulations, are known to underestimate the single channel conductance by about one order of magnitude (73). Indeed, the fully opened state of Core-MT, constructed from the Ca^{2+} -bound Cryo-EM full-length BK structure, is predicted to have a conductance of ~ 6 pS using the same simulation setup (*sim 7*; Figure S3), which is only ~ 4 fold of that of the final state from the voltage activation simulations. We further note that subconductance open states have been observed in single channel recordings of BK channels (74–77). Besides the limitation of the current fixed charge force fields in quantitatively predicting channel conductance, we note that the molecular basis for the large conductance of BK channels is actually poorly understood (78). It is noteworthy that the pore hydration level appears to be an important factor in determining the apparent conductance in the simulation, which has also been proposed in a previous atomistic simulation study of the *Aplysia* BK channel (33). Despite not reaching a fully conductive state, the ability of the dilated and hydrated pore to sustain K^+ permeation throughout the second half of the 10 μ s simulation demonstrates that the hydrophobic gate in the closed channel has been broken and the channel enters a conductive state. Consistent with previous simulations of other K^+ channels (78–80), the conductance follows a multi-ion mechanism, where there are at least two K^+ ions inside the filter, preferentially occupying positions S1/S3 or S2/S3 (**Figure 3b**). The two bound ions subsequently move to S0/S1 when an incoming ion takes the S3 position. Notably, between the ions inside the filter, there can be either one or no water molecule, indicating the coexistence of both soft and hard knock-on mechanisms.

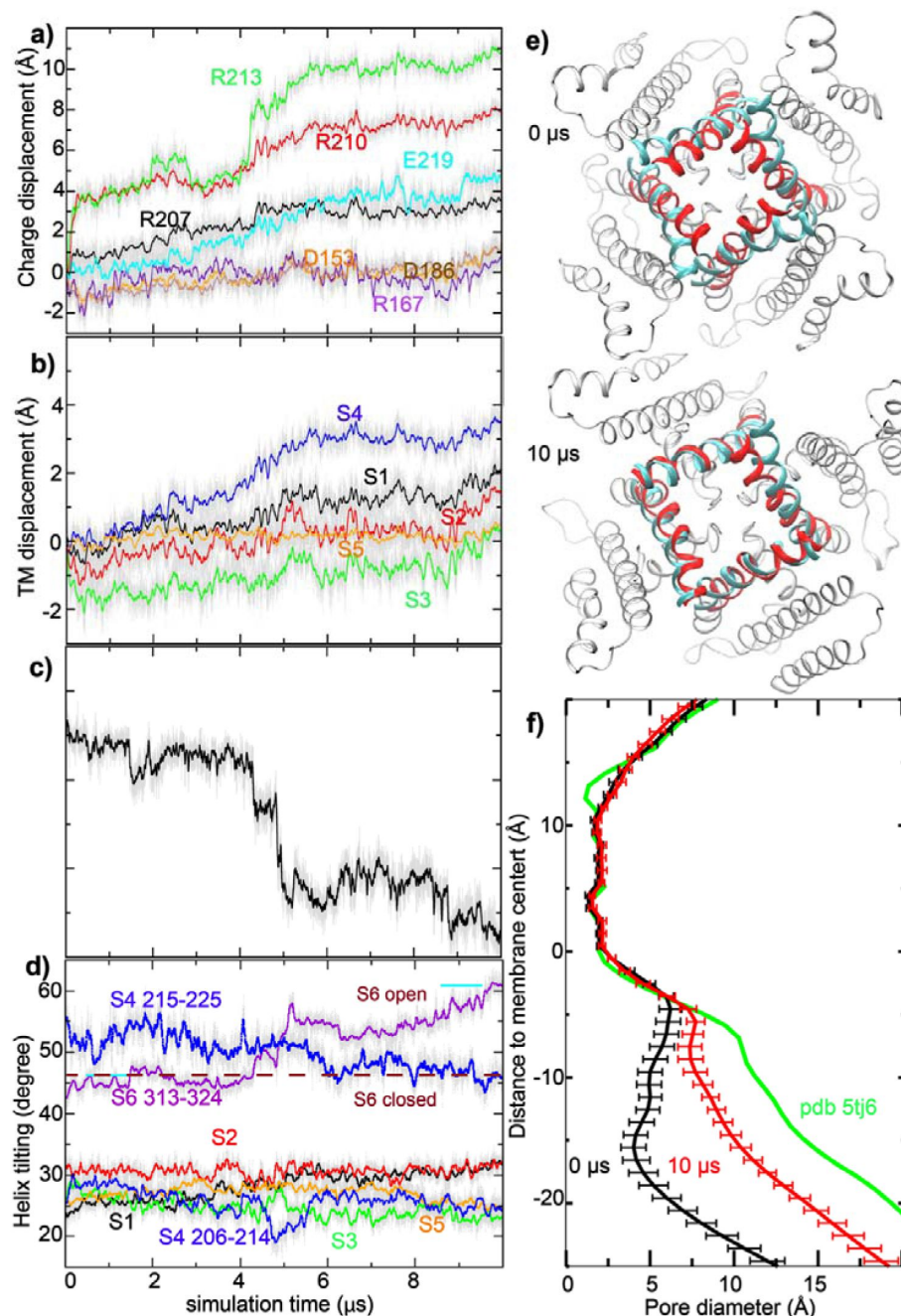


Figure 2.

Voltage activation of Core-MT BK channels.

a-d) Results from a 10- μ s simulation under 750 mV (*sim2b* in Table S1). Each data point represents the average of four subunits for a given snapshot (thin grey lines), and the colored thick lines plot the running average. **a)** z-displacement of key side chain charged groups from initial positions, **b)** z-displacement of centers-of-mass of VSD helices from initial positions, **c)** backbone RMSD of the pore-lining S6 (F307-L325) to the open state, and **d)** tilt angles of all TM helices. The locations of charged groups were taken as those of guanidinium CZ atoms (for Arg) and sidechain carboxyl carbons (for Asp/Glu). Only residues 313-324 of S6 were include in tilt angle calculation, and the values in the open and closed Cryo-EM structures are marked using purple dashed lines for reference in panel d. **e)** Superimposition of the initial (0 μ s) and final (10 μ s) structures of the pore (residues F307-L325; red cartoon), in comparison to the open Cryo-EM structure (cyan cartoon). The view shown is from the bottom (cytosolic side). **f)** Average pore profiles calculated from the first and last 0.1 μ s of *sim2b*, with error bars showing standard error. The pore profile derived from PDB 5tj6 (open state) is shown as a reference.

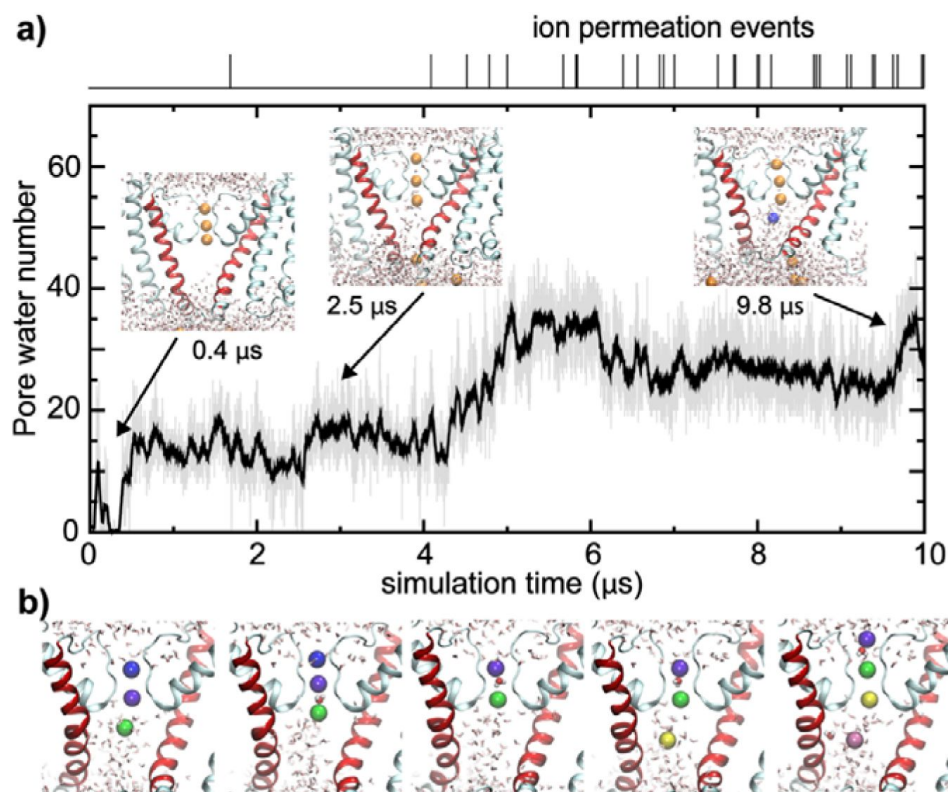


Figure 3.

Pore Rehydration and Ion Conductance.

a) The number of water molecules inside the pore as a function of time during simulation (*sim 2b*), with the upper panel showing recorded ion permeation events. Inserts show snapshots of the pore region at three representative timepoints. **b)** Snapshots illustrating key steps of K⁺ ions passing through the filter. Potassium ions inside or near the filter are colored according to their identities. The water molecule bridging two ions inside the filter is also shown as van der Waals spheres.

Gating charges: how small VSD movements support voltage sensing

The ability of Anton 2 simulations to directly observe voltage-driven activation of Core-MT BK channels allows one to examine the molecular details of VSD activation and VSD-pore coupling. An important observation is that, even though the charged groups of R210 and R213 show z-displacement of 8-10 Å ([Figure 4a](#)), the overall movement of S4 along the membrane normal is only ~3 Å, less than a single α -helical turn ([Figure 2b](#)). This is considerably smaller than that observed in canonical Kv channels, which has been estimated to be ~8 Å ([55](#)) and up to 15 Å ([56](#)). Smaller S4 movements have been suggested in fluorophore quenching experiments ([65](#)–[67](#)) and molecular simulations ([39](#)). Recent Cryo-EM structures of mutant BK channels with constitutively activated VSDs actually reveal minimal S4 movements along the membrane normal ([38](#), [41](#)). Upon voltage activation, R210 switches the salt-bridge partner from D186 on S3 to D153 on S2 and D133 on S1, R213 switches to engage with D153, and R207 switches from engaging with D153 and D133 to becoming exposed to the water/membrane interface ([Figure 4b](#)). Similar movements were also observed in recent high-resolution Cryo-EM structures of R207A mutant BK channels with constitutively activated VSDs at 0 mV ([41](#)), even though the net z-displacements of R210 and R213 charges in Cryo-EM structures are about ~2-4 Å smaller. Both the smaller side chain charge movements and a lack of overall S4 z-displacement in the mutant Cryo-EM structures may be due to the absence of membrane voltage. To further evaluate if z-placements observed at 750 mV are an artifact of unphysical voltage, three independent simulations were initiated from the final state of *sim 2b* at 300 mV for 1.0 μ s (*sim 9* in Table S1). The results, summarized in Figure S1, show that the activated state of VSD remained stable and the pore stably hydrated in all three simulations.

Another important feature is that multiple helices in VSD underwent substantial outward movements in the membrane lateral direction in addition to the modest z-displacement of S4. As shown in [Figure 4c](#), both S1 and S4 displayed ~3 Å outward movements within the x-y plane. These lateral movements are apparently important for driving the opening motion of the pore-lining S6 helices ([Figure 4d](#)), likely mediated by direct S4-S5-S6 interactions (see the next section). Importantly, the predicted VSD movements appear highly consistent with observations from fluorophore quenching experiments ([65](#)–[67](#)), which suggest that S4 likely moves towards S3 and closer to S1/S2 upon activation. Interestingly, the simulations predict that S1 moves during voltage-activation instead of S2, in contrast to the previously proposed model ([66](#)). Lateral movements of S4 at the cytosolic end have also been observed in recent Cryo-EM structures of R202Q mutant *Aplysia* BK channel (equivalent to R213Q in human BK), which presumably locks the VSDs in the activated state ([38](#)), even though the overall movements of VSD are much more subtle in the Cryo-EM structures. The later may again be a direct consequence of the absence of membrane voltage.

To understand how the modest movements of VSDs can support effective voltage sensing in BK channels, we calculated the average electrostatic potential maps for the resting and activated states ([Figure 5](#)). The results reveal how the protein significantly remodels the local electric field, creating large gradients along both the membrane normal and lateral directions. This is similar the electric field focusing effects proposed for VSDs of both Kv and BK channels ([39](#), [81](#)). In particular, residues R210 and R213 reside in a region of high electrostatic potential (> 1100 mV) in the resting state, but move to a region of much lower electrostatic potential (~ 600 mV) in the activated state. Therefore, despite the modest S4 movements, charges on R210 and R213 can effectively sense a voltage change of ~500 mV, compared to the total imposed membrane voltage of 750 mV.

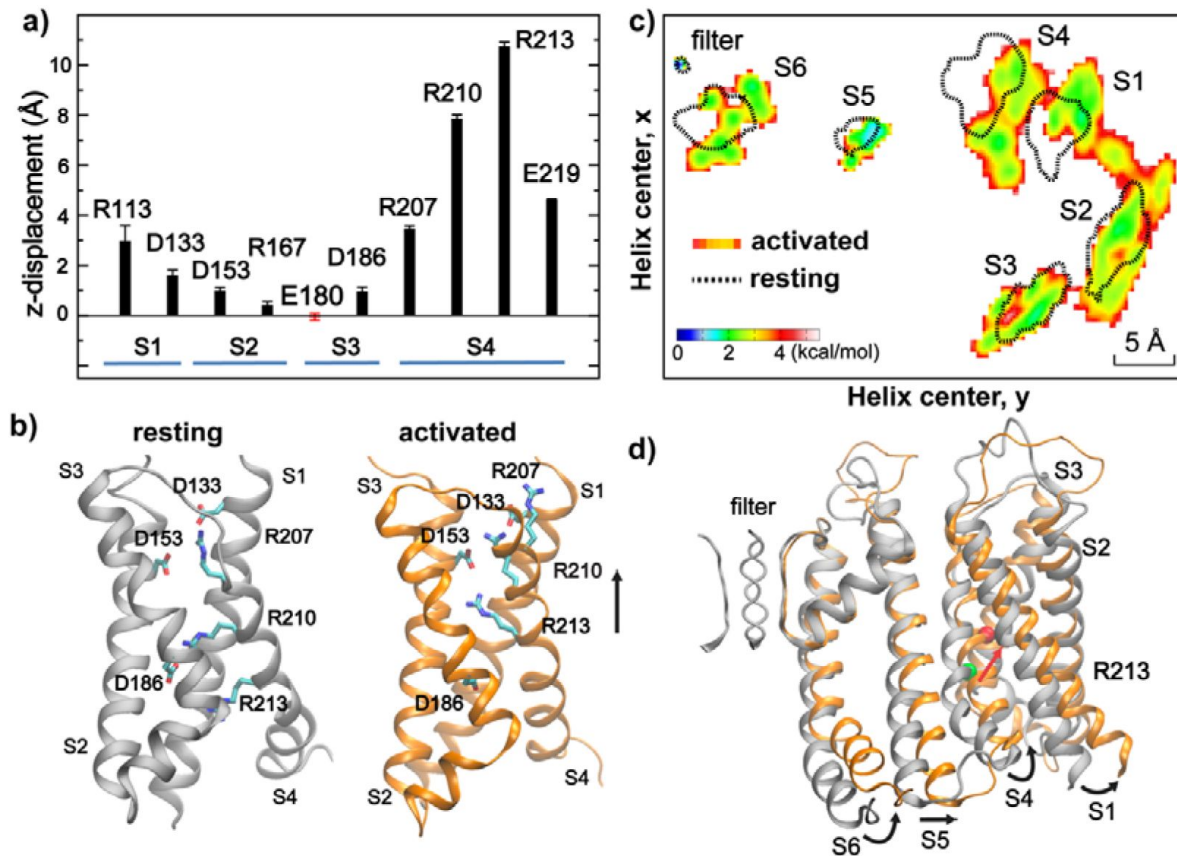


Figure 4.

VSD gating charge and voltage-sensing movements.

a) Average voltage-induced movements of key charges along the membrane normal (z-axis) with respect to the initial resting state structure, derived from last 500 ns of 750 mV simulation *sim 2b*. Error bars show the standard deviations. **b)** Conformations of key charged residues in the resting (silver) and activated (orange) states of BK VSD. The resting and activated states are represented using the snapshots at 0 and 10 μ s of *sim2b*, respectively. **c)** Distributions of centers-of-mass of TM helices along the membrane lateral directions (x and y) (view from the cytosolic side). The distributions for resting and activated states were derived from the first and last 500 ns of the 750 mV simulation *sim 2b*, respectively, which were converted into the free energy scale by $\sim R T \ln P(x,y)$ with $T = 300$ K. The contour for the resting state distribution (dotted lines) is drawn at 4 kcal/mol. **d)** Overlay of the resting (silver) and activated (orange) states of the TM domain of the Core-MT BK channel. The green and red spheres mark the backbone C_α atom of S4 R213 in the resting and activated states. Only one subunit is shown for clarity but all four filter loops shown for reference.

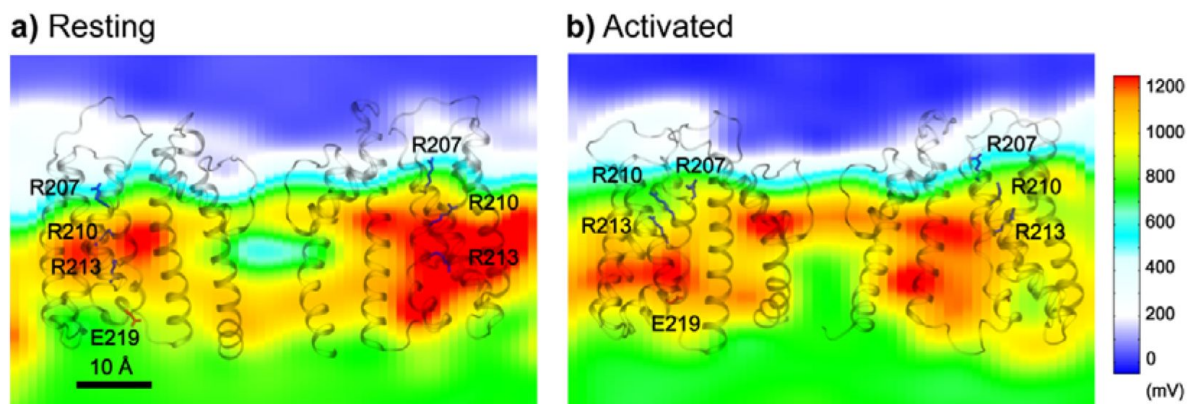


Figure 5.

Electrostatic potential fields of the Core-MT BK channel at 750 mV with resting and activated VSDs.

The fields were calculated as the averages of the first (resting) and last (activated) 250 ns of simulation *sim2b*. The fields are shown on a plane that goes through the filter and R210. Only two subunits of Core-MT BK are shown for clarity, with side chains of key S4 charges shown in sticks.

We performed free energy calculations (82, 83) to further quantify the contributions of key VSD residues to the total gating charge of BK channels (see Methods; Table S1 *sim* 3-6). As summarized in **Table 1**, the calculations estimated the total gating charge per VSD to be $\sim 0.45 e$, in strong agreement with the range of $0.48 - 0.65 e$ measured experimentally for the full-length BK channel (39, 63, 64). The results further identify R210 and R213 as the primary contributors to voltage sensing, accounting for approximately 97% of the total gating charge per VSD (**Table 1**). The calculated residue contributions to gating charge are highly consistent with the latest experimental measurements by Carrasquel-Ursulaez et al (2022) (39), even though earlier experimental studies disagreed on the contributions of several residues, including R207, R167, D153 and D186. It has been recognized that interpretation of gating charge measurements on mutant BK channels is nontrivial (39). For example, neutralizing R213 reduces the total gating charge from $2.62 e$ for WT to $1.31 e$ for R213C; yet replacing it with a negative charge (R213E) does not further reduce the net gating charge (63). The simulations reveal that the complication in experimental analysis may be attributed to modest movements of VSD in both membrane normal and lateral directions and a clear interplay of VSD conformation and local electric field (**Figure 5**). Nonetheless, atomistic simulations together with electrostatic potential and free analyses strongly support a central role of R210 and R213 in BK voltage sensing.

The dominant role of R210 and R213 in voltage sensing of BK channels was further tested by steered MD simulations (*sim* 8), where biasing potentials were applied to steer the movement of the guanidium CZ atoms of R210 and R213 from their positions in the initial resting state to those in the activated state in absence of membrane voltage (see Methods and Figure S4). The simulations show that upward movements of R210 and R213 guanidium tips alone can readily drive pore opening, which were observed in two out of four independent steer MD simulations (Figure S4). In the other two simulations, the S5-S6 packing was ruptured during the first 100 ns, likely due to the rapid rate of pulling. Similar to simulations under 750 mV membrane potential, pore dilation and increase in S6 tilting were observed as the VSD responded to the movement of R210 and R213, along with the breakdown of the hydrophobic barrier, as indicated by an increase in the number of water molecules within the pore. The ability of R210 and R213 charge upward movement alone to drive pore opening in absence of membrane voltage further supports the dominant roles of these two charges in voltage sensing and gating of BK channels.

Central role of the S4-S5-S6 interface in VSD-pore coupling

Examination of the covariance matrices (Figure S5) reveal that all TM helices, particularly, S1-S5, are tightly coupled in both resting and activated states. The lateral movements of S1 and S4 during activation mainly involve the cytosolic ends, concerted with those of S5 and S6 from the pore (**Figure 4b**). In the Cryo-EM structures, a kink is observed in S4 below R213, resulting in approximately a $\sim 30^\circ$ bend in the C-terminal half away from S5 and the pore. During voltage-induced activation, the N-terminal region of S5 maintained strong contacts with the S4 C-terminal segment (Figure S6), such that the upward and outward movement of S4 induces an increase in the tilt and bend of S5 (**Fig. 4d**). This S4-S5 movement in turn increases the tilt angle of S6 C-terminal half (F315 to E324) by $\sim 15^\circ$ to dilate the pore. In particular, the hydrophobic helix-helix contacts between S5 (L235, L239 and F242) and S6 (F315, V319 and I322) are strengthened, while those between N231 and K234 on S5 and E321 and E324 on S6 are weakened during this process (Figure S6). Besides pore dilation, the conformational response of S6 alters the orientation of the conserved E321 and E324 at the cytosolic entrance from membrane interface-facing in the closed state to pore-facing in the open state, as observed in Cryo-EM structures (28–30). The resulting increase in both pore size and surface hydrophilicity promotes pore hydration and disrupts vapor barrier to render the channel conductive.

To further identify structural elements central to BK VSD-pore coupling, we performed unbiased dynamic community and coupling pathway analysis based on the covariance matrices derived from atomistic simulations (see Methods). The results, summarized in **Figure 6a**, reveal that S4-S6 forms a single community distinct from the rest of the VSD (and S0 helix), where the motions of

Residues	Current work ^a	Carrasquel-Ursulaez (2022) ⁽³⁹⁾	Ma (2006) ⁽⁶³⁾	Diaz (1998) ⁽⁶⁴⁾
D153 (S2)	0.08 ± 0.04	-0.03	0.26	
R167 (S2)	0.002 ± 0.03	-0.02	0.15	
D186 (S3)	-0.01 ± 0.03	0.0	0.22	
R207 (S4)	0.04 ± 0.01	-0.01	-0.03	0.275
R210 (S4)	0.25 ± 0.02	0.32 to 0.35 ^b	-0.01	0.325
R213 (S4)	0.19 ± 0.01	0.23 to 0.31 ^b	0.34	0.3
E219 (S4)	-0.09 ± 0.02	0.03	0.02	

^a Statistical error was determined by block averaging.

^b Results from reversed charge mutations are excluded here.

Table 1

Residue contributions to the gating charge per VSD.

residues are more strongly coupled to each other than to the rest of the channel. Indeed, the optimal and suboptimal pathways of allosteric coupling between S4 R213 (a major voltage sensing charge) and S6 E321 (a key position of pore dilation and hydrophilicity increase) pass exclusively through S5 residues (**Figure 6b**). The central role of S5 in mediating VSD-pore coupling is further supported by analyzing the information flow betweenness (84), which provides a global measure how conformational perturbation flows through the network from the “source” (e.g., R213) to the “sink” (e.g., E321). The analysis reveals that S5 residues have the largest contributions to the informational flow besides neighboring residues on S4 or S6 (**Figure 6c**). Other residues on S1 or S3 also contribute significantly to the flow, albeit at much lower levels compared to S5. Interestingly, analysis of the simulation trajectory of Ca^{2+} fully open state (Table S1, *sim 7*) reveals highly similar patterns in the dynamic coupling communities, coupling pathways as well as informational flow (Figure S7), despite substantial differences in the VSD conformation and S4-S5-S6 packing (Figure S8). The implication is that Ca^{2+} - and voltage activation pathways likely have significant overlaps, which is consistent with experimental observations that BK channels can be synergistically or independently activated by membrane depolarization and Ca^{2+} binding. Taken together, the simulation and dynamic network analysis consistently point to a central role of the S4-S5-S6 interface in VSD-pore coupling of BK channels.

Concluding Discussion

Long atomistic simulations in explicit water and membrane performed using special purposed supercomputer Anton 2 have allowed direct observation of voltage-induced activation of the Core-MT BK channels for the first time. These simulations provide crucial new insights into how BK VSDs may sense membrane voltage and how the VSD movements may drive the pore opening and activate channel conductance. In particular, the simulations support that the S4 helix remains the major voltage sensing element and that it undergoes modest movements of $\sim 3 \text{ \AA}$ along both the membrane normal and lateral directions. The ability of BK channels to utilize relatively small VSD movements to sense membrane voltage and drive channel activation is likely attributed to the protein's ability to modify the local electric field, which leads to large electrostatic potential gradient in both vertical and lateral directions. Further analyses reveal that R210 and R213 are the major voltage sensing residues in the wild-type Core-MT BK channel, contributing $\sim 97\%$ to the total gating charge of $\sim 0.45 e$ per VSD. These core features of BK voltage sensing from simulation are largely consistent with a range of existing functional, biophysical and structural studies (38, 39, 41, 66), even though structural studies mutant BK channels with constitutively activated VSDs reveal smaller movements of R210 and R213 guanidinium moieties and minimal vertical movement of S4. The later observations may be a consequence of the complete absence of membrane voltage during structural determination. Importantly, control simulations show that the activated VSD state remains stable at 300 mV, suggesting that the observed vertical S4 movement is not an artifact of high voltage of 750 mV.

Another distinct feature derived from atomistic simulation is that the tightly packed S4-S5-S6 interface in BK channels plays a central role in the VSD-pore coupling, which is further supported by dynamic community, coupling pathway, and informational flow analyses. The modest movements in S4 drive a series of concerted conformational changes in S5 and eventually in S6, with the S4-S5 linker not playing a major role as observed in the canonical Kv channels (55–57). This is supported by the findings that mutations in the S4-S5 linker (N225-K228) do not significantly affect the coupling between VSD and the pore (40). Arguably, the distinct features of voltage-sensing and pore-sensor coupling observed in BK channels are intimately related to the novel non-domain-swapped TM topology. In particular, the VSD and pore domains are more tightly packed compared to the domain swapped configuration (**Figure 1**), which restricts the VSD motions and provides a direct pathway for their inter-talk. Interestingly, a similar non-canonical pathway for VSD-pore coupling, involving the S4/S1 and S1/S5 interfaces, has recently been proposed for the non-domain-swapped cardiac hERG potassium channel (85). We also note

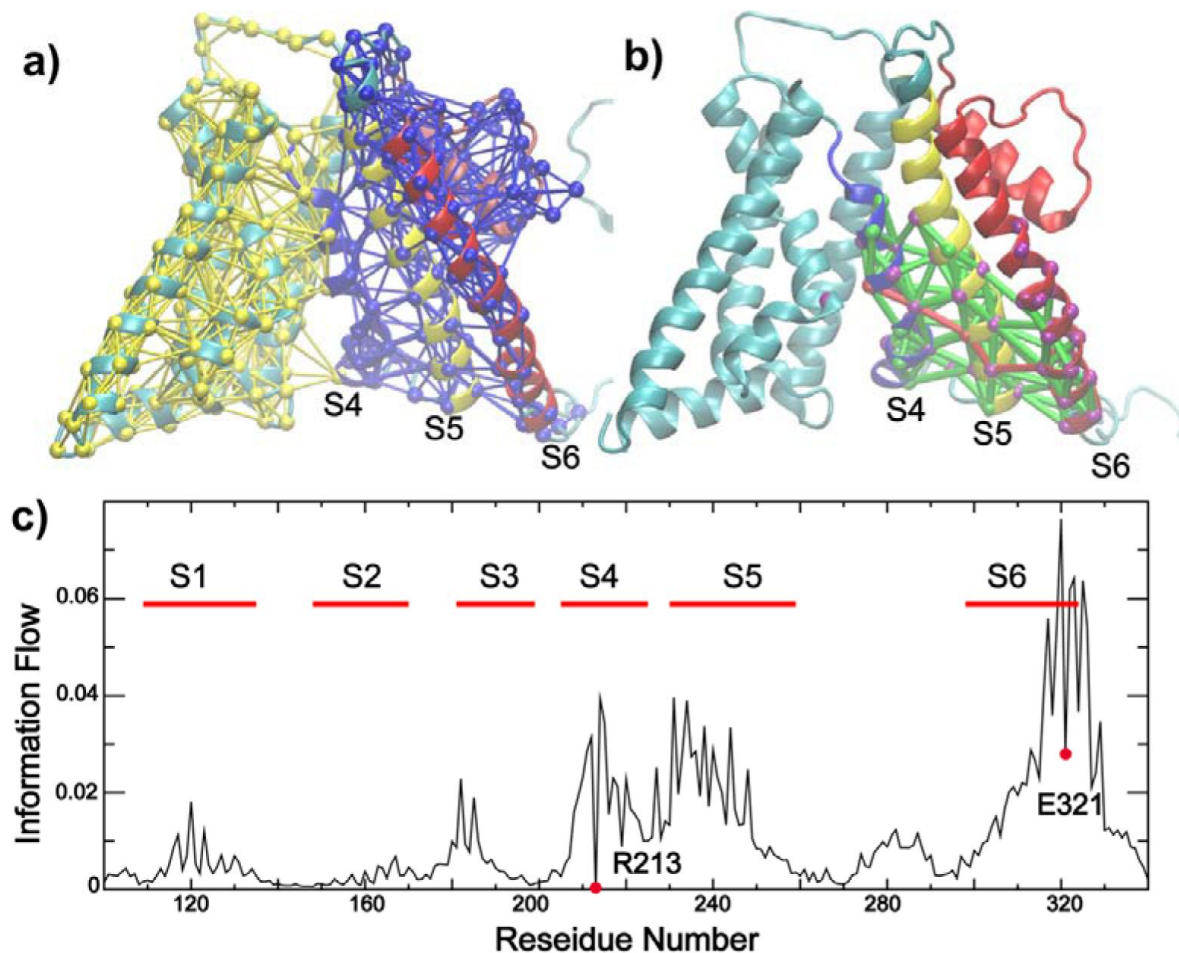


Figure 6

Dynamic community, coupling pathways, and information flow of VSD-pore coupling in BK.

a) Dynamic community analysis showing that TM S4-6 are clustered into single tightly coupled community (blue network). The nodes (residues) and edges (contacts) are colored based on the community number. **b)** Optimal and suboptimal pathways of dynamic coupling between R213 (VSD S4) and E321 (pore-lining S6). All paths are colored green except for the optimal path, which is colored red. Nodes with information flow value > 0.02 is highlighted in purple; **c)** Information flow profile of the Core-MT BK channel with R213 as the source and E321 as the sink node (labeled by red circle), respectively. All dynamic coupling analysis was derived from last 500 ns of *sim 1* (closed state at 0 mV; see Table S1).

that a similar non-canonical VSD-pore coupling involving S4-S5 interactions between neighboring subunits has been suggested to complement the canonical coupling mode even in domain-swapped Kv channels (57 [DOI](#), 86 [DOI](#), 87 [DOI](#)). An increasing number of ion channels have been discovered to adopt non-domain-swapped TM topology besides BK, including hERG (88 [DOI](#)), KvAP (89 [DOI](#)), HCN (90 [DOI](#)), Eag1 (91 [DOI](#)), and CNG channels (92 [DOI](#)). It is possible that the mechanistic features observed for BK channels may apply to voltage gating of the important emerging class of non-domain swapped ion channels in general.

Importantly, full-length BK channels can be independently activated by membrane potential and intracellular Ca^{2+} (22 [DOI](#), 93 [DOI](#), 94 [DOI](#)). The latter requires the C-terminal gating ring that is absent in the Core-MT construct studied in this work. Cryo-EM structures have revealed functional state-dependent interactions between the gating ring and VSD (28 [DOI](#)–30 [DOI](#), 38 [DOI](#)), strongly supporting that the calcium sensor and VSD are coupled. It is not clear how the VSD-gating ring coupling may affect the nature of VSD movements or how VSDs may drive the pore opening. Nonetheless, mechanistic details revealed from the voltage-gating of the Core-MT construct should provide a solid basis for future computational and experimental studies of BK activation and regulation.

Methods and Materials

Molecular modeling and atomistic simulations

The structure of the Core-MT human BK channel in the closed state was derived from the Cryo-EM structures of the *ac*BK channel in deactivated Ca^{2+} -free states (PDB 5tji (29 [DOI](#))) as previously described (32 [DOI](#), 95 [DOI](#)). The simulated construct was truncated at R342. The 11-residue C-terminal Kv mini-tails are not involved in voltage gating (68 [DOI](#), 69 [DOI](#)), and they were thus not included. The dynamic loop (C54-V91) and N-terminal tail (M1-N19) were not included either. Residues before and after the missing segments are capped with either an acetyl group (for N-terminus) or a N-methyl amide (for C-terminus). Standard protonation states under neutral pH were assigned for all titratable residues.

The initial structures was first inserted in model POPC lipid bilayers and then solvated in TIP3P water using the CHARMM-GUI web server (96 [DOI](#)). Even though polarizable water models are probably necessary to capture the precise energetics (and kinetics) of dewetting transitions within a protein cavity, it has also been shown that classical nonpolarizable water models such as TIP3P is sufficient to capture the spontaneous dewetting and rehydration of BK channels (32 [DOI](#)–34 [DOI](#), 36 [DOI](#)). In fact, a recent analysis using the Drude polarizable force field actually under-estimated the hydration free energy of deactivated BK pore (~ 0.5 kcal/mol) (35 [DOI](#)), which is very likely too small to sustain a dry and nonconductive pore. All systems were neutralized and 150 mM KCl added. The final simulation boxes contain about 593 lipid molecules (POPC) and $\sim 50,000$ water molecules and other solutes, with a total of $\sim 250,000$ atoms and dimensions of $\sim 160 \times 160 \times 110$ Å³. The CHARMM36m all-atom force field (97 [DOI](#)) and the CHARMM36 lipid force field (98 [DOI](#)) were used. All simulations were performed using Desmond (99 [DOI](#)) on Anton 2 (70 [DOI](#), 71 [DOI](#)) or CUDA-enabled versions of Gromacs 2020 (100 [DOI](#), 101 [DOI](#)) on GPU clusters. Electrostatic interactions were described by using the Particle Mesh Ewald (PME) algorithm (102 [DOI](#)) with a cutoff of 12 Å. Van der Waals interactions were cutoff at 12 Å with a smooth switching function starting at 10 Å. Covalent bonds to hydrogen atoms were constrained by the SHAKE algorithm (103 [DOI](#)), and the MD time step was set at 2 fs. The temperature was maintained at 298 K using the Nose-Hoover thermostat (104 [DOI](#), 105 [DOI](#)) (in Gromacs). The pressure was maintained semi-isotropically at 1 bar at membrane lateral directions using the Parrinello–Rahman barostat algorithm (106 [DOI](#)).

All systems were first minimized for 5000 steps using the steepest descent algorithm, followed by a series of equilibration steps where the positions of heavy atoms of the protein and lipid were harmonically restrained as prescribed by the CHARMM-GUI Membrane Builder (107 [DOI](#)). For Anton

simulation, additional 70 ns equilibration step with $0.1 \text{ kcal.mol}^{-1}.\text{\AA}^{-2}$ position restraints on all protein heavy atoms was performed before production runs. Note that the inner pore became dewetted during equilibration simulations. All production simulations were performed under NVT (constant particle number, volume and temperature) conditions at 298 K. The P-loop/filter (T273 to D292) and C-terminus were harmonically restrained with a small force constant of $0.1 \text{ kcal.mol}^{-1}.\text{\AA}^{-2}$ in all production simulations to prevent drift in all production simulations. Using Anton 2, a 2- μs control simulation was first performed without membrane voltage (*sim 1*, Table S1). Two independent 10- μs simulations were then performed at 750 mV to directly probe voltage-driven activation transitions (*sim 2*). The voltage was applied as a constant external electric field ($E = V/L_z$) imposed along the z dimension (83 [DOI](#)).

Steered molecular dynamics simulations

Steered molecular dynamics (SMD) simulations were performed to test the role of R210 and R213 in voltage sensing and channel activation. In these simulations, a moving reference point was used to simulate the effect of an external electric field by pulling the CZ atoms of R210 and R213 from their initial positions in the resting state to those observed in the activated state. As illustrated in Figure S4, the initial reference points of SMD were positioned at the location of each CZ atom of R210 and R213 in the resting/closed state. From 0 to 100 ns, these reference points were moved along the z-axis at constant speed until they reached the positions corresponding to the predicted activated VSD state at 100 ns. A harmonic positional restraint of $5 \text{ kcal/mol}.\text{\AA}^2$ was applied between each CZ atom to its respective reference point along z-axis, allowing the CZ atom to track the reference point's movement. After 100 ns, the reference points were held stationary at the positions corresponding to the activated VSD state, with the same harmonic restraints imposed to maintain the CZ atoms in their activated position. Four independent simulations (*sim 8*) were performed, and pore opening was observed in two of the four replicates (replicas 2 and 4). In the other two replicates, the S5-S6 packing was broken during the first 100 ns, likely due to strong steering potentials imposed, and the simulations were terminated.

Free energy analysis of gating charges

The total gating charge ΔQ is the sum of contributions of all charged residues in VSD and can be calculated as (82 [DOI](#), 83 [DOI](#)):

$$\Delta Q = \sum_i q_i [f_c(i) - f_o(i)], \quad (1)$$

where is q_i is the charge of a specific residue, and the dimensionless quantities $f_s(i)$ for a specific conformational state s , which can be open (o) or closed (c), represent the coupling of charge q_i to the transmembrane potential. The value of $f_s(i)$ is derived from the difference between charging free energies calculated at two different voltages V_1 and V_2 ,

$$f_s(i) = \frac{\Delta G_s(V_2, q_i) - \Delta G_s(V_1, q_i)}{q_i(V_2 - V_1)}, \quad (2)$$

where $\Delta G_s(V, q_i)$ is the free energy cost of increasing the charge of residue i from 0 to q_i in state s under membrane voltage V . $\Delta G_s(V, q_i)$ was calculated using thermodynamic integration (TI), with the charge gradually scaled to the final value in increments of 0.1, i.e., $\lambda = 0, 0.1, 0.2, \dots, 1.0$. In this work, we focused on 7 charged residues on VSD, namely, D153, R167, D186, R207, R210, R213 and E219, and performed four TI free energy calculations for each residue (*sim 3-6*, Table S1). The last snapshot of simulation 2b was used to represent the activated state of VSD. For each TI window, the system was first equilibrated 200 ps, and then simulated for 4 ns to calculate the mean force $\langle \delta H / \delta \lambda \rangle_\lambda$.

Structural and electrostatic potential analysis

The displacement of residue charged groups, as well as movement of helices (translocation, titling and rotation) is calculated using MDanalysis (108) together with in-house scripts. The TM helices are defined as: T109-S135 (S1), F148-A170 (S2), V181-L199 (S3), G205-N225 (S4), S230-S259 (S5), and A313-E324 (S6 after the glycine hinge). Residue contacts were identified using a minimal heavy atom distance cutoff of 5 Å. Pore water molecules were identified as those occupying the inner pore cavity below the selectivity filter, roughly from L312 to the plane defined by the center of mass of P320. Pore profiles are calculated using program HOLE(109). To examine how the protein modulates the local electric field, average electrostatic potential maps were then calculated from the first (resting) and last (activated) 250 ns of *sim 2b* using a combination of Python scripts and the VMD's PMEPot and VolMap plugins (110). The electrostatic potential is obtained by solving the Poisson equation $\nabla^2 \phi(r) = -4\pi \sum_i \rho_i(r)$ where the sum runs over all atoms, and $\rho_i(r)$ is the charge electric potential distribution contributed by atom i at position r approximated by a spherical Gaussian: $\rho_i(r) = q_i \left(\frac{\beta}{\pi} \right)^{3/2} e^{-\beta^2 (r-r_i)^2}$. An Ewald factor β of $0.25 \text{ \AA}^{-1}$ was used for the inverse width of the Gaussian and grid size was set to 1 Å. All molecular illustrations were prepared using VMD (111). External electric potential was added to the PMEPot grid file with in-house scripts (see supplementary files).

Dynamic community and coupling pathways

Dynamic network analysis was performed using the *Networkview* (112) plugin of VMD. To build the network, each amino acid was represented as a single node at the Ca position, and a contact (edge) was defined between two nodes if the minimal heavy-atom distance between residues was within a cutoff distance (5 Å) during at least 75% of the trajectory. The resulting contact matrix was weighted based on the covariance of dynamic fluctuation (C_{ij}) calculated from the same MD trajectory as $w_{ij} = -\log(|C_{ij}|)$. The length of a possible pathway D_{ij} between distant nodes i and j is defined as the sum of the edge weights between consecutive nodes along this path.

The shortest path, calculated using Floyd-Warshall algorithm (113), is considered the optimal pathway with the strongest dynamic coupling. Suboptimal paths are identified as alternative top-ranked paths with lengths that deviate by less than 50% from the optimal path. We further performed dynamic community analysis to identify groups of residues are more tightly coupled among themselves (114).

Information flow analysis

Information flow provides a global assessment of the contributions of all nodes to the dynamic coupling between selected “source” and “sink” nodes(115, 116). This analysis complements the dynamic pathway analysis to provide additional insights on how different residues may contribute to sensor-pore coupling. For this, a network similar to the one described above was first constructed. Pairwise mutual information was calculated between node i and j as follows: $M_{ij} = H_i + H_j - H_{ij}$. H_i is calculated as $-\frac{1}{N} \sum_{x=1}^N [-\ln \rho_i(x)]$, where $\rho_i(x)$ is the fluctuation density and x is the distance to the equilibrium position(116). Gaussian mixture model (GMM)(117) is used to estimate the density. The residue network is then defined as $A_{ij} = C_{ij} M_{ij}$, where C_{ij} is the contact map. To analysis the information flow from the source (S_0) to sink (S_f) nodes, the network Laplacian, is calculated as $L = D - A$, where D is diagonal degree matrix: $D_{ii} = \sum_j A_{ij}$. The information flow through a given node (residue) is defined as $f_i = \frac{1}{2} (\sum_j |P_i - P_j| A_{ij})$. The potentials P is given by $P = L^{-1} b$, where L^{-1} is the inverse reduced Laplacian, and b is the supply vector that corresponds to one unit of current entering at the source node that will exit at sink nodes. The magnitude of f_i thus quantifies the contribution of residue i to dynamic coupling between the source and sink nodes.

Data availability statement

All data needed to evaluate the conclusions in the paper are present in the paper and the Supplementary Materials. The PDB structures of Core-MT with resting and activated VSDs, as well as simulation input and analysis script, can be found on GitHub at: https://github.com/mdlab-um/Voltage_gating_Core-MT .

Acknowledgements

The authors thank Jianmin Cui and Guohui Zhang for critical discussions. This work is supported by NIH R35 GM144045 (Chen). Anton 2 computer time was provided by the Pittsburgh Supercomputing Center (PSC) through Grant R01 GM116961 from the National Institutes of Health. The Anton 2 machine at PSC was generously made available by D.E. Shaw Research.

Additional information

Author contributions

Chen and Jia conceived and initiated the study. Jia performed simulations and analysis. Jia and Chen wrote the manuscript.

Funding

National Institutes of Health

National Institutes of Health (R01 GM116961)

Additional files

Supplemental tables and figures 

References

1. Hodgkin A. L., Huxley A. F. (1952) **A quantitative description of membrane current and its application to conduction and excitation in nerve** *J. Physiol* **117**:500–544 [Google Scholar](#)
2. Bezanilla F. (2000) **The voltage sensor in voltage-dependent ion channels** *Physiol. Rev* **80**:555–592 [Google Scholar](#)
3. Sigworth F. J. (2003) **Structural biology: Life's transistors** *Nature* **423**:21–22 [Google Scholar](#)
4. MacKinnon R. (2003) **Potassium channels** *FEBS Lett.* **555**:62–65 [Google Scholar](#)
5. Swartz K. J. (2008) **Sensing voltage across lipid membranes** *Nature* **456**:891–897 [Google Scholar](#)
6. Brayden J. E., Nelson M. T. (1992) **Regulation of arterial tone by activation of calcium-dependent potassium channels** *Science* **256**:532–535 [Google Scholar](#)
7. Wellman G. C., Nelson M. T. (2003) **Signaling between SR and plasmalemma in smooth muscle: sparks and the activation of Ca²⁺-sensitive ion channels** *Cell Calcium* **34**:211–229 [Google Scholar](#)
8. Pluger S., et al. (2000) **Mice with disrupted BK channel beta1 subunit gene feature abnormal Ca²⁺ spark/STOC coupling and elevated blood pressure** *Circ Res* **87**:E53–60 [Google Scholar](#)
9. Nelson M. T., Quayle J. M. (1995) **Physiological roles and properties of potassium channels in arterial smooth muscle** *Am. J. Physiol* **268**:C799–822 [Google Scholar](#)
10. Tanaka Y., Aida M., Tanaka H., Shigenobu K., Toro L. (1998) **Involvement of maxi-KCa channel activation in atrial natriuretic peptide-induced vasorelaxation** *Naunyn Schmiedebergs Arch Pharmacol* **357**:705–708 [Google Scholar](#)
11. Perez G. J., Bonev A. D., Patlak J. B., Nelson M. T. (1999) **Functional coupling of ryanodine receptors to KCa channels in smooth muscle cells from rat cerebral arteries** *J Gen Physiol* **113**:229–238 [Google Scholar](#)
12. Sokolov S., Scheuer T., Catterall W. A. (2007) **Gating pore current in an inherited ion channelopathy** *Nature* **446**:76–78 [Google Scholar](#)
13. Ackerman M. J. (1998) **The long QT syndrome: Ion channel diseases of the heart** *Mayo Clin. Proc* **73**:250–269 [Google Scholar](#)
14. Kaczorowski G. J., Garcia M. L. (1999) **Pharmacology of voltage-gated and calcium-activated potassium channels** *Curr. Opin. Chem. Biol* **3**:448–458 [Google Scholar](#)
15. Lehmann-Horn F., Jurkat-Rott K. (1999) **Voltage-gated ion channels and hereditary disease** *Physiol. Rev* **79**:1317–1372 [Google Scholar](#)

16. Cannon S. C. (2006) **Pathomechanisms in Channelopathies of Skeletal Muscle and Brain** *Annu. Rev. Neurosci.* **29**:387–415 [Google Scholar](#)
17. Meldrum B. S., Rogawski M. A. (2007) **Molecular targets for antiepileptic drug development** *Neurotherapeutics* **4**:18–61 [Google Scholar](#)
18. Reid C. A., Berkovic S. F., Petrou S. (2009) **Mechanisms of human inherited epilepsies** *Prog. Neurobiol* **87**:41–57 [Google Scholar](#)
19. Eijkelkamp N., et al. (2012) **Neurological perspectives on voltage-gated sodium channels** *Brain* **135**:2585–2612 [Google Scholar](#)
20. Bennett D. L. H., Woods C. G. (2014) **Painful and painless channelopathies** *Lancet Neurol.* **13**:587–599 [Google Scholar](#)
21. Marty A. (1981) **Ca-dependent K channels with large unitary conductance in chromaffin cell membranes** *Nature* **291**:497–500 [Google Scholar](#)
22. Horrigan F. T., Aldrich R. W. (2002) **Coupling between Voltage Sensor Activation, Ca²⁺ Binding and Channel Opening in Large Conductance (BK) Potassium Channels** *The Journal of General Physiology* **120**:267–305 [Google Scholar](#)
23. Magleby K. L. (2003) **Gating mechanism of BK (Slo1) channels: So near, yet so far** *The Journal of General Physiology* **121**:81–96 [Google Scholar](#)
24. Salkoff L., Butler A., Ferreira G., Santi C., Wei A. (2006) **High-conductance potassium channels of the SLO family** *Nat Rev Neurosci* **7**:921–931 [Google Scholar](#)
25. Contreras G. F., et al. (2013) **A BK (Slo1) channel journey from molecule to physiology** *Channels* **7**:442–458 [Google Scholar](#)
26. Bentzen B. H., Olesen S.-P., Rønn L. C. B., Grunnet M. (2014) **BK channel activators and their therapeutic perspectives** *Front Physiol* **5** [Google Scholar](#)
27. Yang H., Zhang G., Cui J. (2015) **BK channels: multiple sensors, one activation gate** *Front Physiol* **6** [Google Scholar](#)
28. Hite R. K., Tao X., MacKinnon R. (2017) **Structural basis for gating the high-conductance Ca²⁺-activated K⁺ channel** *Nature* **541**:52–57 [Google Scholar](#)
29. Tao X., Hite R. K., MacKinnon R. (2017) **Cryo-EM structure of the open high-conductance Ca²⁺-activated K⁺ channel** *Nature* **541**:46–51 [Google Scholar](#)
30. Tao X., MacKinnon R. (2019) **Molecular structures of the human Slo1 K(+) channel in complex with beta4** *eLife* **8** [Google Scholar](#)
31. Long S. B., Campbell E. B., MacKinnon R. (2005) **Crystal structure of a mammalian voltage-dependent Shaker family K⁺ channel** *Science* **309**:897–903 [Google Scholar](#)
32. Jia Z., Yazdani M., Zhang G., Cui J., Chen J. (2018) **Hydrophobic gating in BK channels** *Nat. Commun* **9**:3408 [Google Scholar](#)
33. Gu R. X., de Groot B. L. (2023) **Central cavity dehydration as a gating mechanism of potassium channels** *Nat. Commun* **14** [Google Scholar](#)

34. Nordquist E. B., Jia Z., Chen J. (2023) **Inner pore hydration free energy controls the activation of big potassium channels** *Biophys. J* **122**:1158–1167 [Google Scholar](#)
35. Deng J., Cui Q. (2024) **Electronic Polarization Leads to a Drier Dewetted State for Hydrophobic Gating in the Big Potassium Channel** *The journal of physical chemistry letters* :7436–7441 [Google Scholar](#)
36. Coronel L., Muccio G. D., Rothberg B., Giacomello A., Carnevale V. (2024) **Lipid-mediated hydrophobic gating in the BK potassium channel** *ArXiv* [Google Scholar](#)
37. Zhou Y., Yang H., Cui J., Lingle C. J. (2017) **Threading the biophysics of mammalian Slo1 channels onto structures of an invertebrate Slo1 channel** *J. Gen. Physiol* **149**:985–1007 [Google Scholar](#)
38. Gustavo F. C., Rong S., Ramon L., Eduardo P. (2023) **Structural Basis of Voltage-Dependent Gating in BK Channels and Its Coupling to the Calcium Sensor** *bioRxiv* [Google Scholar](#)
39. Carrasquel-Ursulaez W., et al. (2022) **Mechanism of voltage sensing in Ca(2+)- and voltage-activated K(+) (BK) channels** *Proc. Natl. Acad. Sci. U. S. A* **119**:e2204620119 [Google Scholar](#)
40. Sun L., Horrigan F. T. (2022) **A gating lever and molecular logic gate that couple voltage and calcium sensor activation to opening in BK potassium channels** *Sci Adv* **8**:eabq5772 [Google Scholar](#)
41. Kallure G. S., Pal K., Zhou Y., Lingle C. J., Chowdhury S. (2023) **High-resolution structures illuminate key principles underlying voltage and LRRC26 regulation of Slo1 channels** *bioRxiv* [Google Scholar](#)
42. Larsson H. P., Baker O. S., Dhillon D. S., Isacoff E. Y. (1996) **Transmembrane movement of the Shaker K⁺ channel S4** *Neuron* **16**:387–397 [Google Scholar](#)
43. Yifrach O., MacKinnon R. (2002) **Energetics of pore opening in a voltage-gated K⁺ channel** *Cell* **111**:231–239 [Google Scholar](#)
44. Schmidt D., Jiang Q.-X., MacKinnon R. (2006) **Phospholipids and the origin of cationic gating charges in voltage sensors** *Nature* **444**:775–779 [Google Scholar](#)
45. Islas L. D., Sigworth F. J. (1999) **Voltage sensitivity and gating charge in Shaker and Shab family potassium channels** *J. Gen. Physiol* **114**:723–741 [Google Scholar](#)
46. Catterall W. A. (2010) **Ion channel voltage sensors: structure, function, and pathophysiology** *Neuron* **67**:915–928 [Google Scholar](#)
47. Catterall W. A., Zheng N. (2015) **Deciphering voltage-gated Na(+) and Ca(2+) channels by studying prokaryotic ancestors** *Trends Biochem. Sci* **40**:526–534 [Google Scholar](#)
48. Wu J., et al. (2016) **Structure of the voltage-gated calcium channel Ca(v)1.1 at 3.6 Å resolution** *Nature* **537**:191–196 [Google Scholar](#)
49. Lenaeus M. J., et al. (2017) **Structures of closed and open states of a voltage-gated sodium channel** *Proc. Natl. Acad. Sci. U. S. A* **114**:E3051–E3060 [Google Scholar](#)
50. Matthies D., et al. (2018) **Single-particle cryo-EM structure of a voltage-activated potassium channel in lipid nanodiscs** *eLife* **7** [Google Scholar](#)

51. Schoppa N. E., McCormack K., Tanouye M. A., Sigworth F. J. (1992) **The Size of Gating Charge in Wild-Type and Mutant Shaker Potassium Channels** *Science* **255**:1712–1715 [Google Scholar](#)
52. Aggarwal S. K., MacKinnon R. (1996) **Contribution of the S4 segment to gating charge in the Shaker K⁺ channel** *Neuron* **16**:1169–1177 [Google Scholar](#)
53. Long S. B., Tao X., Campbell E. B., MacKinnon R. (2007) **Atomic structure of a voltage-dependent K⁺ channel in a lipid membrane-like environment** *Nature* **450**:376–382 [Google Scholar](#)
54. Seoh S. A., Sigg D., Papazian D. M., Bezanilla F. (1996) **Voltage-sensing residues in the S2 and S4 segments of the Shaker K⁺ channel** *Neuron* **16**:1159–1167 [Google Scholar](#)
55. Fowler P. W., Sansom M. S. P. (2013) **The pore of voltage-gated potassium ion channels is strained when closed** *Nat. Commun* **4**:1872 [Google Scholar](#)
56. Jensen M. O., et al. (2012) **Mechanism of voltage gating in potassium channels** *Science* **336**:229–233 [Google Scholar](#)
57. Hou P., et al. (2020) **Two-stage electro-mechanical coupling of a KV channel in voltage-dependent activation** *Nat. Commun* **11**:676 [Google Scholar](#)
58. Horrigan F. T., Aldrich R. W. (1999) **Allosteric voltage gating of potassium channels II *Mslo* channel gating charge movement in the absence of Ca²⁺**. *J. Gen. Physiol.* **114**:305–336 [Google Scholar](#)
59. Lorenzo-Ceballos Y., Carrasquel-Ursulaez W., Castillo K., Alvarez O., Latorre R. (2019) **Calcium-driven regulation of voltage-sensing domains in BK channels** *eLife* **8**:e 44934 [Google Scholar](#)
60. Carrasquel-Ursulaez W., et al. (2015) **Hydrophobic interaction between contiguous residues in the S6 transmembrane segment acts as a stimuli integration node in the BK channel** *J. Gen. Physiol* **145**:61–74 [Google Scholar](#)
61. Contreras G. F., Neely A., Alvarez O., Gonzalez C., Latorre R. (2012) **Modulation of BK channel voltage gating by different auxiliary β subunits** *Proceedings of the National Academy of Sciences* **109**:18991–18996 [Google Scholar](#)
62. Stefani E., et al. (1997) **Voltage-controlled gating in a large conductance Ca²⁺-sensitive K⁺ channel (hslo)** *Proc. Natl. Acad. Sci. U. S. A* **94**:5427–5431 [Google Scholar](#)
63. Ma Z., Lou X. J., Horrigan F. T. (2006) **Role of charged residues in the S1-S4 voltage sensor of BK channels** *J. Gen. Physiol* **127**:309–328 [Google Scholar](#)
64. Diaz L., et al. (1998) **Role of the S4 segment in a voltage-dependent calcium-sensitive potassium (hSlo) channel** *J. Biol. Chem* **273**:32430–32436 [Google Scholar](#)
65. Pantazis A., Kohanteb A. P., Olcese R. (2010) **Relative motion of transmembrane segments S0 and S4 during voltage sensor activation in the human BK(Ca) channel** *J. Gen. Physiol* **136**:645–657 [Google Scholar](#)
66. Pantazis A., Olcese R. (2012) **Relative transmembrane segment rearrangements during BK channel activation resolved by structurally assigned fluorophore-quencher pairing** *J. Gen. Physiol* **140**:207–218 [Google Scholar](#)

67. Savalli N., Kondratiev A., Toro L., Olcese R. (2006) **Voltage-dependent conformational changes in human Ca(2+)- and voltage-activated K(+) channel, revealed by voltage-clamp fluorometry** *Proc. Natl. Acad. Sci. U. S. A* **103**:12619–12624 [Google Scholar](#)
68. Budelli G., Geng Y. Y., Butler A., Magleby K. L., Salkoff L. (2013) **Properties of Slo1 K+ channels with and without the gating ring** *Proc. Natl. Acad. Sci. U. S. A* **110**:16657–16662 [Google Scholar](#)
69. Zhang G., et al. (2017) **Deletion of cytosolic gating ring decreases gate and voltage sensor coupling in BK channels** *J. Gen. Physiol* **149**:373–387 [Google Scholar](#)
70. Shaw D. E., et al. (2008) **Anton, a special-purpose machine for molecular dynamics simulation** *Commun. ACM* **51**:91–97 [Google Scholar](#)
71. Shaw D. E., et al. (2014) **Anton 2: Raising the bar for performance and programmability in a special-purpose molecular dynamics supercomputer** *Int Conf High Perfor* :41–53 [Google Scholar](#)
72. Jiang T., Yu K., Hartzell H. C., Tajkhorshid E. (2017) **Lipids and ions traverse the membrane by the same physical pathway in the nhTMEM16 scramblase** *eLife* **6**:e28671 <https://doi.org/10.7554/eLife.28671> | [Google Scholar](#)
73. Furini S., Domene C. (2020) **Critical Assessment of Common Force Fields for Molecular Dynamics Simulations of Potassium Channels** *J. Chem. Theory Comput* **16**:7148–7159 [Google Scholar](#)
74. Ferguson W. B., McManus O. B., Magleby K. L. (1993) **Opening and closing transitions for BK channels often occur in two steps via sojourns through a brief lifetime subconductance state** *Biophys. J* **65**:702–714 [Google Scholar](#)
75. Sun X. P., Schlichter L. C., Stanley E. F. (1999) **Single-channel properties of BK-type calcium-activated potassium channels at a cholinergic presynaptic nerve terminal** *Journal of Physiology-London* **518**:639–651 [Google Scholar](#)
76. Gonzalez-Perez V., Zeng X. H., Henzler-Wildman K., Lingle C. J. (2012) **Stereospecific binding of a disordered peptide segment mediates BK channel inactivation** *Nature* **485**:133–136 [Google Scholar](#)
77. Takacs Z., Imredy J. P., Bingham J. P., Zhorov B. S., Moczydlowski E. G. (2014) **Interaction of the BKCa channel gating ring with dendrotoxins** *Channels* **8**:421–432 [Google Scholar](#)
78. Mironenko A., Zachariae U., de Groot B. L., Kopec W. (2021) **The Persistent Question of Potassium Channel Permeation Mechanisms** *J. Mol. Biol* **433** [Google Scholar](#)
79. Bernèche S., Roux B. (2001) **Energetics of ion conduction through the K+ channel** *Nature* **414**:73–77 [Google Scholar](#)
80. Roux B. (2005) **Ion conduction and selectivity in K+ channels** *Annu. Rev. Biophys. Biomol. Struct* **34**:153–171 [Google Scholar](#)
81. Starace D. M., Bezannilla F. (2004) **A proton pore in a potassium channel voltage sensor reveals a focused electric field** *Nature* **427**:548–553 [Google Scholar](#)
82. Khalili-Araghi F., et al. (2010) **Calculation of the gating charge for the Kv1.2 voltage-activated potassium channel** *Biophysical Journal* **98**:2189–2198 [Google Scholar](#)

83. Roux B. (2008) **The membrane potential and its representation by a constant electric field in computer simulations** *Biophys. J* **95**:4205–4216 [Google Scholar](#)
84. Botello-Smith W. M., Luo Y. (2019) **Robust Determination of Protein Allosteric Signaling Pathways** *J. Chem. Theory Comput* **15**:2116–2126 [Google Scholar](#)
85. Bassetto C. A. Z., Costa F., Guardiani C., Bezanilla F., Giacomello A. (2023) **Noncanonical electromechanical coupling paths in cardiac hERG potassium channel** *Nature Communications* **14**:1110 [Google Scholar](#)
86. Fernandez-Marino A. I., Harpole T. J., Oelstrom K., Delemotte L., Chanda B. (2018) **Gating interaction maps reveal a noncanonical electromechanical coupling mode in the Shaker K(+) channel** *Nat Struct Mol Biol* **25**:320–326 [Google Scholar](#)
87. Carvalho-de-Souza J. L., Bezanilla F. (2019) **Noncanonical mechanism of voltage sensor coupling to pore revealed by tandem dimers of Shaker** *Nat. Commun* **10**:3584 [Google Scholar](#)
88. Wang W., MacKinnon R. (2017) **Cryo-EM Structure of the Open Human Ether-a-go-go-Related K(+) Channel hERG** *Cell* **169**:422–430 [Google Scholar](#)
89. Tao X., MacKinnon R. (2019) **Cryo-EM structure of the KvAP channel reveals a non-domain-swapped voltage sensor topology** *eLife* **8** [Google Scholar](#)
90. Lee C. H., MacKinnon R. (2017) **Structures of the Human HCN1 Hyperpolarization-Activated Channel** *Cell* **168**:111–120 [Google Scholar](#)
91. Whicher J. R., MacKinnon R. (2016) **Structure of the voltage-gated K(+) channel Eag1 reveals an alternative voltage sensing mechanism** *Science* **353**:664–669 [Google Scholar](#)
92. Li M., et al. (2017) **Structure of a eukaryotic cyclic-nucleotide-gated channel** *Nature* **542**:60–65 [Google Scholar](#)
93. Shi J., Cui J. (2001) **Intracellular Mg²⁺ Enhances the Function of Bk-Type Ca²⁺-Activated K⁺ Channels** *The Journal of General Physiology* **118**:589–606 [Google Scholar](#)
94. Gessner G., et al. (2012) **Molecular mechanism of pharmacological activation of BK channels** *Proceedings of the National Academy of Sciences* **109**:3552–3557 [Google Scholar](#)
95. Yazdani M., et al. (2020) **Aromatic interactions with membrane modulate human BK channel activation** *eLife* **9**:e55571 <https://doi.org/10.7554/eLife.55571> | [Google Scholar](#)
96. Lee J., et al. (2016) **CHARMM-GUI input generator for NAMD, GROMACS, AMBER, OpenMM, and CHARMM/OpenMM simulations using the CHARMM36 additive force field** *J. Chem. Theory Comput.* **12**:405–413 [Google Scholar](#)
97. Huang J., et al. (2017) **CHARMM36m: an improved force field for folded and intrinsically disordered proteins** *Nat. Methods* **14**:71–73 [Google Scholar](#)
98. Klauda J. B., et al. (2010) **Update of the CHARMM all-atom additive force field for lipids: validation on six lipid types** *J Phys Chem B* **114**:7830–7843 [Google Scholar](#)
99. Bowers K. J., et al. (2006) , **paper presented at the Proceedings of the 2006 ACM/IEEE conference on Supercomputing, Tampa Florida** [Google Scholar](#)

100. Hess B., Kutzner C., van der Spoel D., Lindahl E. (2008) **GROMACS 4: algorithms for highly efficient, load-balanced, and scalable molecular simulation** *J. Chem. Theory Comput* **4**:435–447 [Google Scholar](#)
101. Abraham M. J., et al. (2015) **GROMACS: High performance molecular simulations through multi-level parallelism from laptops to supercomputers** *SoftwareX* **1**:1–2 [Google Scholar](#)
102. Darden T., York D., Pedersen L. (1993) **Particle mesh Ewald: An N-log (N) method for Ewald sums in large systems** *The Journal of Chemical Physics* **98** [Google Scholar](#)
103. Ryckaert J.-P., Ciccotti G., Berendsen H. J. C. (1977) **Numerical integration of the cartesian equations of motion of a system with constraints: molecular dynamics of n-alkanes** *J. Comput. Phys* **23**:327–341 [Google Scholar](#)
104. Nosé S. (1984) **A unified formulation of the constant temperature molecular dynamics methods** *The Journal of Chemical Physics* **81**:511–519 [Google Scholar](#)
105. Hoover W. G. (1985) **Canonical dynamics: Equilibrium phase-space distributions** *Phys. Rev. A* **31**:1695–1697 [Google Scholar](#)
106. Parrinello M., Rahman A. (1981) **Polymorphic transitions in single crystals: A new molecular dynamics method** *J. Appl. Phys* **52**:7182–7190 [Google Scholar](#)
107. Jo S., Kim T., Im W. (2007) **Automated Builder and Database of Protein/Membrane Complexes for Molecular Dynamics Simulations** *PLoS One* **2**:e880 [Google Scholar](#)
108. Michaud-Agrawal N., Denning E. J., Woolf T. B., Beckstein O. (2011) **MDAnalysis: A toolkit for the analysis of molecular dynamics simulations** *J. Comput. Chem* **32**:2319–2327 [Google Scholar](#)
109. Smart O. S., Neduvilil J. G., Wang X., Wallace B. A., Sansom M. S. P. (1996) **HOLE: A program for the analysis of the pore dimensions of ion channel structural models** *J. Mol. Graphics Modell* **14**:354 [Google Scholar](#)
110. Aksimentiev A., Schulten K. (2005) **Imaging alpha-hemolysin with molecular dynamics: ionic conductance, osmotic permeability, and the electrostatic potential map** *Biophys. J* **88**:3745–3761 [Google Scholar](#)
111. Humphrey W., Dalke A., Schulten K. (1996) **VMD: Visual molecular dynamics** *Journal of Molecular Graphics* **14**:33–38 [Google Scholar](#)
112. Eargle J., Luthey-Schulten Z. (2012) **NetworkView: 3D display and analysis of protein.RNA interaction networks** *Bioinformatics* **28**:3000–3001 [Google Scholar](#)
113. Floyd R. W. (1962) **Algorithm 97: Shortest path** *Commun. ACM* **5** [Google Scholar](#)
114. Girvan M., Newman M. E. (2002) **Community structure in social and biological networks** *Proc. Natl. Acad. Sci. U. S. A* **99**:7821–7826 [Google Scholar](#)
115. Kang P. W., et al. (2020) **Calmodulin acts as a state-dependent switch to control a cardiac potassium channel opening** *bioRxiv* [Google Scholar](#)
116. Westerlund A. M., Fleetwood O., Perez-Conesa S., Delemotte L. (2020) **Network analysis reveals how lipids and other cofactors influence membrane protein allostery** *J. Chem. Phys*

117. Dempster A. P., Laird N. M., Rubin D. B. (1977) **Maximum Likelihood from Incomplete Data Via the EM Algorithm** *Journal of the Royal Statistical Society: Series B (Methodological)* **39**:1–22 [Google Scholar](#)

Author information

Zhiguang Jia

Department of Chemistry, University of Massachusetts, Amherst, United States

Jianhan Chen

Department of Chemistry, University of Massachusetts, Amherst, United States

ORCID iD: [0000-0002-5281-1150](#)

For correspondence: jianhanc@umass.edu

Editors

Reviewing Editor

Merritt Maduke

Stanford University, Stanford, United States of America

Senior Editor

Merritt Maduke

Stanford University, Stanford, United States of America

Reviewer #1 (Public review):

Summary:

This study provides new insight into the non-canonical voltage-gating mechanism of BK channels through prolonged (10 us) MD simulations of the Slo1 transmembrane domain conformation and K⁺ conduction in response to high imposed voltages (300, 750 mV). The results support previous conclusions based on functional and structural data and MD simulations that the voltage-sensor domain (VSD) of Slo1 undergoes limited conformational changes compared to Kv channels, and predicts gating charge movement comparable in magnitude to experimental results. The gating charge calculations further indicate that R213 and R210 in S4 are the main contributors owing to their large side chain movements and the presence of a locally focused electric field, consistent with recent experimental and MD simulation results by Carrasquel-Ursulaez et al., 2022. Most interestingly, changes in pore conformation and K⁺ conduction driven by VSD activation are resolved, providing information regarding changes in VSD/pore interaction through S4/S5/S6 segments proposed to underly electromechanical coupling.

Strengths:

Include that the prolonged timescale and high voltage of the simulation allow apparent equilibration in the voltage-sensor domain (VSD) conformational changes and at least partial opening of the pore. The study extends the results of previous MD simulations of VSD activation by providing quantitative estimates of gating charge movement, showing how the

electric field distribution across the VSD is altered in resting and activated states, and testing the hypothesis that R213 and R210 are the primary gating charges by steered MD simulations. The ability to estimate gating charge contributions of individual residues in the WT channel is useful as a comparison to experimental studies based on mutagenesis which have yielded conflicting results that could reflect perturbations in structure. Use of dynamic community analysis to identify coupling pathways and information flow for VSD-pore (electromechanical) coupling as well as analysis of state-dependent S4/S5/S6 interactions that could mediate coupling provide useful predictions extending beyond what has been experimentally tested.

Weaknesses:

Weaknesses include that a truncated channel (lacking the C-terminal gating ring) was used for simulations, which is known to have reduced single channel conductance and electromechanical coupling compared to the full-length channel. In addition, as VSD activation in BK channels is much faster than opening, the timescale of simulations was likely insufficient to achieve a fully open state as supported by differences in the degree of pore expansion in replicate simulations, which are also smaller than observed in Ca-bound open structures of the full-length channel. Taken together, these limitations suggest that inferences regarding coupling pathways and interactions in the fully open voltage-activated channel may be only partially supported and therefore incomplete. That said, adequate discussion regarding these limitations are provided together with dynamic community analysis based on the Ca-bound open structure. The latter supports the main conclusions based on simulations, while providing an indication of potential interaction differences between simulated and fully open conformations. Another limitation is that while the simulations convincingly demonstrate voltage-dependent channel opening as evidenced by pore expansion and conduction of K⁺ and water through the pore, single channel conductance is underestimated by at least an order of magnitude, as in previous studies of other K⁺ channels. These quantitative discrepancies suggest that MD simulations may not yet be sufficiently advanced to provide insight into mechanisms underlying the extraordinarily large conductance of BK channels.

Comments on revisions:

My previous questions and concerns have been adequately addressed.

<https://doi.org/10.7554/eLife.105895.3.sa2>

Reviewer #2 (Public review):

Summary:

The manuscript by Jia and Chen addresses the structural basis of voltage-activation of BK channels using computational approaches. Although a number of experimental studies using gating current and patch-clamp recording have analyzed voltage-activation in terms of observed charge movements and the apparent energetic coupling between voltage-sensor movement and channel opening, the structural changes that underlie this phenomenon have been unclear. The present studies use a reduced molecular system comprising the transmembrane portion of the BK channel (i.e. the cytosolic domain was deleted), embedded in a POPC membrane, with either 0 or 750 mV applied across the membrane. This system enabled acquisition of long simulations of 10 microseconds, to permit tracking of conformational changes of the channel. The authors principal findings were that the side chains of R210 and R213 rapidly moved toward the extracellular side of the membrane (by 8 - 10 Å), with greater displacements than any of the other charged transmembrane residues. These movements appeared tightly coupled to movement of the pore-lining helix, pore

hydration, and ion permeation. The authors estimate that R210 and R213 contribute 0.25 and 0.19 elementary charges per residue to the gating current, which is roughly consistent with estimates based on electrophysiological measurements that used the full-length channel.

Strengths:

The methodologies used in this work are sound, and these studies certainly contribute to our understanding of voltage-gating of BK channels. An intriguing observation is the strongly coupled movement of the S4, S5, and S6 helices that appear to underlie voltage-dependent opening. Based on Fig 2a-d, the substantial movements of the R210 and R213 side chains occur nearly simultaneously to the S6 movement (between 4 - 5 usec of simulation time). This seems to provide support for a "helix-packing" mechanism of voltage gating in the so-called "non-domain-swapped" voltage-gated K channels.

Weaknesses:

The main limitation is that these studies used a truncated version of the BK channel, and there are likely to be differences in VSD-pore coupling in the context of the full-length channels that will not be resolved in the present work. Nonetheless, the authors provide a strong rationale for their use of the truncated channel, and the results presented will provide a good starting point for future computational studies of this channel.

<https://doi.org/10.7554/eLife.105895.3.sa1>

Author response:

The following is the authors' response to the previous reviews

Recommendations for the authors:

Reviewing Editor Comments:

The resubmitted version of the manuscript adequately addressed several initial comments made by reviewing editors, including a more detailed analysis of the results (such as those of bilayer thickness). This version was seen by 2 reviewers. Both reviewers recognize this work as being an important contribution to the field of BK and voltage-dependent ion channels in general. The long trajectories and the rigorous/novel analyses have revealed important insights into the mechanisms of voltage-sensing and electromechanical coupling in the context of a truncated variant of the BK channel. Many of these observations are consistent with structural and functional measurements of the channel, available thus far. The authors also identify a novel partially expanded state of the channel pore that is accessed after gating-charge displacement, which informs the sequence of structural events accompanying voltage-dependent opening of BK.

However, there are key concerns regarding the use of the truncated channel in the simulations. While many gating features of BK are preserved in the truncated variant, studies have suggested that opening of the channel pore to voltage-sensing domain rearrangement is impaired upon gating-ring deletion. So the inferences made here might only represent a partial view of the mechanism of electromechanical coupling.

It is also not entirely clear whether the partially expanded pore represents a functionally open, sub-conductance, or another closed state. Although the authors provide evidence that the inner pore is hydrated in this partially open state, in the absence of additional structural/functional restraints, a confident assignment of a functional state to this structure state is difficult. Functional measurements of the truncated channel seem to suggest that not only is their single channel conductance lower than full-length channels,

but they also appear to have a voltage-independent step that causes the gates to open. It is unclear whether it is this voltage-independent step that remains to be captured in these MD trajectories. A clean cut resolution of this conundrum might not be feasible at this time, but it could help present the various possibilities to the readers.

We appreciate the positive comments and agree that there will likely be important differences between the mechanistic details of voltage activation between the Core-MT and full-length constructs of BK channels. We also agree that the dilated pore observed in the simulation may not be the fully open state of Core-MT.

Nonetheless, the notion that the simulation may not have captured the full pore opening transition or the contribution of the CTD should not render the current work “incomplete”, because a complete understanding of BK activation would be an unrealistic goal beyond the scope of this work. We respectfully emphasize that the main insights of the current simulations are the mechanisms of voltage sensing (e.g., the nature of VSD movements, contributions of various charged residues, how small charge movements allow voltage sensing, etc.) as well as the role of the S4-S5-S6 interface in VSD-pore coupling. As noted by the Editor and reviewers, these insights represent important steps towards establishing a more complete understanding of BK activation.

Below are the specific comments of the two experts who have assessed the work and made specific suggestions to improve the manuscript.

Reviewer #1 (Recommendations for the authors):

(1) Although the successful simulation of V-dependent K⁺ conduction through the BK channel pore and analysis of associated state dependent VSD/pore interactions and coupling analysis is significant, there are two related questions that are relevant to the conclusions and of interest to the BK channel community which I think should be addressed or discussed.

One key feature of BK channels is their extraordinarily large conductance compared to other K⁺ selective channels. Do the simulations of K⁺ conductance provide any insight into this difference? Is the predicted conductance of BK larger than that of other K⁺ channels studied by similar methods? Is there any difference in the conductance mechanism (e.g., the hard and soft knock-on effects mentioned for BK)?

The molecular basis of the large conductance of BK channels is indeed an interesting and fundamental question. Unfortunately, this is beyond the scope of this work and the current simulation does not appear to provide any insight into the basis of large conductance. It is interesting to note, though, the conductance is apparently related to the level of pore dilation and the pore hydration level, as increasing hydration level from ~30 to ~40 waters in the pore increases the simulated conductance from ~1.5 to 6 pS (page 8). This is consistent with previous atomistic simulations (Gu and de Groot, Nature Communications 2023; ref. 33) showing that the pore hydration level is strongly correlated with observed conductance. As noted in the manuscript, the conductance mechanism through the filter appears highly similar to previous simulations of other K⁺ channels (Page 8). Given the limit conductance events observed in the current simulations, we will refrain from discussing possible basis of the large conductance in BK channels except commenting on the role of pore hydration (page 8; also see below in response to #5).

The pore in the MD simulations does not open as wide as the Ca-bound open structure, which (as the authors note) may mean that full opening requires longer than 10 us. I think that is highly likely given that the two 750 mV simulations yielded different degrees of opening and that in BK channels opening is generally much slower than charge

movement. Therefore, a question is - do any of the conclusions illustrated in Figures 6, S5, S6 differ if the Ca-bound structure is used as the open state? For example, I expect the interactions between S5 and S6 might at least change to some extent as S6 moves to its final position. In this case, would conclusions about which residues interact, and get stronger or weaker, be the same as in Figures S6 b,c? Providing a comparison may help indicate to what extent the conclusions are dependent on achieving a fully open conformation.

We appreciate the reviewer's suggestion and have further analyzed the information flow and coupling pathways using the simulation trajectory initiated from the Ca^{2+} -bound cryo-EM structure (sim 7, Table S1). The new results are shown in two new SI Figures S7 and S8, and new discussion has been added to pages 14-15. Comparing Figures 5 and S7, we find that dynamic community, coupling pathways, and information flow are highly similar between simulation of the open and closed states, even though there are significant differences in S5 contacts in the simulated open state vs Ca^{2+} -bound open state (Figure S8). Interestingly, there are significant differences in S4-S5 packing in the simulated and Ca^{2+} -bound open states (Figure S8 top panel), which likely reflect important difference in VSD/pore interactions during voltage vs Ca^{2+} activation.

(2) P4 Significance -"first, successful direct simulation of voltage-activation"

This statement may need rewording. As noted above Carrasquel-Ursulaez et al.,2022 (reference 39) simulated voltage sensor activation under comparable conditions to the current manuscript (3.9 us simulation at +400 mV), and made some similar conclusions regarding R210, R213 movement, and electric field focusing within the VSD. However, they did not report what happens to the pore or simulate K^+ movement. So do the authors here mean something like "first, successful direct simulation of voltage-dependent channel opening"?

We agree with the reviewer and have revised the statement to “ ... the first successful direct simulation of voltage-dependent activation of the big potassium (BK) channel, ..”

(3) P5 "We compare the membrane thickness at 300 and 750 mV and the results reveal no significant difference in the membrane thickness (Figure S2)"

The figure also shows membrane thickness at 0 mV and indicates it is 1.4 Angstroms less than that at 300 or 750 mV. Whether or not this difference is significant should be stated, as the question being addressed is whether the structure is perturbed owing to the use of non-physiological voltages (which would include both 300 and 750 mV).

We have revised the Figure S2 caption to clarify that one-way ANOVA suggest the difference is not significant.

(4) P7 "It should be noted that the full-length BK channel in the Ca^{2+} bound state has an even larger intracellular opening (Figure 2f, green trace), suggesting that additional dilation of the pore may

occur at longer timescales."

As noted above, I agree it is likely that additional pore dilation may occur at longer timescales. However, for completeness, I suppose an alternative hypothesis should be noted, e.g. "...suggesting that additional dilation of the pore may occur at longer timescales, or in response to Ca-binding to the full length channel."

This is a great suggestion. Revised as suggested.

(5) Since the authors raise the possibility that they are simulating a subconductance state, some more discussion on this point would be helpful, especially in relation to the hydrophobic gate concept. Although the Magleby group concluded that the cytoplasmic mouth of the (fully open) pore has little impact on single channel conductance, that doesn't rule out that it becomes limiting in a partially open conformation. The simulation in Figure 3A shows an initial hydration of the pore with ~15 waters with little conductance events, suggesting that hydration per se may not suffice to define a fully open state. Indeed, the authors indicate that the simulated open state (w/ ~30-40 waters) has 1/4th the simulated conductance of the open structure (w/ ~60 waters). So is it the degree of hydration that limits conductance? Or is there a threshold of hydration that permits conductance and then other factors that limit conductance until the pore widens further? Addressing these issues might also be relevant to understanding the extraordinarily large conductance of fully open BK compared to other K channels.

We agree with the reviewer's proposal that pore hydration seems to be a major factor that can affect conductance. This is also well in-line with the previous computational study by Gu and de Groot (2023). We have now added a brief discussion on page 8, stating "Besides the limitation of the current fixed charge force fields in quantitatively predicting channel conductance, we note that the molecular basis for the large conductance of BK channels is actually poorly understood (78). It is noteworthy that the pore hydration level appears to be an important factor in determining the apparent conductance in the simulation, which has also been proposed in a previous atomistic simulation study of the Aplysia BK channel (33)."

Minor points

(1) P5 "the fully relaxed pore profile (red trace in Figure S1d, top row) shows substantial differences compared to that of the Ca²⁺-free Cryo-EM structure of the full-length channel."

For clarity, I suggest indicating which is the Ca-free profile - "... Ca²⁺-free Cryo-EM structure of the full-length channel (black trace)."

We greatly appreciate the thoughtful suggestion. Revised as suggested.

(2) P8 "Consistent with previous simulations (78-80), the conductance follows a multi-ion mechanism, where there are at least two K⁺ ions inside the filter"

For clarity, I suggest indicating these are not previous simulations of BK channels (e.g., "previous simulations of other K⁺ channels ...").

Author response: Revised as suggested. Thank you.

(3) Figure 2, S1 - grey traces representing individual subunits are very difficult to see (especially if printed). I wonder if they should be made slightly darker. Similar traces in Figure 3 are easier to see.

The traces in Figure S1 are actually the same thickness in Figure 3 and they appear lighter due to the size of the figure. Figure 2 panels a-c have been updated to improve the resolution.

(4) Figure 2 - suggest labeling S6 as "S6 313-324" (similar to S4 notation) to indicate it is not the entire segment.

Figure 2 panel d) has been updated as suggested.

(5) Figure 2 legend - "Voltage activation of Core-MT BK channels. a-d)..."

It would be easier to find details corresponding to individual panels if they were referenced individually. For example:

"a-d) results from a 10- μ s simulation under 750 mV (sim2b in Table S1). Each data point represents the average of four subunits for a given snapshot (thin grey lines), and the colored thick lines plot the running average. a) z-displacement of key side chain charged groups from initial positions. The locations of charged groups were taken as those of guanidinium CZ atoms (for Arg) and sidechain carboxyl carbons (for Asp/Glu) b) z-displacement of centers-of-mass of VSD helices from initial positions, c) backbone RMSD of the pore-lining S6 (F307-L325) to the open state, and d) tilt angles of all TM helices. Only residues 313-324 of S6 were included in the tilt angle calculation, and the values in the open and closed Cryo-EM structures are marked using purple dashed lines. "

We appreciate the thoughtful suggestion and have revised the caption as suggested.

(6) Figure S1 - column labels a,b,c, and d should be referenced in the legend.

The references to column labels have been added to Figure S1 caption.

(7) References need to be double-checked for duplicates and formatting.

a) I noticed several duplicate references, but did not do a complete search: Budelli et al 2013 (#68, 100), Horrigan Aldrich 2002 (#22,97), Sun Horrigan 2022 (#40, 86), Jensen et al 2012 (#56,81).

b) Reference #38 is incorrectly cited with the first name spelled out and the last name abbreviated.

We appreciate the careful proofreading of the reviewer. The duplicated references were introduced by mistake due to the use of multiple reference libraries. We have gone through the manuscript and removed a total of 5 duplicated references.

Response to additional reviewer comments

My only new comment is that the numbering of residues in Fig. S8 does not match the standard convention for hSlo and needs to be doublechecked. For the residues I checked, the numbers appear to be shifted 3 compared hSlo (e.g. Y315, P317, E318, G324 should be Y318, P320, E321, G327).

We greatly appreciate the reviewer for catching the errors in residue labels. Figure S8 has now been updated to include correct residue labels. Thanks!

Reviewer #2 (Recommendations for the authors):

This manuscript has been through a previous level of review. The authors have provided their responses to the previous reviewers, which appear to be satisfactory, and I have no additional comments, beyond the caveats concerning interpretations based on the truncated channel, which are noted above.

We greatly appreciate the constructive comments and insightful advice. Please see above response to the Reviewing Editor's comments for response and changes regarding the caveats concerning interpretations of the current simulations.

<https://doi.org/10.7554/eLife.105895.3.sa0>